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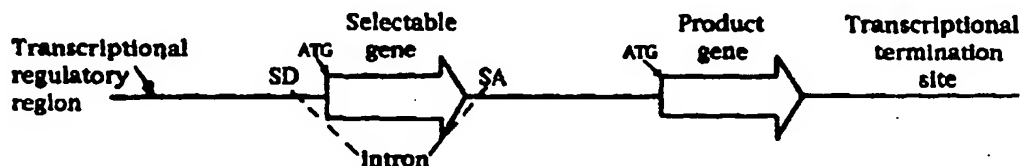
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(54) Title: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS



## (57) Abstract

A method for selecting recombinant host cells expressing high levels of a desired protein is described. This method utilizes eukaryotic host cells harboring a DNA construct comprising a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene. The selectable gene is positioned within an intron defined by a splice donor site and a splice acceptor site and the selectable gene and product gene are under the transcriptional control of a single transcriptional regulatory region. The splice donor site is generally an efficient splice donor site and thereby regulates expression of the product gene using the transcriptional regulatory region. The transfected cells are cultured so as to express the gene encoding the product in a selective medium comprising an amplifying agent for sufficient time to allow amplification to occur, whereupon either the desired product is recovered or cells having multiple copies of the product gene are identified.

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METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLSBACKGROUND OF THE INVENTIONField of the Invention

This invention relates to a method of selecting for high-expressing  
5 host cells, a method of producing a protein of interest in high yields and  
a method of producing eukaryotic cells having multiple copies of a sequence  
encoding a protein of interest.

Description of Background and Related Art

The discovery of methods for introducing DNA into living host cells  
10 in a functional form has provided the key to understanding many fundamental  
biological processes, and has made possible the production of important  
proteins and other molecules in commercially useful quantities.

Despite the general success of such gene transfer methods, several  
common problems exist that may limit the efficiency with which a gene  
15 encoding a desired protein can be introduced into and expressed in a host  
cell. One problem is knowing when the gene has been successfully  
transferred into recipient cells. A second problem is distinguishing  
between those cells that contain the gene and those that have survived the  
transfer procedures but do not contain the gene. A third problem is  
20 identifying and isolating those cells that contain the gene and that are  
expressing high levels of the protein encoded by the gene.

In general, the known methods for introducing genes into eukaryotic  
cells tend to be highly inefficient. Of the cells in a given culture, only  
a small proportion take up and express exogenously added DNA, and an even  
25 smaller proportion stably maintain that DNA.

Identification of those cells that have incorporated a product gene  
encoding a desired protein typically is achieved by introducing into the  
same cells another gene, commonly referred to as a selectable gene, that  
encodes a selectable marker. A selectable marker is a protein that is  
30 necessary for the growth or survival of a host cell under the particular  
culture conditions chosen, such as an enzyme that confers resistance to an  
antibiotic or other drug, or an enzyme that compensates for a metabolic or  
catabolic defect in the host cell. For example, selectable genes commonly  
used with eukaryotic cells include the genes for aminoglycoside  
35 phosphotransferase (APH), hygromycin phosphotransferase (hyg),  
dihydrofolate reductase (DHFR), thymidine kinase (tk), neomycin, puromycin,  
glutamine synthetase, and asparagine synthetase.

The method of identifying a host cell that has incorporated one gene  
on the basis of expression by the host cell of a second incorporated gene  
40 encoding a selectable marker is referred to as cotransfection (or  
cotransfection). In that method, a gene encoding a desired polypeptide and  
a selection gene typically are introduced into the host cell  
simultaneously, although they may be introduced sequentially. In the case  
of simultaneous cotransfection, the gene encoding the desired polypeptide



and the selectable gene may be present on a single DNA molecule or on separate DNA molecules prior to being introduced into the host cells. Wigler et al., Cell, 16:777 (1979). Cells that have incorporated the gene encoding the desired polypeptide then are identified or isolated by  
5 culturing the cells under conditions that preferentially allow for the growth or survival of those cells that synthesize the selectable marker encoded by the selectable gene.

The level of expression of a gene introduced into a eukaryotic host cell depends on multiple factors, including gene copy number, efficiency  
10 of transcription, messenger RNA (mRNA) processing, stability, and translation efficiency. Accordingly, high level expression of a desired polypeptide typically will involve optimizing one or more of those factors.

For example, the level of protein production may be increased by covalently joining the coding sequence of the gene to a "strong" promoter  
15 or enhancer that will give high levels of transcription. Promoters and enhancers are nucleotide sequences that interact specifically with proteins in a host cell that are involved in transcription. Kriegler, Meth. Enzymol., 185:512 (1990); Maniatis et al., Science, 236:1237 (1987). Promoters are located upstream of the coding sequence of a gene and  
20 facilitate transcription of the gene by RNA polymerase. Among the eukaryotic promoters that have been identified as strong promoters for high-level expression are the SV40 early promoter, adenovirus major late promoter, mouse metallothionein-I promoter, Rous sarcoma virus long terminal repeat, and human cytomegalovirus immediate early promoter (CMV).

Enhancers stimulate transcription from a linked promoter. Unlike  
25 promoters, enhancers are active when placed downstream from the transcription initiation site or at considerable distances from the promoter, although in practice enhancers may overlap physically and functionally with promoters. For example, all of the strong promoters  
30 listed above also contain strong enhancers. Bendig, Genetic Engineering, 7:91 (Academic Press, 1988).

The level of protein production also may be increased by increasing the gene copy number in the host cell. One method for obtaining high gene copy number is to directly introduce into the host cell multiple copies of  
35 the gene, for example, by using a large molar excess of the product gene relative to the selectable gene during cotransfection. Kaufman, Meth. Enzymol., 185:537 (1990). With this method, however, only a small proportion of the cotransfected cells will contain the product gene at high copy number. Furthermore, because no generally applicable, convenient  
40 method exists for distinguishing such cells from the majority of cells that contain fewer copies of the product gene, laborious and time-consuming screening methods typically are required to identify the desired high-copy number transfectants.

Another method for obtaining high gene copy number involves cloning  
45 the gene in a vector that is capable of replicating autonomously in the host cell. Examples of such vectors include mammalian expression vectors

derived from Epstein-Barr virus or bovine papilloma virus, and yeast 2-micron plasmid vectors. Stephens & Hentschel, Biochem. J., 248:1 (1987); Yates et al., Nature, 313:812 (1985); Beggs, Genetic Engineering, 2:175 (Academic Press, 1981).

5 Yet another method for obtaining high gene copy number involves gene amplification in the host cell. Gene amplification occurs naturally in eukaryotic cells at a relatively low frequency. Schimke, J. Biol. Chem., 263:5989 (1988). However, gene amplification also may be induced, or at least selected for, by exposing host cells to appropriate selective  
10 pressure. For example, in many cases it is possible to introduce a product gene together with an amplifiable gene into a host cell and subsequently select for amplification of the marker gene by exposing the cotransfected cells to sequentially increasing concentrations of a selective agent. Typically the product gene will be coamplified with the marker gene under  
15 such conditions.

The most widely used amplifiable gene for that purpose is a DHFR gene, which encodes a dihydrofolate reductase enzyme. The selection agent used in conjunction with a DHFR gene is methotrexate (Mtx). A host cell is cotransfected with a product gene encoding a desired protein and a DHFR  
20 gene, and transfectants are identified by first culturing the cells in culture medium that contains Mtx. A suitable host cell when a wild-type DHFR gene is used is the Chinese Hamster Ovary (CHO) cell line deficient in DHFR activity, prepared and propagated as described by Urlaub & Chasin, Proc. Nat. Acad. Sci. USA, 77:4216 (1980). The transfected cells then are  
25 exposed to successively higher amounts of Mtx. This leads to the synthesis of multiple copies of the DHFR gene, and concomitantly, multiple copies of the product gene. Schimke, J. Biol. Chem., 263:5989 (1988); Axel et al., U.S. Patent No. 4,399,216; Axel et al., U.S. Patent No. 4,634,665. Other references directed to co-transfection of a gene together with a genetic  
30 marker that allows for selection and subsequent amplification include Kaufman in Genetic Engineering, ed. J. Setlow (Plenum Press, New York), Vol. 9 (1987); Kaufman and Sharp, J. Mol. Biol., 159:601 (1982); Ringold et al., J. Mol. Appl. Genet., 1:165-175 (1981); Kaufman et al., Mol. Cell Biol., 5:1750-1759 (1985); Kaetzel and Nilson, J. Biol. Chem., 263:6244-  
35 6251 (1988); Hung et al., Proc. Natl. Acad. Sci. USA, 83:261-264 (1986); Kaufman et al., EMBO J., 6:87-93 (1987); Johnston and Kucey, Science, 242:1551-1554 (1988); Urlaub et al., Cell, 33:405-412 (1983).

To extend the DHFR amplification method to other cell types, a mutant DHFR gene that encodes a protein with reduced sensitivity to methotrexate  
40 may be used in conjunction with host cells that contain normal numbers of an endogenous wild-type DHFR gene. Simonsen and Levinson, Proc. Natl. Acad. Sci. USA, 80:2495 (1983); Wigler et al., Proc. Natl. Acad. Sci. USA, 77:3567-3570 (1980); Haber and Schimke, Somatic Cell Genetics, 8:499-508 (1982).

45 Alternatively, host cells may be co-transfected with the product gene, a DHFR gene, and a dominant selectable gene, such as a neo<sup>r</sup> gene. Kim

and Wold, Cell, 42:129 (1985); Capon et al., U.S. Pat. No. 4,965,199. Transfectants are identified by first culturing the cells in culture medium containing neomycin (or the related drug G418), and the transfectants so identified then are selected for amplification of the DHFR gene and the product gene by exposure to successively increasing amounts of Mtx.

As will be appreciated from this discussion, the selection of recombinant host cells that express high levels of a desired protein generally is a multi-step process. In the first step, initial transfectants are selected that have incorporated the product gene and the selectable gene. In subsequent steps, the initial transfectants are subject to further selection for high-level expression of the selectable gene and then random screening for high-level expression of the product gene. To identify cells expressing high levels of the desired protein, typically one must screen large numbers of transfectants. The majority of transfectants produce less than maximal levels of the desired protein. Further, Mtx resistance in DHFR transformants is at least partially conferred by varying degrees of gene amplification. Schimke, Cell, 37:705-713 (1984). The inadequacies of co-expression of the non-selected gene have been reported by Wold et al., Proc. Natl. Acad. Sci. USA, 76:5684-5688 (1979). Instability of the amplified DNA is reported by Kaufman and Schimke, Mol. Cell Biol., 1:1069-1076 (1981); Haber and Schimke, Cell, 26:355-362 (1981); and Fedespiel et al., J. Biol. Chem., 259:9127-9140 (1984).

Several methods have been described for directly selecting such recombinant host cells in a single step. One strategy involves co-transfecting host cells with a product gene and a DHFR gene, and selecting those cells that express high levels of DHFR by directly culturing in medium containing a high concentration of Mtx. Many of the cells selected in that manner also express the co-transfected product gene at high levels. Page and Sydenham, Bio/Technology, 9:64 (1991). This method for single-step selection suffers from certain drawbacks that limit its usefulness. High-expressing cells obtained by direct culturing in medium containing a high level of a selection agent may have poor growth and stability characteristics, thus limiting their usefulness for long-term production processes. Page and Snyderman, Bio/Technology, 9:64 (1991). Single-step selection for high-level resistance to Mtx may produce cells with an altered, Mtx-resistant DHFR enzyme, or cells that have altered Mtx transport properties, rather than cells containing amplified genes. Haber et al., J. Biol. Chem., 256:9501 (1981); Assaraf and Schimke, Proc. Natl. Acad. Sci. USA, 84:7154 (1987).

Another method involves the use of polycistronic mRNA expression vectors containing a product gene at the 5' end of the transcribed region and a selectable gene at the 3' end. Because translation of the selectable gene at the 3' end of the polycistronic mRNA is inefficient, such vectors exhibit preferential translation of the product gene and require high levels of polycistronic mRNA to survive selection. Kaufman, Meth.

Enzymol., 185:487 (1990); Kaufman, Meth. Enzymol., 185:537 (1990); Kaufman et al., EMBO J., 6:187 (1987). Accordingly, cells expressing high levels of the desired protein product may be obtained in a single step by culturing the initial transfectants in medium containing a selection agent  
5 appropriate for use with the particular selectable gene. However, the utility of these vectors is variable because of the unpredictable influence of the upstream product reading frame on selectable marker translation and because the upstream reading frame sometimes becomes deleted during methotrexate amplification (Kaufman et al., J. Mol. Biol., 159:601-621  
10 [1982]; Levinson, Methods in Enzymology, San Diego: Academic Press, Inc. [1990]). Later vectors incorporated an internal translation initiation site derived from members of the picornavirus family which is positioned between the product gene and the selectable gene (Pelletier et al., Nature, 334:320 [1988]; Jang et al., J. Virol., 63:1651 [1989]).

15 A third method for single-step selection involves use of a DNA construct with a selectable gene containing an intron within which is located a gene encoding the protein of interest. See U.S. Patent No. 5,043,270 and Abrams et al., J. Biol. Chem., 264(24): 14016-14021 (1989). In yet another single-step selection method, host cells are co-transfected  
20 with an intron-modified selectable gene and a gene encoding the protein of interest. See WO 92/17566, published October 15, 1992. The intron-modified gene is prepared by inserting into the transcribed region of a selectable gene an intron of such length that the intron is correctly spliced from the corresponding mRNA precursor at low efficiency, so that  
25 the amount of selectable marker produced from the intron-modified selectable gene is substantially less than that produced from the starting selectable gene. These vectors help to insure the integrity of the integrated DNA construct, but transcriptional linkage is not achieved as selectable gene and the protein gene are driven by separate promoters.

30 Other mammalian expression vectors that have single transcription units have been described. Retroviral vectors have been constructed (Cepko et al., Cell, 37:1053-1062 [1984]) in which a cDNA is inserted between the endogenous Moloney murine leukemia virus (M-MuLV) splice donor and splice acceptor sites which are followed by a neomycin resistance gene. This  
35 vector has been used to express a variety of gene products following retroviral infection of several cell types.

With the above drawbacks in mind, it is one object of the present invention to increase the level of homogeneity with regard to expression levels of stable clones transfected with a product gene of interest, by  
40 expressing a selectable marker (DHFR) and the protein of interest from a single promoter.

It is another object to provide a method for selecting stable, recombinant host cells that express high levels of a desired protein product, which method is rapid and convenient to perform, and reduces the  
45 numbers of transfected cells which need to be screened. Furthermore, it is

an object to allow high levels of single and two unit polypeptides to be rapidly generated from clones or pools of stable host cell transfectants.

It is an additional object to provide expression vectors which bias for active integration events (i.e. have an increased tendency to generate transformants wherein the DNA construct is inserted into a region of the genome of the host cell which results in high level expression of the product gene) and can accommodate a variety of product genes without the need for modification.

10

#### SUMMARY OF THE INVENTION

Accordingly, the present invention is directed to a DNA construct (DNA molecule) alternative terminology comprising a 5' transcriptional initiation site and a 3' transcriptional termination site, a selectable gene (preferably an amplifiable gene) and a product gene provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene positioned within an intron defined by a splice donor site and a splice acceptor site. The splice donor site preferably comprises an effective splice donor sequence as herein defined and thereby regulates expression of the product gene using the transcriptional regulatory region.

In another embodiment, the invention provides a method for producing a product of interest comprising culturing a eukaryotic cell which has been transfected with the DNA construct described above, so as to express the product gene and recovering the product.

In a further embodiment, the invention provides a method for producing eukaryotic cells having multiple copies of the product gene comprising transfecting eukaryotic cells with the DNA construct described above (where the selectable gene is an amplifiable gene), growing the cells in a selective medium comprising an amplifying agent for a sufficient time for amplification to occur, and selecting cells having multiple copies of the product gene. Preferably transfection of the cells is achieved using electroporation.

After transfection of the host cells, most of the transfectants fail to exhibit the selectable phenotype characteristic of the protein encoded by the selectable gene, but surprisingly a small proportion of the transfectants do exhibit the selectable phenotype, and among those transfectants, the majority are found to express high levels of the desired product encoded by the product gene. Thus, the invention provides an improved method for the selection of recombinant host cells expressing high levels of a desired product, which method is useful with a wide variety of eukaryotic host cells and avoids the problems inherent in existing cell selection technology.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1A-1D illustrate schematically various DNA constructs encompassed by the instant invention. The large arrows represent the selectable gene and the product gene, the V formed by the dashed lines shows the region of the precursor RNA internal to the 5' splice donor site (SD) and 3' splice acceptor site (SA) that is excised from vectors that contain a functional SD. The transcriptional regulatory region, selectable gene, product gene and transcriptional termination site are depicted in Figure 1A. Figure 1B depicts the DNA constructs of Example 1. The various splice donor sequences are depicted, i.e., wild type ras splice donor sequence (WT ras), mutant ras splice donor sequence (MUTANT ras) and non-functional splice donor sequence ( $\Delta$ GT). The probes used for Northern blot analysis in Example 1 are shown in Figure 1B. Figure 1C depicts the DNA constructs of Example 2 and Figure 1D depicts the DNA construct of Example 3 used for expression of anti-IgE  $V_H$ .

Figure 2 depicts schematically the control DNA construct used in Example 1.

Figures 3A-Q depict the nucleotide sequence (SEQ ID NO: 1) of the DHFR/intron-(WT ras SD)-tPA expression vector of Example 1.

Figure 4 is a bar graph which shows the number of colonies that form in selective medium after electroporation of linearized duplicate miniprep DNA's prepared in parallel from the three vectors shown in Figure 1B (i.e. with wild type ras splice donor sequence [WT ras], mutant ras splice donor sequence [MUTANT ras] and non-functional splice donor sequence [ $\Delta$ GT]) and from the control vector that has DHFR under control of SV40 promoter and tPA under control of CMV promoter (see Figure 2). Cells were selected in nucleoside free medium and counted with an automated colony counter.

Figures 5A-C are bar graphs depicting expression of tPA from stable pools and clones generated from the vectors shown in Figure 1B. In Figure 5A greater than 100 clones from each vector transfection were mixed, plated in 24 well plates, and assayed by tPA ELISA at "saturation". In Figure 5B, twenty clones chosen at random derived from each of the vectors were assayed by tPA ELISA at "saturation". In Figure 5C, the pools mentioned in Figure 5A (except the  $\Delta$ GT pool) were exposed to 200nM Mtx to select for DHFR amplification and then pooled and assayed for tPA expression.

Figures 6A-P depict the nucleotide sequence (SEQ ID NO: 2) of the DHFR/intron-(WT ras SD)-TNFr-IgG expression vector of Example 2.

Figures 7A-B are bar graphs depicting expression of TNFr-IgG using dicistronic or control vectors (see Example 2). Vectors containing TNFr-IgG (but otherwise identical to those described for tPA expression in Example 1) were constructed (see Figure 1C), introduced into dp12.CHO cells by electroporation, pooled, and assayed for product expression before (Figure 7A) and after (Figure 7B) being subjected to amplification in 200nM Mtx.

Figure 8 depicts schematically the DNA construct used for expression of the  $V_L$  of anti-IgE in Example 3.

Figures 9A-O depict the nucleotide sequence (SEQ ID NO: 3) of the anti-IgE V<sub>H</sub> expression vector of Example 3.

Figures 10A-Q depict the nucleotide sequence (SEQ ID NO: 4) of the anti-IgE V<sub>L</sub> expression vector of Example 3.

Figure 11 is a bar graph depicting anti-IgE expression in Example 3. Heavy (V<sub>H</sub>) and light (V<sub>L</sub>) chain expression vectors were constructed, co-electroporated into CHO cells, clones were selected and assayed for antibody expression. Additionally, pools were established and assessed with regard to expression before and after Mtx selection at 200nM and 1μM.

#### DESCRIPTION OF THE PREFERRED EMBODIMENTS

##### Definitions:

The "DNA construct" disclosed herein comprises a non-naturally occurring DNA molecule which can either be provided as an isolate or integrated in another DNA molecule e.g. in an expression vector or the chromosome of an eukaryotic host cell.

The term "selectable gene" as used herein refers to a DNA that encodes a selectable marker necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. Accordingly, a host cell that is transformed with a selectable gene will be capable of growth or survival under certain cell culture conditions wherein a non-transfected host cell is not capable of growth or survival. Typically, a selectable gene will confer resistance to a drug or compensate for a metabolic or catabolic defect in the host cell. Examples of selectable genes are provided in the following table. See also Kaufman, Methods in Enzymology, 185: 537-566 (1990), for a review of these.

**TABLE 1**  
**Selectable Genes and their Selection Agents**

Selection Agent	Selectable Gene
Methotrexate	Dihydrofolate reductase
Cadmium	Metallothionein
PALA	CAD
Xyl-A-or adenosine and 2'-deoxycorformycin	Adenosine deaminase
Adenine, azaserine, and coformycin	Adenylate deaminase
6-Azauridine, pyrazofuran	UMP Synthetase
Mycophenolic acid	IMP 5'-dehydrogenase

	Mycophenolic acid with limiting xanthine	Xanthine-guanine phosphoribosyltransferase
	Hypoxanthine, aminopterin, and thymidine (HAT)	Mutant HGPRTase or mutant thymidine kinase
5	5-Fluorodeoxyuridine	Thymidylate synthetase
	Multiple drugs e.g. adriamycin, vincristine or colchicine	P-glycoprotein 170
	Aphidicolin	Ribonucleotide reductase
10	Methionine sulfoximine	Glutamine synthetase
	$\beta$ -Aspartyl hydroxamate or Albizziin	Asparagine synthetase
	Canavanine	Arginosuccinate synthetase
	$\alpha$ -Difluoromethylornithine	Ornithine decarboxylase
15	Compactin	HMG-CoA reductase
	Tunicamycin	N-Acetylglucosaminyl transferase
	Borrelidin	Threonyl-tRNA synthetase
	Ouabain	Na <sup>+</sup> K <sup>+</sup> -ATPase

The preferred selectable gene is an amplifiable gene. As used herein, the term "amplifiable gene" refers to a gene which is amplified (i.e. additional copies of the gene are generated which survive in intrachromosomal or extrachromosomal form) under certain conditions. The amplifiable gene usually encodes an enzyme (i.e. an amplifiable marker) which is required for growth of eukaryotic cells under those conditions. For example, the gene may encode DHFR which is amplified when a host cell transformed therewith is grown in Mtx. According to Kaufman, the selectable genes in Table 1 above can also be considered amplifiable genes. An example of a selectable gene which is generally not considered to be an amplifiable gene is the neomycin resistance gene (Cepko et al., supra).

As used herein, "selective medium" refers to nutrient solution used for growing eukaryotic cells which have the selectable gene and therefore includes a "selection agent". Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are exemplary nutrient solutions. In addition, any of the media described in Ham and Wallace, Meth. Enz., 58:44 (1979), Barnes and Sato, Anal. Biochem., 102:255



(1980), U.S. Patent Nos. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Patent Re. 30,985; or U.S. Patent No. 5,122,469, the disclosures of all of which are incorporated herein by reference, may be used as culture media. Any of these media may be  
5 supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as Gentamycin™ drug), trace elements (defined as inorganic compounds usually  
10 present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The preferred nutrient solution comprises fetal bovine serum.

The term "selection agent" refers to a substance that interferes with  
15 the growth or survival of a host cell that is deficient in a particular selectable gene. Examples of selection agents are presented in Table 1 above. The selection agent preferably comprises an "amplifying agent" which is defined for purposes herein as an agent for amplifying copies of the amplifiable gene, such as Mtx if the amplifiable gene is DHFR. See Table  
20 1 for examples of amplifying agents.

As used herein, the term "transcriptional initiation site" refers to the nucleic acid in the DNA construct corresponding to the first nucleic acid incorporated into the primary transcript, i.e., the mRNA precursor, which site is generally provided at, or adjacent to, the 5' end of the DNA  
25 construct.

The term "transcriptional termination site" refers to a sequence of DNA, normally represented at the 3' end of the DNA construct, that causes RNA polymerase to terminate transcription.

As used herein, "transcriptional regulatory region" refers to a  
30 region of the DNA construct that regulates transcription of the selectable gene and the product gene. The transcriptional regulatory region normally refers to a promoter sequence (i.e. a region of DNA involved in binding of RNA polymerase to initiate transcription) which can be constitutive or inducible and, optionally, an enhancer (i.e. a cis-acting DNA element,  
35 usually from about 10-300 bp, that acts on a promoter to increase its transcription).

As used herein, "product gene" refers to DNA that encodes a desired protein or polypeptide product. Any product gene that is capable of expression in a host cell may be used, although the methods of the  
40 invention are particularly suited for obtaining high-level expression of a product gene that is not also a selectable or amplifiable gene. Accordingly, the protein or polypeptide encoded by a product gene typically will be one that is not necessary for the growth or survival of a host cell under the particular cell culture conditions chosen. For example, product  
45 genes suitably encode a peptide, or may encode a polypeptide sequence of

amino acids for which the chain length is sufficient to produce higher levels of tertiary and/or quaternary structure.

Examples of bacterial polypeptides or proteins include, e.g., alkaline phosphatase and  $\beta$ -lactamase. Examples of mammalian polypeptides or proteins include molecules such as renin; a growth hormone, including human growth hormone, and bovine growth hormone; growth hormone releasing factor; parathyroid hormone; thyroid stimulating hormone; lipoproteins; alpha-1-antitrypsin; insulin A-chain; insulin B-chain; proinsulin; follicle stimulating hormone; calcitonin; luteinizing hormone; glucagon; clotting factors such as factor VIIIC, factor IX, tissue factor, and von Willebrands factor; anti-clotting factors such as Protein C; atrial natriuretic factor; lung surfactant; a plasminogen activator, such as urokinase or human urine or tissue-type plasminogen activator (t-PA); bombesin; thrombin; hemopoietic growth factor; tumor necrosis factor-alpha and -beta; enkephalinase; RANTES (regulated on activation normally T-cell expressed and secreted); human macrophage inflammatory protein (MIP-1-alpha); a serum albumin such as human serum albumin; mullerian-inhibiting substance; relaxin A-chain; relaxin B-chain; prorelaxin; mouse gonadotropin-associated peptide; a microbial protein, such as beta-lactamase; DNase; inhibin; activin; vascular endothelial growth factor (VEGF); receptors for hormones or growth factors; integrin; protein A or D; rheumatoid factors; a neurotrophic factor such as bone-derived neurotrophic factor (BDNF), neurotrophin-3, -4, -5, or -6 (NT-3, NT-4, NT-5, or NT-6), or a nerve growth factor such as NGF- $\beta$ ; platelet-derived growth factor (PDGF); fibroblast growth factor such as aFGF and bFGF; epidermal growth factor (EGF); transforming growth factor (TGF) such as TGF-alpha and TGF-beta, including TGF- $\beta$ 1, TGF- $\beta$ 2, TGF- $\beta$ 3, TGF- $\beta$ 4, or TGF- $\beta$ 5; insulin-like growth factor-I and -II (IGF-I and IGF-II); des(1-3)-IGF-I (brain IGF-I), insulin-like growth factor binding proteins; CD proteins such as CD-3, CD-4, CD-8, and CD-19; erythropoietin; osteoinductive factors; immunotoxins; a bone morphogenetic protein (BMP); an interferon such as interferon-alpha, -beta, and -gamma; colony stimulating factors (CSFs), e.g., M-CSF, GM-CSF, and G-CSF; interleukins (ILs), e.g., IL-1 to IL-10; superoxide dismutase; T-cell receptors; surface membrane proteins; decay accelerating factor; viral antigen such as, for example, a portion of the AIDS envelope; transport proteins; homing receptors; addressins; regulatory proteins; antibodies; chimeric proteins such as immunoadhesins and fragments of any of the above-listed polypeptides.

The product gene preferably does not consist of an anti-sense sequence for inhibiting the expression of a gene present in the host. Preferred proteins herein are therapeutic proteins such as TGF- $\beta$ , TGF- $\alpha$ , PDGF, EGF, FGF, IGF-I, DNase, plasminogen activators such as t-PA, clotting factors such as tissue factor and factor VIII, hormones such as relaxin and insulin, cytokines such as IFN- $\gamma$ , chimeric proteins such as TNF receptor IgG immunoadhesin (TNFr-IgG) or antibodies such as anti-IgE.

The term "intron" as used herein refers to a nucleotide sequence present within the transcribed region of a gene or within a messenger RNA precursor, which nucleotide sequence is capable of being excised, or spliced, from the messenger RNA precursor by a host cell prior to translation. Introns suitable for use in the present invention are suitably prepared by any of several methods that are well known in the art, such as purification from a naturally occurring nucleic acid or *de novo* synthesis. The introns present in many naturally occurring eukaryotic genes have been identified and characterized. Mount, Nuc. Acids Res., 10:459 (1982). Artificial introns comprising functional splice sites also have been described. Winey et al., Mol. Cell Biol., 9:329 (1989); Gattermann et al., Mol. Cell Biol., 9:1526 (1989). Introns may be obtained from naturally occurring nucleic acids, for example, by digestion of a naturally occurring nucleic acid with a suitable restriction endonuclease, or by PCR cloning using primers complementary to sequences at the 5' and 3' ends of the intron. Alternatively, introns of defined sequence and length may be prepared synthetically using various methods in organic chemistry. Narang et al., Meth. Enzymol., 68:90 (1979); Caruthers et al., Meth. Enzymol., 154:287 (1985); Froehler et al., Nuc. Acids Res., 14:5399 (1986).

As used herein "splice donor site" or "SD" refers to the DNA sequence immediately surrounding the exon-intron boundary at the 5' end of the intron, where the "exon" comprises the nucleic acid 5' to the intron. Many splice donor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. An "efficient splice donor sequence" refers to a nucleic acid sequence encoding a splice donor site wherein the efficiency of splicing of messenger RNA precursors having the splice donor sequence is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. Examples of efficient splice donor sequences include the wild type (WT) ras splice donor sequence and the GAC:GTAAGT sequence of Example 3. Other efficient splice donor sequences can be readily selected using the techniques for measuring the efficiency of splicing disclosed herein.

The terms "PCR" and "polymerase chain reaction" as used herein refer to the *in vitro* amplification method described in US Patent No. 4,683,195 (issued July 28, 1987). In general, the PCR method involves repeated cycles of primer extension synthesis, using two DNA primers capable of hybridizing preferentially to a template nucleic acid comprising the nucleotide sequence to be amplified. The PCR method can be used to clone specific DNA sequences from total genomic DNA, cDNA transcribed from cellular RNA, viral or plasmid DNAs. Wang & Mark, in PCR Protocols, pp. 70-75 (Academic Press, 1990); Scharf, in PCR Protocols, pp. 84-98; Kawasaki & Wang, in PCR Technology, pp. 89-97 (Stockton Press, 1989). Reverse transcription-polymerase chain reaction (RT-PCR) can be used to analyze RNA samples containing mixtures of spliced and unspliced mRNA transcripts. Fluorescently tagged primers designed to span the intron are used to

amplify both spliced and unspliced targets. The resultant amplification products are then separated by gel electrophoresis and quantitated by measuring the fluorescent emission of the appropriate band(s). A comparison is made to determine the amount of spliced and unspliced transcripts present in the RNA sample.

One preferred splice donor sequence is a "consensus splice donor sequence". The nucleotide sequences surrounding intron splice sites, which sequences are evolutionarily highly conserved, are referred to as "consensus splice donor sequences". In the mRNAs of higher eukaryotes, the 5' splice site occurs within the consensus sequence AG:GUAAGU (wherein the colon denotes the site of cleavage and ligation). In the mRNAs of yeast, the 5' splice site is bounded by the consensus sequence :GUAUGU. Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986).

The expression "splice acceptor site" or "SA" refers to the sequence immediately surrounding the intron-exon boundary at the 3' end of the intron, where the "exon" comprises the nucleic acid 3' to the intron. Many splice acceptor sites have been characterized and Ohshima et al., J. Mol. Biol., 195:247-259 (1987) provides a review of these. The preferred splice acceptor site is an efficient splice acceptor site which refers to a nucleic acid sequence encoding a splice acceptor site wherein the efficiency of splicing of messenger RNA precursors having the splice acceptor site is between about 80 to 99% and preferably 90 to 95% as determined by quantitative PCR. The splice acceptor site may comprise a consensus sequence. In the mRNAs of higher eukaryotes, the 3' splice acceptor site occurs within the consensus sequence (U/C)<sub>1</sub>NCAG:G. In the mRNAs of yeast, the 3' acceptor splice site is bounded by the consensus sequence (C/U)AG:. Padgett, et al., *supra*.

As used herein "culturing for sufficient time to allow amplification to occur" refers to the act of physically culturing the eukaryotic host cells which have been transformed with the DNA construct in cell culture media containing the amplifying agent, until the copy number of the amplifiable gene (and preferably also the copy number of the product gene) in the host cells has increased relative to the transformed cells prior to this culturing.

The term "expression" as used herein refers to transcription or translation occurring within a host cell. The level of expression of a product gene in a host cell may be determined on the basis of either the amount of corresponding mRNA that is present in the cell or the amount of the protein encoded by the product gene that is produced by the cell. For example, mRNA transcribed from a product gene is desirably quantitated by northern hybridization. Sambrook, et al., Molecular Cloning: A Laboratory Manual, pp. 7.3-7.57 (Cold Spring Harbor Laboratory Press, 1989). Protein encoded by a product gene can be quantitated either by assaying for the biological activity of the protein or by employing assays that are independent of such activity, such as western blotting or radioimmunoassay using antibodies that are capable of reacting with the protein. Sambrook,

et al., Molecular Cloning: A Laboratory Manual, pp. 18.1-18.88 (Cold Spring Harbor Laboratory Press, 1989).

#### Modes for Carrying Out the Invention

Methods and compositions are provided for enhancing the stability and/or copy number of a transcribed sequence in order to allow for elevated levels of a RNA sequence of interest. In general, the methods of the present invention involve transfecting a eukaryotic host cell with an expression vector comprising both a product gene encoding a desired polypeptide and a selectable gene (preferably an amplifiable gene).

Selectable genes and product genes may be obtained from genomic DNA, cDNA transcribed from cellular RNA, or by in vitro synthesis. For example, libraries are screened with probes (such as antibodies or oligonucleotides of about 20-80 bases) designed to identify the selectable gene or the product gene (or the protein(s) encoded thereby). Screening the cDNA or genomic library with the selected probe may be conducted using standard procedures as described in chapters 10-12 of Sambrook et al., Molecular Cloning: A Laboratory Manual (New York: Cold Spring Harbor Laboratory Press, 1989). An alternative means to isolate the selectable gene or product gene is to use PCR methodology as described in section 14 of Sambrook et al., *supra*.

A preferred method of practicing this invention is to use carefully selected oligonucleotide sequences to screen cDNA libraries from various tissues known to contain the selectable gene or product gene. The oligonucleotide sequences selected as probes should be of sufficient length and sufficiently unambiguous that false positives are minimized.

The oligonucleotide generally is labeled such that it can be detected upon hybridization to DNA in the library being screened. The preferred method of labeling is to use <sup>32</sup>P- labeled ATP with polynucleotide kinase, as is well known in the art, to radiolabel the oligonucleotide. However, other methods may be used to label the oligonucleotide, including, but not limited to, biotinylation or enzyme labeling.

Sometimes, the DNA encoding the selectable gene and product gene is preceded by DNA encoding a signal sequence having a specific cleavage site at the N-terminus of the mature protein or polypeptide. In general, the signal sequence may be a component of the expression vector, or it may be a part of the selectable gene or product gene that is inserted into the expression vector. If a heterologous signal sequence is used, it preferably is one that is recognized and processed (i.e., cleaved by a signal peptidase) by the host cell. For yeast secretion the native signal sequence may be substituted by, e.g., the yeast invertase leader, alpha factor leader (including *Saccharomyces* and *Kluyveromyces*  $\alpha$ -factor leaders, the latter described in U.S. Pat. No. 5,010,182 issued 23 April 1991), or acid phosphatase leader, the *C. albicans* glucoamylase leader (EP 362,179 published 4 April 1990), or the signal described in WO 90/13646 published 15 November 1990. In mammalian cell expression the native signal sequence

of the protein of interest is satisfactory, although other mammalian signal sequences may be suitable, such as signal sequences from secreted polypeptides of the same or related species, as well as viral secretory leaders, for example, the herpes simplex gD signal. The DNA for such precursor region is ligated in reading frame to the selectable gene or product gene.

As shown in Figure 1A, the selectable gene is generally provided at the 5' end of the DNA construct and this selectable gene is followed by the product gene. Therefore, the full length (non-spliced) message will contain DHFR as the first open reading frame and will therefore generate DHFR protein to allow selection of stable transfectants. The full length message is not expected to generate appreciable amounts of the protein of interest as the second AUG in a dicistronic message is an inefficient initiator of translation in mammalian cells (Kozak, J. Cell Biol., 115: 887-903 [1991]).

The selectable gene is positioned within an intron. Introns are noncoding nucleotide sequences, normally present within many eukaryotic genes, which are removed from newly transcribed mRNA precursors in a multiple-step process collectively referred to as splicing.

A single mechanism is thought to be responsible for the splicing of mRNA precursors in mammalian, plant, and yeast cells. In general, the process of splicing requires that the 5' and 3' ends of the intron be correctly cleaved and the resulting ends of the mRNA be accurately joined, such that a mature mRNA having the proper reading frame for protein synthesis is produced. Analysis of a variety of naturally occurring and synthetically constructed mutant genes has shown that nucleotide changes at many of the positions within the consensus sequences at the 5' and 3' splice sites have the effect of reducing or abolishing the synthesis of mature mRNA. Sharp, Science, 235:766 (1987); Padgett, et al., Ann. Rev. Biochem., 55:1119 (1986); Green, Ann. Rev. Genet., 20:671 (1986). Mutational studies also have shown that RNA secondary structures involving splicing sites can affect the efficiency of splicing. Solnick, Cell, 43:667 (1985); Konarska, et al., Cell, 42:165 (1985).

The length of the intron may also affect the efficiency of splicing. By making deletion mutations of different sizes within the large intron of the rabbit beta-globin gene, Wieringa, et al. determined that the minimum intron length necessary for correct splicing is about 69 nucleotides. Cell, 37:915 (1984). Similar studies of the intron of the adenovirus E1A region have shown that an intron length of about 78 nucleotides allows correct splicing to occur, but at reduced efficiency. Increasing the length of the intron to 91 nucleotides restores normal splicing efficiency, whereas truncating the intron to 63 nucleotides abolishes correct splicing. Ulfendahl, et al., Nuc. Acids Res., 13:6299 (1985).

To be useful in the invention, the intron must have a length such that splicing of the intron from the mRNA is efficient. The preparation of introns of differing lengths is a routine matter, involving methods well known in the art, such as *de novo* synthesis or *in vitro* deletion

mutagenesis of an existing intron. Typically, the intron will have a length of at least about 150 nucleotides, since introns which are shorter than this tend to be spliced less efficiently. The upper limit for the length of the intron can be up to 30 kB or more. However, as a general  
5 proposition, the intron is generally less than about 10 kB in length.

The intron is modified to contain the selectable gene not normally present within the intron using any of the various known methods for modifying a nucleic acid *in vitro*. Typically, a selectable gene will be introduced into an intron by first cleaving the intron with a restriction  
10 endonuclease, and then covalently joining the resulting restriction fragments to the selectable gene in the correct orientation for host cell expression, for example by ligation with a DNA ligase enzyme.

The DNA construct is dicistronic, i.e. the selectable gene and product gene are both under the transcriptional control of a single  
15 transcriptional regulatory region. As mentioned above, the transcriptional regulatory region comprises a promoter. Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem., 255:2073 [1980]) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Reg., 7:149 [1968]; and Holland,  
20 Biochemistry, 17:4900 [1978]), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the  
25 additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and  
30 promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Expression control sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30  
35 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide.

Product gene transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as  
40 polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis-B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters,  
45 and from the promoter normally associated with the product gene, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication. Fiers et al., Nature, 273:113 (1978); Mulligan and Berg, Science, 209:1422-1427 (1980); Pavlakis et al., Proc. Natl. Acad. Sci. USA, 78:7398-7402 (1981). The immediate early promoter of the human cytomegalovirus (CMV) is conveniently obtained as a HindIII E restriction fragment. Greenaway et al., Gene, 18:355-360 (1982). A system for expressing DNA in mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature, 295:503-508 (1982) on expressing cDNA encoding immune interferon in monkey cells; , Reyes et al., Nature, 297:598-601 (1982) on expression of human  $\beta$ -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. USA, 79:5166-5170 (1982) on expression of the human interferon  $\beta$ 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. USA, 79:6777-6781 (1982) on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

Preferably the transcriptional regulatory region in higher eukaryotes comprises an enhancer sequence. Enhancers are relatively orientation and position independent having been found 5' (Lainins et al., Proc. Natl. Acad. Sci. USA, 78:993 [1981]) and 3' (Lusky et al., Mol. Cell Bio., 3:1108 [1983]) to the transcription unit, within an intron (Banerji et al., Cell, 33:729 [1983]) as well as within the coding sequence itself (Osborne et al., Mol. Cell Bio., 4:1293 [1984]). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin,  $\alpha$ -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer (CMV), the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature, 297:17-18 (1982) on enhancing elements for activation of eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to the product gene, but is preferably located at a site 5' from the promoter.

The DNA construct has a transcriptional initiation site following the transcriptional regulatory region and a transcriptional termination region following the product gene (see Figure 1A). These sequences are provided in the DNA construct using techniques which are well known in the art.

The DNA construct normally forms part of an expression vector which may have other components such as an origin of replication (i.e., a nucleic acid sequence that enables the vector to replicate in one or more selected host cells) and, if desired, one or more additional selectable gene(s). Construction of suitable vectors containing the desired coding and control sequences employs standard ligation techniques. Isolated plasmids or DNA



fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

Generally, in cloning vectors the origin of replication is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating sequences. Such sequences are well known. The 2 $\mu$  plasmid origin of replication is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e., they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

For analysis to confirm correct sequences in plasmids constructed, plasmids from the transformants are prepared, analyzed by restriction, and/or sequenced by the method of Messing et al., Nucleic Acids Res., 9:309 (1981) or by the method of Maxam et al., Methods in Enzymology, 65:499 (1980).

The expression vector having the DNA construct prepared as discussed above is transformed into a eukaryotic host cell. Suitable host cells for cloning or expressing the vectors herein are yeast or higher eukaryote cells.

Eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing the product gene. *Saccharomyces cerevisiae*, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are commonly available and useful herein, such as *S. pombe* [Beach and Nurse, Nature, 290:140 (1981)], *Kluyveromyces lactis* [Louvencourt et al., J. Bacteriol., 737 (1983)], *Yarrowia* [EP 402,226], *Pichia pastoris* [EP 183,070], *Trichoderma reesia* [EP 244,234], *Neurospora crassa* [Case et al., Proc. Natl. Acad. Sci. USA, 76:5259-5263 (1979)], and *Aspergillus* hosts such as *A. nidulans* [Ballance et al., Biochem. Biophys. Res. Commun., 112:284-289 (1983); Tilburn et al., Gene, 26:205-221 (1983); Yelton et al., Proc. Natl. Acad. Sci. USA, 81:1470-1474 (1984)] and *A. niger* [Kelly and Hynes, EMBO J., 4:475-479 (1985)].

Suitable host cells for the expression of the product gene are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from vertebrate or invertebrate culture. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as *Spodoptera frugiperda*

(caterpillar), *Aedes aegypti* (mosquito), *Aedes albopictus* (mosquito), *Drosophila melanogaster* (fruitfly), and *Bombyx mori* host cells have been identified. See, e.g., Luckow et al., Bio/Technology, 6:47-55 (1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., eds., Vol. 8 (Plenum Publishing, 1986), pp. 277-279; and Maeda et al., Nature, 315:592-594 (1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of *Autographa californica* NPV and the Bm-5 strain of *Bombyx mori* NPV, and such viruses may be used as the virus herein according to the present invention, particularly for transfection of *Spodoptera frugiperda* cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium *Agrobacterium tumefaciens*, which has been previously manipulated to contain the product gene. During incubation of the plant cell culture with *A. tumefaciens*, the product gene is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express the product gene. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen., 1:561 (1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. EP 321,196 published 21 June 1989.

However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years [Tissue Culture, Academic Press, Kruse and Patterson, editors (1973)]. Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen Virol., 36:59 [1977]); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. USA, 77:4216 [1980]); dpi2.CHO cells (EP 307,247 published 15 March 1989); mouse sertoli cells (TM4, Mather, Biol. Reprod., 23:243-251 [1980]); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HELA, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci., 383:44-68 [1982]); MRC 5 cells; FS4 cells; and a human hepatoma line (Hep G2).

Host cells are transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting transformants, or amplifying the genes encoding the desired sequences.

Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene, 23:315 (1983) and WO 89/05859 published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology, 52:456-457 (1978) may be used. General aspects of mammalian cell host system transformations have been described by Axel in U.S. 4,399,216 issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bact., 130:946 (1977) and Hsiao et al., Proc. Natl. Acad. Sci. (USA), 76:3829 (1979). However, other methods for introducing DNA into cells such as by nuclear injection or by protoplast fusion may also be used.

In the preferred embodiment the DNA is introduced into the host cells using electroporation. See Andreason, J. Tiss. Cult. Meth., 15:56-62 (1993), for a review of electroporation techniques useful for practicing the instantly claimed invention. It was discovered that electroporation techniques for introducing the DNA construct into the host cells were preferable over calcium phosphate precipitation techniques insofar as the latter could cause the DNA to break up and forming concatemers.

The mammalian host cells used to express the product gene herein may be cultured in a variety of media as discussed in the definitions section above. The media contains the selection agent used for selecting transformed host cells which have taken up the DNA construct (either as an intra- or extra-chromosomal element). To achieve selection of the transformed eukaryotic cells, the host cells may be grown in cell culture plates and individual colonies expressing the selectable gene (and thus the product gene) can be isolated and grown in growth medium until the nutrients are depleted. The host cells are then analyzed for transcription and/or transformation as discussed below. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, Northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. USA, 77:5201-5205 [1980]), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe, based on the sequences provided herein. Various labels may be employed, most commonly radioisotopes, particularly <sup>32</sup>P. However, other techniques may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescens, enzymes, or the like. Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the assay may be carried out where the duplex is bound to a surface, so that upon the

formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by immunological methods, such as immunohistochemical staining of tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels, fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use in the present invention is described by Hsu et al., Am. J. Clin. Path., 75:734-738 (1980).

In the preferred embodiment, the mRNA is analyzed by quantitative PCR (to determine the efficiency of splicing) and protein expression is measured using ELISA as described in Example 1 herein.

The product of interest preferably is recovered from the culture medium as a secreted polypeptide, although it also may be recovered from host cell lysates when directly expressed without a secretory signal. When the product gene is expressed in a recombinant cell other than one of human origin, the product of interest is completely free of proteins or polypeptides of human origin. However, it is necessary to purify the product of interest from recombinant cell proteins or polypeptides to obtain preparations that are substantially homogeneous as to the product of interest. As a first step, the culture medium or lysate is centrifuged to remove particulate cell debris. The product of interest thereafter is purified from contaminant soluble proteins and polypeptides, for example, by fractionation on immunoaffinity or ion-exchange columns; ethanol precipitation; reverse phase HPLC; chromatography on silica or on a cation exchange resin such as DEAE; chromatofocusing; SDS-PAGE; ammonium sulfate precipitation; gel electrophoresis using, for example, Sephadex G-75; chromatography on plasminogen columns to bind the product of interest and protein A Sepharose columns to remove contaminants such as IgG.

The following examples are offered by way of illustration only and are not intended to limit the invention in any manner. All patent and literature references cited herein are expressly incorporated by reference.

#### EXAMPLE 1

##### tPA production using the dicistronic expression vectors

It was sought to increase the level of homogeneity with regard to expression levels of stable clones by expressing a selectable marker (such as DHFR) and the protein of interest from a single promoter. These vectors divert most of the transcript to product expression while linking it at a fixed ratio to DHFR expression via differential splicing.

Vectors were constructed which were derived from the vector pRK (Suva et al., Science, 237:893-896 [1987]) which contains an intron between the cytomegalovirus immediate early promoter (CMV) and the cDNA that encodes

the polypeptide of interest. The intron of pRK is 139 nucleotides in length, has a splice donor site derived from cytomegalovirus immediate early gene (CMVIE), and a splice acceptor site from an IgG heavy chain variable region (V<sub>H</sub>) gene (Eaton et al., Biochem., 25:8343 [1986]).

5 DHFR/intron vectors were constructed by inserting an EcoRV linker into the BSTX1 site present in the intron of pRK7. An 830 base-pair fragment containing a mouse DHFR coding fragment was inserted to obtain DHFR intron expression vectors which differ only in the sequence that comprises the splice donor site. Those sequences were altered by  
10 overlapping PCR mutagenesis to obtain sequences that match splice donor sites found between exons 3 and 4 of normal and mutant Ras genes. PCR was also used to destroy the splice donor site.

A mouse DHFR cDNA fragment (Simonsen et al., Proc. Natl. Acad. Sci. USA, 80:2495-2499 [1983]) was inserted into the intron of this vector 59  
15 nucleotides downstream of the splice donor site. The splice donor site of this vector was altered by mutagenesis to change the ratio of spliced to non-spliced message in transfected cells. It has previously been shown that a single nucleotide change (G to A) converted a relatively efficient splice donor site found in the normal ras gene into an inefficient splice  
20 site (Cohen et al., Nature, 334:119-124 [1988]). This effect has been demonstrated in the context of the ras gene and confirmed when these sequences were transferred to human growth hormone constructs (Cohen et al., Cell, 58:461-472 [1989]). Additionally, a non functional 5' splice site (GT to CA) was constructed as a control ( $\Delta$ GT). A polylinker was  
25 inserted 35 nucleotides downstream of the 3' splice site to accept the cDNA of interest. A vector containing tPA (Pennica et al., Nature, 301:214-221 [1983]) was linearized downstream of the polyadenylation site before it was introduced into CHO cells (Potter et al., Proc. Natl. Acad. Sci. USA, 81:7161 [1984]).

30 Plasmid DNA's that contained DHFR/intron, tPA and (a) wild type ras (WT ras), i.e. Figure 3 (SEQ ID NO: 1), (b) mutant ras, or (c) non-functional splice donor site ( $\Delta$ GT) were introduced into CHO DHFR minus cells by electroporation. The intron vectors were each linearized downstream of the polyadenylation site by restriction endonuclease  
35 treatment. The control vector was linearized downstream of the second polyadenylation site. The DNA's were ethanol precipitated after phenol/chloroform extraction and were resuspended in 20 $\mu$ l 1/10 Tris EDTA. Then, 10 $\mu$ g of DNA was incubated with 10<sup>7</sup> CHO.dp12 cells (EP 307,247 published 15 March 1989) in 1 ml of PBS on ice for 10 min. before  
40 electroporation at 400 volts and 330 $\mu$ f using a BRL Cell Porator.

Cells were returned to ice for 10 min. before being plated into non-selective medium. After 24 hours cells were fed nucleoside-free medium to select for stable DHFR+ clones which were pooled. The pooled DHFR+ clones were lysed and mRNA's were prepared.

45 To prepare the mRNA, RNA was extracted from 5 x 10<sup>7</sup> cells which were grown from pools of more than 200 clones derived from the stable

transfection of the three vectors, the essential construction of which is shown in Figure 1B and from non-transfected CHO cells. RNA was purified over oligo-DT cellulase (Collaborative Biomedical Products). 10µg of mRNA was then subjected to Northern blotting which involved running the mRNA on  
5 a 1.2% agarose, 6.6% formaldehyde gel, and transferring it to a nylon filter (Stratagene Duralon-UV membrane), prehybridized, probed and washed according to the manufacturer's instructions.

The filter was probed sequentially using probes (shown in Figure 1B) that would detect (a) the full length message, (b) both full length and  
10 spliced message, or (c) beta actin. Probing with the long probe showed that the vector that contains the efficient splice donor site (i.e. WT ras) generates predominately a mRNA of the size predicted for the spliced product while the other two vectors gave rise primarily to a mRNA that corresponds in size to non-spliced message. The DHFR probe detected only  
15 full length message and demonstrated that the WT ras splice donor derived vector generates very little full length message with which to confer a DHFR positive phenotype.

Figure 4 shows the number of DHFR positive colonies obtained after duplicate electroporations with the three intron vectors described above  
20 and from a conventional vector that has a CMV promoter driving tPA and a SV40 promoter driving DHFR (see Figure 2). The increase in colony number parallels the increase in full length message that accumulates with the modification of the splice donor sites. The conventional vector efficiently generates colonies and does not vary significantly from the ΔGT  
25 construct.

The level of tPA expression was determined by seeding cells in 1 ml of F12:DMEM (50:50, with 5% FBS) in 24 well dishes to near confluency. Growth of the cells continued until the media was exhausted. Media was then assayed by ELISA for tPA production. Briefly, anti-tPA antibody was  
30 coated onto the wells of an ELISA microtiter plate, media samples were added to the wells followed by washing. Binding of the antigen (tPA) was then quantified using horse radish peroxidase (HRPO) labelled anti-tPA antibody.

Figure 5A depicts the titers of secreted tPA protein after pooling  
35 the clones of each group shown in Figure 4. While the number of colonies increased with a weakening of splice donor function, the inverse was seen with respect to tPA expression. The expression levels are consistent with the RNA products that are observed; as more of the dicistronic message is spliced an increased amount of message will contain tPA as the first open  
40 reading frame resulting in increased tPA expression. A mutation of GT to CA in the splice donor site results in an abundance of DHFR positive colonies which express undetectable levels of tPA, possibly resulting from inefficient utilization of the second AUG. Importantly, Figure 5A also shows that expression levels obtained from one of the dicistronic vectors  
45 (with WT ras SD) was about threefold higher than that obtained with the control vector containing a CMV promoter/enhancer driving tPA, SV40

promoter/enhancer controlling DHFR and SV40 polyadenylation signals controlling the expression of tPA and DHFR.

Additionally, the homogeneity of expression in the pools was investigated. Figure 5B shows that all 20 clones generated by the WT ras splice donor site derived dicistronic vectors express detectable levels of tPA while only 4 of 20 clones generated by the control vector express tPA. None of the clones transfected with the non-splicing ( $\Delta$ GT) vector expressed tPA levels detectable by ELISA. This finding is consistent with previous observations that relatively few clones generated by conventional vectors make useful levels of protein.

Expression of tPA was increased following methotrexate amplification of pools. Figure 5C shows that 2 of the dicistronic vector derived pools (i.e. with WT ras and MUTANT ras SD sites) increased in expression markedly (8.4 and 7.7 fold), while the pool generated by the conventional vector increased only slightly (2.8 fold) when each was subjected to 200 nM Mtx. An overall increase of 9 fold was obtained using the best dicistronic (WT ras SD) versus the conventional vector following amplification. Growth of the highest expressing amplified pool in nutrient rich production medium yielded titers of 4.2  $\mu$ g/ml tPA.

It was shown that manipulation of the splice donor sequence alters the ratio of spliced to full length message and the number of colonies that form in selective medium. It was also shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Surprisingly, it was possible to isolate high expressors which had the efficient WT ras splice donor site by selection for DHFR<sup>r</sup> cells despite the efficiency with which the DHFR gene was spliced from the RNA precursors formed in these cells.

#### EXAMPLE 2

##### TNFr-IgG production using the dicistronic expression vectors

To prove the general applicability of this approach, a second product was evaluated in the dicistronic vector system containing, as the DNA of interest, an immunoadhesin (TNFr-IgG) capable of binding tumor necrosis factor (TNF) (Ashkenazi et al., *Proc. Natl. Acad. Sci. USA*, 88:10535-10539 [1991]). The experiments described in Example 1 above were essentially repeated except that the product gene encoded the immunoadhesin TNFr-IgG. Plasmid DNA's that contained a TNFr-IgG cDNA and (a) WT ras, i.e. Figure 6 (SEQ ID NO: 2), (b) mutant ras or (c) nonfunctional splice donor site ( $\Delta$ GT) were introduced into the dp12.CHO cells as discussed for Example 1. See Figure 1C for an illustration of the DNA constructs.

It was discovered that the number of DHFR positive colonies generated by three of these vectors was similar to that seen with the tPA constructs. Expression of TNFr-IgG also paralleled that seen with the tPA constructs (Figure 7A). Amplification of pools from two of the constructs showed a marked increase in expression of immunoadhesin (9.6 and 6.8 fold) (Figure

7B). The best of these amplified pools expressed 9.5 µg/ml when grown in nutrient rich production medium.

Thus, it was again shown that dicistronic expression vectors generate clones that express high levels of recombinant proteins. Furthermore, contrary to expectations, it was discovered that isolation of high product expressing host DHFR<sup>+</sup> cells was possible using an efficient splice donor site (i.e. the WT ras splice donor site).

### EXAMPLE 3

#### Antibody production using a dicistronic expression vector

10 The usefulness of this system for antibody expression was evaluated by testing production of an antibody directed against IgE (Presta et al., Journal of Immunology, 151:2623-2632 [1993]). Further, the flexibility of the system with regard to transcription initiation was tested by replacing the CMV promoter/enhancer present in the previous vectors with the  
15 promoter/ enhancer derived from the early region of SV40 virus (Griffin, B., Structure and Genomic Organization of SV40 and Polyoma Virus, In J. Tooze [Ed] DNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York). The heavy chain of the antibody was inserted downstream of DHFR as described in the earlier tPA and TNFr-IgG constructs.  
20 Additionally, a new splice donor site sequence (GAC:GTAAGT) was engineered into the vector which matches the consensus splice donor site more closely than did the splice donor sites present in the vectors tested in Examples 1 and 2. The resultant expression vector is shown in Figures 1D and 9.

It was discovered that this vector produced fewer colonies than the  
25 vectors previously tested, and produced predominantly a spliced RNA product. A second vector was constructed to have the light chain of the antibody under control of the SV40 promoter/enhancer and poly-A and the hygromycin B resistance gene under control of the CMV promoter/enhancer and SV40 poly-A. These vectors were linearized at unique HpaI sites downstream  
30 of the poly-A signal, mixed at a ratio of light chain vector to heavy chain vector of 10:3 and electroporated into CHO cells using an optimized protocol (as discussed in Examples 1 and 2).

Figure 11 shows the levels of antibody expressed by clones and pools after selection in hygromycin B followed by selection for DHFR expression.  
35 All 20 of the clones analyzed expressed high levels of antibody when grown in rich medium and varied from one another by only a factor of four. A pool of antibody producing clones was generated and assayed shortly after it was established. That pool was grown continuously for 6 weeks without a significant decrease in productivity demonstrating that its stability was  
40 sufficient to generate gram quantities of protein from its large scale culture.

The pool was subjected to methotrexate amplification at 200nM and 1µM and achieved a greater than 2 fold increase in antibody titer. The 1µM Mtx resistant pool achieved a titer of 41 mg/L when grown under optimal  
45 conditions in suspension culture.



The structure of the expressed antibody was examined. Proteins expressed by the 200nM methotrexate resistant pool and by a well characterized expression clone generated by conventional vectors (Presta et al. [1993], supra) were metabolically labeled with S<sup>35</sup> cysteine and methionine. In particular, confluent 35mm plates of cells were metabolically labeled with 50μCi each S-35 methionine and S-35 cysteine (Amersham) in serum free cysteine and methionine free F12:DMEM. After one hour, nutrient rich production media was added and labeled proteins were allowed to "chase" into the medium for six more hours. Proteins were run on a 12% SDS/PAGE gel (NOVEX) non-reduced or following reduction with B-mercaptoethanol. Dried gels were exposed to film for 16 hours. CHO control cells were also labeled.

The majority of the antibody protein is secreted with a molecular weight of about 155 kilodaltons, consistent with a properly disulfide-linked antibody molecule with 2 light and 2 heavy chains. Upon reduction the molecular weight shifts to 2 approximately equally abundant proteins of 22.5 and 55 kilodaltons. The protein generated from the pool is indistinguishable from the antibody produced by the well characterized expression clone, with no apparent increase of free heavy or light chain expressed by the pool.

#### CONCLUSION

The efficient expression system described herein utilizes vectors consisting of promoter/enhancer elements followed by an intron containing the selectable marker coding sequence, followed by the cDNA of interest and a polyadenylation signal.

Several splice donor site sequences were tested for their effect on colony number and expression of the cDNA of interest. A non-functional splice donor site, splice donor sites found in an intron between exons 3 and 4 of mutant (mutant ras) and normal (WT ras) forms of the Harvey Ras gene and another efficient SD site (see Example 3) were used. The vectors were designed to direct expression of dicistronic primary transcripts. Within a transfected cell some of the transcripts remain full length while the remainder are spliced to excise the DHFR coding sequence. When the splice donor site is weakened or destroyed an increase in colony number is observed.

Expression levels show the inverse pattern, with the most efficient splice donor sites generating the highest levels of tPA, TNF $\alpha$  immunoadhesin or anti-IgE V<sub>H</sub>.

The homogeneity of expression of clones generated by the ras splice donor site intron DHFR vectors was compared to clones generated from a conventional vector with a separate promoter/enhancer and polyadenylation signal for each DHFR and tPA. The DHFR intron vector gives rise to colonies that are much more homogeneous with regard to expression than those generated by the conventional vector. Non-expressing clones derived from the conventional vector may be the result of breaks in the tPA or

TNFr-IgG domain of the plasmid during integration into the genome or the result of methylation of promoter elements (Busslinger et al., Cell, 34:197-206 [1983]; Watt et al., Genes and Development, 2:1136-1143 [1988]) driving tPA or TNFr-IgG expression. Promoter silencing by methylation or  
5 breaks in the DHFR-intron vectors would very likely render them incapable of conferring a DHFR positive phenotype.

It was found that pools generated by the DHFR-intron vectors could be amplified in methotrexate and would increase in expression by a factor of 8.4 (tPA), or 9.8 (TNFr-IgG). Pools from conventional vectors increased  
10 by only 2.8 and 3.0 fold for tPA and TNFr-IgG when amplified similarly. Amplified pools resulted in 9 fold higher tPA levels and 15 fold higher TNFr-IgG levels when compared to the conventional vector amplified pools.

Without being limited to any theory, the increase in expression of methotrexate resistant pools derived from the dicistronic vectors is likely  
15 due to the transcriptional linkage of DHFR and the product; when cells are selected for increased DHFR expression they consistently over-express product. Conventional approaches lack selectable marker and cDNA expression linkage and therefore methotrexate amplification often generates DHFR overexpression without the concomitant increase in product expression.

20 A further increase of 4 and 6.3 fold in expression were obtained when amplified tPA and TNFr-IgG pools were transferred from the media used for the selections and amplifications to a nutrient rich production medium.

In Example 3, the expression vector had a splice donor site that more closely matches the consensus splice donor sequence and had the heavy chain  
25 of a humanized anti-IgE antibody inserted downstream. This vector was linearized and co-electroporated with a second linearized vector that expresses the hygromycin resistance gene and the light chain of the antibody each under the control of its own promoter/enhancer and poly-A signals. An excess of light chain expression vector over the heavy chain  
30 dicistronic expression vector was used to bias in favor of light chain expression. Clones and a pool were generated after hygromycin B and DHFR selections. The clones were found to express relatively consistent, high levels of antibody, as did the pool. The 1 $\mu$ M pool achieved a titer of 41mg/L when grown under optimal conditions in suspension culture.

35 The anti-IgE antibody was assessed by metabolic labeling followed by SDS/PAGE under reducing and non reducing conditions and found to be indistinguishable from the protein expressed by a highly characterized clonal cell line. Of particular importance is the finding that no free light chain is observed in the pool relative to the clone.

40 A stable expression system for CHO cells has been developed that produces high levels of recombinant proteins rapidly and with less effort than that required by other expression systems. The vector system generates stable clones that express consistently high levels thereby reducing the number of clones that must be screened to obtain a highly  
45 productive clonal line. Alternatively, pools have been used to conveniently generate moderate to high levels of protein. This approach

may be particularly useful when a number of related proteins are to be expressed and compared.

Without being limited to this theory, it is possible the vectors that have very efficient splice donor sites generate very productive clones  
5 because so little transcript remains non spliced that only integration events that lead to the generation of high levels of RNA produce enough DHFR protein to give rise to colonies in selective medium. The high level of spliced message from such clones is then translated into abundant amounts of the protein of interest. Pools of clones made concurrently by  
10 introducing conventional vectors expressed lower levels of protein, and were unstable with regard to long term expression, and expression could not be appreciably increased when the cells were subjected to methotrexate amplification.

The system developed herein is versatile in that it allows high  
15 levels of single and multiple subunit polypeptides to be rapidly generated from clones or pools of stable transfectants. This expression system combines the advantages of transient expression systems (rapid and labor non intensive generation of research amounts of protein) with the concurrent development of highly productive stable production cell lines.

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

- 5 (i) APPLICANT: GENENTECH, INC.
- (ii) TITLE OF INVENTION: METHOD FOR SELECTING HIGH-EXPRESSING HOST CELLS
- 10 (iii) NUMBER OF SEQUENCES: 4
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: Genentech, Inc.
- (B) STREET: 460 Point San Bruno Blvd
- 15 (C) CITY: South San Francisco
- (D) STATE: California
- (E) COUNTRY: USA
- (F) ZIP: 94080
- (v) COMPUTER READABLE FORM:
- 20 (A) MEDIUM TYPE: 5.25 inch, 360 Kb floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: patin (Genentech)
- 25 (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER:
- (B) FILING DATE:
- (C) CLASSIFICATION:
- 30 (vii) PRIOR APPLICATION DATA:
- (A) APPLICATION NUMBER: 08/286740
- (B) FILING DATE: 05-AUG-1994
- (viii) ATTORNEY/AGENT INFORMATION:
- 35 (A) NAME: Lee, Wendy M.
- (B) REGISTRATION NUMBER: 00,000
- (C) REFERENCE/DOCKET NUMBER: 798PCT
- (ix) TELECOMMUNICATION INFORMATION:
- 40 (A) TELEPHONE: 415/225-1994
- (B) TELEFAX: 415/952-9881
- (C) TELEX: 910/371-7168
- (2) INFORMATION FOR SEQ ID NO:1:
- 45 (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 7360 bases
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- 50 (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

55 TTCGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50

TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGCG GTTACATAAC 100

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ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200

65 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250

ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300

5 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350

TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400

10 GGTTTTGGA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450

TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500

15 AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550

AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

20 TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650

25 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750

30 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800

GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850

35 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900

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55 GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTTGTGA CAAGGATCAT 1200

GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250

60 ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300

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5 AGTACAATCT GCTCTGATGC CGCATAGTTA AGCCAACTCC GCTATCGCTA 4950

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25 CATTCAAATA TGTATCCGCT CATGAGACAA TAACCCTGAT AAATGCTTCA 5300

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30 TTATTCCCTT TTTTGCGGCA TTTTGCCCTC CTGTTTTTGC TCACCCAGAA 5400

ACGCTGGTGA AAGTAAAAGA TGCTGAAGAT CAGTTGGGTG CACGAGTGGG 5450

35 TTACATCGAA CTGGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTCGCC 5500

CCGAAGAACG TTTTCCAATG ATGAGCACTT TTAAAGTTCT GCTATGTGGC 5550

GCGGTATTAT CCCGTGATGA CGCCGGGCAA GAGCAACTCG GTCGCCGCAT 5600

45 ACACTATTCT CAGAATGACT TGGTTGAGTA CTCACCAGTC ACAGAAAAGC 5650

ATCTTACGGA TGGCATGACA GTAAGAGAAT TATGCAGTGC TGCCATAACC 5700

50 ATGAGTGATA ACACTGCGGC CAACTTACTT CTGACAACGA TCGGAGGACC 5750

GAAGGAGCTA ACCGCTTTTT TGCACAACAT GGGGGATCAT GTAACGCGC 5800

TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACCAA CGACGAGCGT 5850

60 GACACCACGA TGCCAGCAGC AATGGCAACA ACGTTGCGCA AACTATTAAC 5900

TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTAATA GACTGGATGG 5950

65 AGGCGGATAA AGTTGCAGGA CCACTTCTGC GCTCGGCCCT TCCGGCTGGC 6000

TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT 6050  
CATTGCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT 6100  
5 ACACGACGGG GAGTCAGGCA ACTATGGATG AACGAAATAG ACAGATCGCT 6150  
GAGATAGGTG CCTCACTGAT TAAGCATTGG TAACTGTCAG ACCAAGTTTA 6200  
CTCATATATA CTTTAGATTG ATTTAAAACT TCATTTTAA TTTAAAAGGA 6250  
15 TCTAGGTGAA GATCCTTTTT GATAATCTCA TGACCAAAAT CCCTTAACGT 6300  
GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAAGGATC 6350  
20 TTCTTGAGAT CCTTTTTTTC TGC CGTAAT CTGCTGCTTG CAAACAAAAA 6400  
AACCACCGCT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT 6450  
CTTTTTCCGA AGGTAAGTGG CTTGAGCAGA GCGCAGATAC CAAATACTGT 6500  
30 CCTTCTAGTG TAGCCGTAGT TAGGCCACCA CTTCAAGAAC TCTGTAGCAC 6550  
CGCCTACATA CCTCGCTCTG CTAATCCTGT TACCAGTGGC TGCTGCCAGT 6600  
35 GCGGATAAGT CGTGTCTTAC CGGGTTGGAC TCAAGACGAT AGTTACCGGA 6650  
TAAGGCGCAG CGGTCGGGCT GAACGGGGGG TTCGTGCACA CAGCCCAGCT 6700  
TGGAGCGAAC GACCTACACC GAACTGAGAT ACCTACAGCG TGAGCATTGA 6750  
45 GAAAGCGCCA CGCTTCCCGA AGGGAGAAAG GCGGACAGGT ATCCGGTAAG 6800  
CGGCAGGGTC GGAACAGGAG AGCGCACGAG GGAGCTTCCA GGGGGAAACG 6850  
50 CCTGGTATCT TTATAGTCCT GTCGGGTTTC GCCACCTCTG ACTTGAGCGT 6900  
CGATTTTTGT GATGCTCGTC AGGGGGGCGG AGCCTATGGA AAAACGCCAG 6950  
CAACGCGGCC TTTTACGGT TCCTGGCCTT TTGCTGCGCT TTTGCTCACA 7000  
60 TGTTCTTTCC TGC GTTATCC CTGATTCTG TGGATAACCG TATTACCGCC 7050  
TTTGAGTGAG CTGATACCGC TCGCCGAGC CGAACGACCG AGCGCAGCGA 7100  
65 GTCAGTGAGC GAGGAAGCGG AAGAGCGCCC AATACGCAAA CCGCCTCTCC 7150

CCGCGCGTTG GCCGATTCAT TAATCCAGCT GGCACGACAG GTTCCCGAC 7200  
5 TGGAAAGCGG GCAGTGAGCG CAACGCAATT AATGTGAGTT ACCTCACTCA 7250  
TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG 7300  
10 GAATTGTGAG CGGATAACAA TTTCACACAG GAAACAGCTA TGACCATGAT 7350  
TACGAATTAA 7360

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## (2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:  
20 (A) LENGTH: 6889 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: linear

25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

TTGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50  
30 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCGC GTTACATAAC 100  
TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATG 150  
35 ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200  
40 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250  
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300  
45 AAATGGCCCG CCTGGCATTG TGCCCAAGTAC ATGACCTTAT GGGACTTTCC 350  
TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400  
50 GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450  
55 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500  
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
60 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600  
TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650  
65 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCCG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA 750  
5 GCGATAAGAG GATTTTATCC CCGCTGCCAT CATGGTTCGA CCATTGAACT 800  
GCATCGTCGC CGTGTCCCAA AATATGGGGA TTGGCAAGAA CGGAGACCTA 850  
10 CCCTGCCCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC 900  
AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAAA 950  
15 CCTGGTTCTC CATTCTGAG AAGAATCGAC CTTTAAAGGA CAGAATTAAT 1000  
ATAGTTCTCA GTAGAGAACT CAAAGAACCA CCACGAGGAG CTCATTTTCT 1050  
20 TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG 1100  
25 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTTTACCAG 1150  
GAAGCCATGA ATCAACCAGG CCACCTTAGA CTCTTGTGA CAAGGATCAT 1200  
30 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAAATTGAT TTGGGGAAAT 1250  
ATAAACCTCT CCCAGAATAC CCAGGCGTCC TCTCTGAGGT CCAGGAGGAA 1300  
35 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA 1350  
40 AGATGCTTTC AAGTTCTCTG CTCCCCTCCT AAAGCTATGC ATTTTATATA 1400  
GACCATGGGA CTTTTGCTGG CTTTAGACCC CCTTGGCTTC GTTAGAACGC 1450  
45 GGCTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA 1500  
CACTATAGAA TAACATCCAC TTTGCCTTTC TCTCCACAGG TGTCACCTCA 1550  
50 GGTCAACTGC ACCTCGGTTC TATCGATTGA ATTCCCGGC CATAGCTGTC 1600  
55 TGGCATGGGC CTCTCCACCG TGCCTGACCT GCTGCTGCCG CTGGTGCTCC 1650  
TGGAGCTGTT GGTGGGAATA TACCCCTCAG GGGTTATTGG ACTGGTCCCT 1700  
60 CACCTAGGGG ACAGGGAGAA GAGAGATAGT GTGTGTCCCC AAGGAAAATA 1750  
TATCCACCCT CAAAATAATT CGATTTGCTG TACCAAGTGC CACAAAGGAA 1800  
65 CCTACTTGTA CAATGACTGT CCAGGCCCGG GGCAGGATAC GGAAGTGCAGG 1850

GAGTGTGAGA GCGGCTCCTT CACCGCTTCA GAAAACCACC TCAGACACTG 1900  
5 CCTCAGCTGC TCCAAATGCC GAAAGGAAAT GGGTCAGGTG GAGATCTCTT 1950  
CTTGCACAGT GGACCGGGAC ACCGTGTGTG GCTGCAGGAA GAACCAGTAC 2000  
10 CGGCATTATT GGAGTGAAAA CCTTTTCCAG TGCTTCAATT GCAGCCTCTG 2050  
CCTCAATGGG ACCGTGCACC TCTCCTGCCA GGAGAAACAG AACACCGTGT 2100  
15 GCACCTGCCA TGCAGGTTTC TTTCTAAGAG AAAACGAGTG TGTCTCCTGT 2150  
AGTAACTGTA AGAAAAGCCT GGAGTGCACG AAGTTGTGCC TACCCAGAT 2200  
20 TGAGAATGTT AAGGGCACTG AGGACTCAGG CACCACAGAC AAGAGAGTTG 2250  
AGCTCAAAAC CCCACTTGGT GACACAATC ACACATGCCC ACGGTGCCCA 2300  
GAGCCCAAAT CTTGTGACAC ACCTCCCCCG TGCCCACGGT GCCCAGAGCC 2350  
30 CAAATCTTGT GACACACCTC CCCCATGCCC ACGGTGCCCA GAGCCCAAAT 2400  
CTTGTGACAC ACCTCCCCCA TGCCCACGGT GCCCAGCACC TGAACCTCTG 2450  
35 GGAGGACCGT CAGTCTTCCT CTTCCCCCA AAACCAAGG ATACCCTTAT 2500  
GATTTCCCGG ACCCCTGAGG TCACGTGCGT GGTGGTGGAC GTGAGCCACG 2550  
AAGACCCCGA GGTCCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT 2600  
45 AATGCCAAGA CAAAGCCGCG GGAGGAGCAG TTCAACAGCA CGTTCCGTGT 2650  
GGTCAGCGTC CTCACCGTCC TGCACCAGGA CTGGCTGAAC GGCAAGGAGT 2700  
50 ACAAGTGCAA GGTCTCCAAC AAAGCCCTCC CAGCCCCCAT CGAGAAAACC 2750  
ATCTCCAAAA CCAAAGGACA GCCCCGAGAA CCACAGGTGT ACACCCTGCC 2800  
CCCATCCCGG GAGGAGATGA CCAAGAACCA GGTGAGCCTG ACCTGCCTGG 2850  
60 TCAAAGGCTT CTACCCAGC GACATCGCCG TGGAGTGGGA GAGCAGCGGG 2900  
CAGCCGGAGA ACAACTACAA CACCACGCCT CCCATGCTGG ACTCCGACGG 2950  
65 CTCCTTCTTC CTCTACAGCA AGCTCACCGT GGACAAGAGC AGGTGGCAGC 3000

AGGGGAACAT CTTCTCATGC TCCGTGATGC ATGAGGCTCT GCACAACCGC 3050

5 TTCACGCAGA AGAGCCTCTC CCTGTCTCCG GGTAAATGAG TGCGACGGCC 3100

GGGGATCCTC TAGAGTCGAC CTGCAGAAGC TTGGCCGCCA TGGCCCAACT 3150

10 TGTTTATTGC AGCTTATAAT GGTTACAAAT AAAGCAATAG CATCACAAAT 3200

TTCACAAATA AAGCATTTTT TCACTGCAT TCTAGTTGTG GTTTGTCCAA 3250

15 ACTCATCAAT GTATCTTATC ATGTCTGGAT CGATCGGGAA TTAATTCGGC 3300

GCAGCACCAT GGCCTGAAAT AACCTCTGAA AGAGGAACTT GGTTAGGTAC 3350

20 CTTCTGAGGC GGAAAGAACC AGCTGTGGAA TGTGTGTCAG TTAGGGTGTG 3400

25 GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAAG CATGCATCTC 3450

AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG 3500

30 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACCATA GTCCCGCCCC 3550

TAACTCCGCC CATCCCGCCC CTA ACTCCGC CCAGTTCCGC CCATTCTCCG 3600

35 CCCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGGCCG AGGCCGCCTC 3650

40 GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCCTAG 3700

GCTTTTGCAA AAAGCTGTTA ACAGCTGGC ACTGGCCGTC GTTTTACAAC 3750

45 GTCGTGACTG GGAAAACCCT GCGGTTACCC AACTTAATCG CCTTGCAGCA 3800

CATCCCCCT TCGCCAGCTG GCGTAATAGC GAAGAGGCC GCACCGATCG 3850

50 CCCTTCCCAA CAGTTGCGTA GCCTGAATGG CGAATGGCGC CTGATGCGGT 3900

55 ATTTTCTCCT TACGCATCTG TCGGTATTT CACACCGCAT ACGTCAAAGC 3950

AACCATAGTA CGCGCCCTGT AGCGGCGCAT TAAGCGCGG GGGTGTGGTG 4000

60 GTTACGCGCA GCGTGACCGC TACACTTGCC AGCGCCCTAG CGCCCGCTCC 4050

TTTCGCTTTC TTCCCTTCCT TTCTCGCCAC GTTCGCGGC TTTCCCGTC 4100

65 AAGCTCTAAA TCGGGGGCTC CCTTTAGGGT TCCGATTTAG TGCTTTACGG 4150

CACCTCGACC CCAAAAACT TGATTGGGT GATGGTTCAC GTAGTGGGCC 4200

5 ATCGCCCTGA TAGACGGTTT TTCGCCCTTT GACGTGGAG TCCACGTTCT 4250

TTAATAGTGG ACTCTGTTC CAACTGGAA CAACACTCAA CCCTATCTCG 4300

10 GGCTATTCTT TTGATTATA AGGGATTTG CCGATTTCGG CCTATTGGTT 4350

AAAAATGAG CTGATTAAAC AAAAATTAA CGCGAATTTT AACAAAATAT 4400

15 TAACGTTTAC AATTTTATGG TGCACTCTCA GTACAATCTG CTCTGATGCC 4450

GCATAGTTAA GCCAACTCCG CTATCGCTAC GTGACTGGGT CATGGCTGCG 4500

20 CCCCACACC CGCCAACACC CGCTGACGCG CCCTGACGGG CTTGTCTGCT 4550

25 CCCGGCATCC GCTTACAGAC AAGCTGTGAC CGTCTCCGGG AGCTGCATGT 4600

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30 AGACGAAAGG GCCTCGTGAT ACGCCTATTT TTATAGGTTA ATGTCATGAT 4700

AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA AATGTGCGCG 4750

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ATGAGACAAT AACCTGATA AATGCTTCAA TAATATTGAA AAAGGAAGAG 4850

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45 TTTGCCTTCC TGTTTTTGCT CACCCAGAAA CGCTGGTGAA AGTAAAGAT 4950

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50 CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT TTTCCAATGA 5050

55 TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGATGAC 5100

GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC AGAATGACTT 5150

60 GGTGAGTAC TCACCACTCA CAGAAAAGCA TCTTACGGAT GGCATGACAG 5200

TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA CACTGCGGCC 5250

65 AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA CCGCTTTTTT 5300

GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG GAACCGGAGC 5350  
5 TGAATGAAGC CATACCAAAC GACGAGCGTG ACACCACGAT GCCAGCAGCA 5400  
ATGGCAACAA CGTTGCGCAA ACTATTA ACT GGC GAACTAC TTACTCTAGC 5450  
10 TTCCCGGCAA CAATTAATAG ACTGGATGGA GCGGATAAA GTTGCAGGAC 5500  
CACTTCTGCG CTCGGCCCTT CCGGCTGGCT GGT TTTATTGC TGATAAATCT 5550  
15 GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC TGGGGCCAGA 5600  
TGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGG AGTCAGGCAA 5650  
20 CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC CTCACTGATT 5700  
AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA 5750  
TTTAAACTT CATTTTAAAT TTAAAGGAT CTAGGTGAAG ATCCTTTTGTG 5800  
30 ATAATCTCAT GACCAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG 5850  
TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC CTTTTTTTCT 5900  
35 GCGCGTAATC TGCTGCTTGC AAACAAAAA ACCACCGCTA CCAGCGGTGG 5950  
40 TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC 6000  
TTCAGCAGAG CGCAGATACC AAATACTGTC CTTCTAGTGT AGCCGTAGTT 6050  
45 AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC 6100  
TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC GTGTCTTACC 6150  
50 GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC GGTCGGGCTG 6200  
AACGGGGGGT TCGTGCACAC AGCCAGCTT GGAGCGAACG ACCTACACCG 6250  
AACTGAGATA CCTACAGCGT GAGCATTGAG AAAGCGCCAC GCTTCCCGAA 6300  
60 GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG GAACAGGAGA 6350  
GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT TATAGTCCTG 6400  
65 TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTGTG ATGCTCGTCA 6450



GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACGCGGCCT TTTTACGGTT 6500  
5 CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTTCCT GCGTTATCCC 6550  
CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC TGATACCGCT 6600  
10 CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG AGGAAGCGGA 6650  
AGAGCGCCCA ATACGCAAAC CGCCTCTCCC CGCGCGTTGG CCGATTCAAT 6700  
15 AATCCAGCTG GCACGACAGG TTTCCCGACT GGAAAGCGGG CAGTGAGCGC 6750  
AACGCAATTA ATGTGAGTTA CCTCACTCAT TAGGCACCCC AGGCTTTACA 6800  
20 CTTTATGCTT CCGGCTCGTA TGTTGTGTGG AATTGTGAGC GGATAACAAT 6850  
25 TTCACACAGG AAACAGCTAT GACCATGATT ACGAATTAA 6889

## (2) INFORMATION FOR SEQ ID NO:3:

30 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 6557 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
35 (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

40 TTCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG 50  
GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGCAGGCA 100  
45 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACCAG GTGTGGAAAG 150  
TCCCCAGGCT CCCGAGCAGG CAGAAGTATG CAAAGCATGC ATCTCAATTA 200  
50 GTCAGCAACC ATAGTCCCGC CCCTAACTCC GCCCATCCCG CCCCTAACTC 250  
CGCCCAGTTC CGCCCATCTC CCGCCCCATG GCTGACTAAT TTTTTTTATT 300  
55 TATGCAGAGG CCGAGGCCGC CTCGGCCTCT GAGCTATTCC AGAAGTAGTG 350  
60 AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAGCTA GCTTATCCGG 400  
CCGGGAACGG TGCATTGGAA CGCGGATTCC CCGTGCCAAG AGTGACGTAA 450  
65 GTACCGCCTA TAGAGCGATA AGAGGATTTT ATCCCCGCTG CCATCATGGT 500

TCGACCATTG AACTGCATCG TCGCCGTGTC CCAAATATG GGGATTGGCA 550  
5 AGAACGGAGA CCTACCCTGG CCTCCGCTCA GGAACGAGTT CAAGTACTTC 600  
CAAAGAATGA CCACAACCTC TTCAGTGGAA GGTAACAGA ATCTGGTGAT 650  
10 TATGGGTAGG AAAACCTGGT TCTCCATTCC TGAGAAGAAT CGACCTTTAA 700  
AGGACAGAAT TAATATAGTT CTCAGTAGAG AACTCAAAGA ACCACCACGA 750  
15 GGAGCTCATT TTCTTGCCAA AAGTTTGGAT GATGCCTTAA GACTTATTGA 800  
ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTTGGATA GTCGGAGGCA 850  
20 GTTCTGTTTA CCAGGAAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT 900  
25 GTGACAAGGA TCATGCAGGA ATTTGAAAGT GACACGTTTT TCCCAGAAAT 950  
TGATTTGGGG AAATATAAAC CTCTCCAGA ATACCCAGGC GTCCTCTCTG 1000  
30 AGGTCCAGGA GGAAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAAG 1050  
AAAGACTAAC AGGAAGATGC TTTCAAGTTC TCTGCTCCCC TCCTAAAGCT 1100  
35 ATGCATTTTT ATAAGACCAT GGGACTTTTG CTGGCTTTAG ATCCCCTTGG 1150  
40 CTTGTTAGA ACGCAGCTAC AATTAATACA TAACCTTATG TATCATACAC 1200  
ATACGATTTA GGTGACACTA TAGATAACAT CCACTTTGCC TTTCTCTCCA 1250  
45 CAGGTGTCCA CTCCCAGGTC CAACTGCACC TCGGTTCTAT CGATTGAATT 1300  
CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT 1350  
50 GGAGTACATT CAGAAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTGCA 1400  
55 GCCAGGGGGC TCACTCCGTT TGTCTGTGC AGTTTCTGGC TACTCCATCA 1450  
CCTCCGGATA TAGCTGGAAC TGGATCCGTC AGGCCCCGGG TAAGGGCCTG 1500  
60 GAATGGGTTG CATCGATTAC GTATGCCGGA TCGACTAACT ATAACCCTAG 1550  
CGTCAAGGGC CGTATCACTA TAAGTCGCGA CGATTCCAAA AACACATTCT 1600  
65 ACCTGCAGAT GAACAGCCTG CGTGCTGAGG AACTGCCGT CTATTATTGT 1650

GCTCGAGGCA GCCACTATTT CGGCGCCTGG CACTTCGCCG TGTGGGGTCA 1700  
5 AGGAACCCTG GTCACCGTCT CCTCGGCCTC CACCAAGGGC CCATCGGTCT 1750  
TCCCCCTGGC ACCCTCCTCC AAGAGCACCT CTGGGGGCAC AGCGGCCCTG 1800  
10 GGCTGCCTGG TCAAGGACTA CTTCCCCGAA CCGGTGACGG TGTCGTGGAA 1850  
CTCAGGCGCC CTGACCAGCG GCGTGACAC CTTCCCGGCT GTCCTACAGT 1900  
15 CCTCAGGACT CTACTCCCTC AGCAGCGTGG TGACTGTGCC CTCTAGCAGC 1950  
TTGGGCACCC AGACCTACAT CTGCAACGTG AATCACAAGC CCAGCAACAC 2000  
20 CAAGGTGGAC AAGAAAGTTG AGCCCAAATC TTGTGACAAA ACTCACACAT 2050  
25 GCCCACCGTG CCCAGCACCT GAACTCCTGG GGGGACCGTC AGTCTTCCTC 2100  
TTCCCCCAA AACCCAAGGA CACCCTCATG ATCTCCCGGA CCCCTGAGGT 2150  
30 CACATGCGTG GTGGTGGACG TGAGCCACGA AGACCCTGAG GTCAAGTTCA 2200  
ACTGGTACGT GGACGGCGTG GAGGTGCATA ATGCCAAGAC AAAGCCGCGG 2250  
35 GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTCAGCGTCC TCACCGTCCT 2300  
GCACCAGGAC TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA 2350  
AAGCCCTCCC AGCCCCATC GAGAAAACCA TCTCCAAAGC CAAAGGGCAG 2400  
45 CCCCAGAAAC CACAGGTGTA CACCCTGCCC CCATCCCGGG AAGAGATGAC 2450  
CAAGAACCAG GTCAGCCTGA CTTGCCTGGT CAAAGGCTTC TATCCCAGCG 2500  
50 ACATCGCCGT GGAGTGGGAG AGCAATGGGC AGCCGGAGAA CAACTACAAG 2550  
55 ACCACGCCTC CCGTGCTGGA CTCCGACGGC TCCTTCTTCC TCTACAGCAA 2600  
GCTCACCGTG GACAAGAGCA GGTGGCAGCA GGGGAACGTC TTCTCATGCT 2650  
60 CCGTGATGCA TGAGGCTCTG CACAACCACT ACACGCAGAA GAGCCTCTCC 2700  
CTGTCTCCGG GTAAATGAGT GCGACGGCCC TAGAGTCGAC CTGCAGAAGC 2750  
65 TTGGCCGCCA TGGCCCAACT TGTTTATTGC AGCTTATAAT GGTACAAAT 2800

AAAGCAATAG CATCACAAAT TTCACAAATA AAGCATTTTT TTCACTGCAT 2850

5 TCTAGTTGTG GTTTGTCCAA ACTCATCAAT GTATCTTATC ATGTCTGGAT 2900

CGATCGGGAA TTAATTCGGC GCAGCACCAT GGCCTGAAAT AACCTCTGAA 2950

10 AGAGGAACTT GGTTAGGTAC CTTCTGAGGC GGAAAGAACC AGCTGTGGAA 3000

TGTGTGTCAG TTAGGGTGTG GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA 3050

15 GTATGCAAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC 3100

CCAGGCTCCC CAGCAGGCAG AAGTATGCAA AGCATGCATC TCAATTAGTC 3150

20 AGCAACCATA GTCCCGCCCC TAACTCCGCC CATCCCGCCC CTA ACTCCGC 3200

25 CCAGTTCCGC CCATTCTCCG CCCCATGGCT GACTAATTTT TTTTATTAT 3250

GCAGAGGCCG AGGCCGCCTC GGCCTCTGAG CTATTCCAGA AGTAGTGAGG 3300

30 AGGCTTTTTT GGAGGCCTAG GCTTTTGCAA AAAGCTGTTA CCTCGAGCGG 3350

CCGCTTAATT AAGGCGCGCC ATTTAAATCC TGCAGGTAAC AGCTTGGCAC 3400

35 TGGCCGTCGT TTTACAACGT CGTGACTGGG AAAACCCTGG CGTTACCCAA 3450

40 CTTAATCGCC TTGCAGCACA TCCCCCTTC GCCAGCTGGC GTAATAGCGA 3500

AGAGGCCCCG ACCGATCGCC CTTCCCAACA GTTGCGTAGC CTGAATGGCG 3550

45 AATGGCGCCT GATGCGGTAT TTTCTCCTTA CGCATCTGTG CGGTATTTC 3600

CACCGCATAC GTCAAAGCAA CCATAGTACG CGCCCTGTAG CGGCGCATT 3650

50 AGCGCGGCGG GTGTGGTGGT TACGCGCAGC GTGACCGCTA CACTTGCCAG 3700

55 CGCCCTAGCG CCCGCTCCTT TCGCTTCTT CCCTTCCTTT CTCGCCACGT 3750

TCGCCGGCTT TCCCCGTCAA GCTCTAAATC GGGGGCTCCC TTTAGGGTTC 3800

60 CGATTTAGTG CTTTACGGCA CCTCGACCCC AAAAACTTG ATTTGGGTGA 3850

TGGTTCACGT AGTGGGCCAT CGCCCTGATA GACGGTTTTT CGCCCTTGA 3900

65 CGTTGGAGTC CACGTTCTTT AATAGTGGAC TCTTGTTCCTA AACTGGAACA 3950

ACACTCAACC CTATCTCGGG CTATTCTTTT GATTTATAAG GGATTTTGCC 4000  
GATTTCGGCC TATTGGTTAA AAAATGAGCT GATTTAACAA AAATTTAACG 4050  
5 CGAATTTTAA CAAAATATTA ACGTTTACAA TTTTATGGTG CACTCTCAGT 4100  
10 ACAATCTGCT CTGATGCCGC ATAGTTAAGC CAACTCCGCT ATCGCTACGT 4150  
GACTGGGTCA TGGCTGCGCC CCGACACCCG CCAACACCCG CTGACGCGCC 4200  
15 CTGACGGGCT TGTCTGCTCC CGGCATCCGC TTACAGACAA GCTGTGACCG 4250  
TCTCCGGGAG CTGCATGTGT CAGAGGTTTT CACCGTCATC ACCGAAACGC 4300  
20 GCGAGGCAGT ATTCTTGAAG ACGAAAGGGC CTCGTGATAC GCCTATTTTT 4350  
25 ATAGGTTAAT GTCATGATAA TAATGGTTTC TTAGACGTCA GGTGGCACTT 4400  
TTCGGGGAAA TGTGCGCGGA ACCCCTATTT GTTTATTTTT CTAAATACAT 4450  
30 TCAAATATGT ATCCGCTCAT GAGACAATAA CCCTGATAAA TGCTTCAATA 4500  
ATATTGAAAA AGGAAGAGTA TGAGTATTCA ACATTTCCGT GTCGCCCTTA 4550  
35 TTCCCTTTTT TGCGGCATTT TGCCTTCCTG TTTTGTCTCA CCCAGAAACG 4600  
40 CTGGTGAAAG TAAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGGTTA 4650  
CATCGAACTG GATCTCAACA GCGGTAAGAT CCTTGAGAGT TTTCGCCCCG 4700  
45 AAGAACGTTT TCCAATGATG AGCACTTTTA AAGTTCTGCT ATGTGGCGCG 4750  
GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTC GCCGCATACA 4800  
50 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAAGCATC 4850  
55 TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTGCTGC CATAACCATG 4900  
AGTGATAACA CTGCGGCCAA CTTACTTCTG ACAACGATCG GAGGACCGAA 4950  
60 GGAGCTAACC GCTTTTTTGC ACAACATGGG GGATCATGTA ACTCGCCTTG 5000  
ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCAAACGA CGAGCGTGAC 5050  
65 ACCACGATGC CAGCAGCAAT GGCAACAACG TTGCGCAAAC TATTAAGTGG 5100

CGAACTACTT ACTCTAGCTT CCCGGCAACA ATTAATAGAC TGGATGGAGG 5150

5 CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGGCTGG 5200

TTTATTGCTG ATAAATCTGG AGCCGGTGAG CGTGGGTCTC GCGGTATCAT 5250

10 TGCAGCACTG GGGCCAGATG GTAAGCCCTC CCGTATCGTA GTTATCTACA 5300

CGACGGGGAG TCAGGCAACT ATGGATGAAC GAAATAGACA GATCGCTGAG 5350

15 ATAGGTGCCT CACTGATTAA GCATTGGTAA CTGTCAGACC AAGTTTACTC 5400

ATATATACTT TAGATTGATT TAAAACTTCA TTTTAAATTT AAAAGGATCT 5450

20 AGGTGAAGAT CCTTTTGTAT AATCTCATGA CCAAATCCC TTAACGTGAG 5500

TTTTCGTTCC ACTGAGCGTC AGACCCCGTA GAAAAGATCA AAGGATCTTC 5550

TTGAGATCCT TTTTCTCTGC GCGTAATCTG CTGCTTGCAA ACAAAAAAAC 5600

30 CACCGCTACC AGCGGTGGTT TGTTCGCCG ATCAAGAGCT ACCAACTCTT 5650

TTTCCGAAGG TAACTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCCT 5700

35 TCTAGTGTAG CCGTAGTTAG GCCACCATT CAAGAACTCT GTAGCACCGC 5750

CTACATACCT CGCTCTGCTA ATCCTGTTAC CAGTGGCTGC TGCCAGTGGC 5800

GATAAGTCGT GTCTTACCGG GTTGGACTCA AGACGATAGT TACCGGATAA 5850

45 GGCGCAGCGG TCGGGCTGAA CGGGGGGTTC GTGCACACAG CCCAGCTTGG 5900

AGCGAACGAC CTACACCGAA CTGAGATACC TACAGCGTGA GCATTGAGAA 5950

50 AGCGCCACGC TTCCGAAGG GAGAAAGGCG GACAGGTATC CGGTAAGCGG 6000

CAGGGTCGGA ACAGGAGAGC GCACGAGGGA GCTTCCAGGG GGAAACGCCT 6050

GGTATCTTTA TAGTCCTGTC GGGTTTCGCC ACCTCTGACT TGAGCGTCGA 6100

60 TTTTGTGAT GCTCGTCAGG GGGGCGGAGC CTATGGAAAA ACGCCAGCAA 6150

CGCGGCCTTT TTACGGTTCC TGGCCTTTTG CTGGCCTTTT GCTCACATGT 6200

65 TCTTCTCTGC GTTATCCCCT GATTCTGTGG ATAACCGTAT TACCGCCTTT 6250

GAGTGAGCTG ATACCGCTCG CCGCAGCCGA ACGACCGAGC GCAGCGAGTC 6300  
AGTGAGCGAG GAAGCGGAAG AGCGCCCAAT ACGCAAACCG CCTCTCCCCG 6350  
5 CGCGTTGGCC GATTCATTAA TCCAGCTGGC ACGACAGGTT TCCCGACTGG 6400  
10 AAAGCGGGCA GTGAGCGCAA CGCAATTAAT GTGAGTTACC TCACTCATT 6450  
GGCACCCAG GCTTTACTT TATGCTTCC GGCTCGTATG TTGTGTGGAA 6500  
15 TTGTGAGCGG ATAACAATTT CACACAGGAA ACAGCTATGA CCATGATTAC 6550  
GAATTAA 6557  
20

## (2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 7305 bases  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: double  
(D) TOPOLOGY: linear

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TTGAGCTCG CCCGACATTG ATTATTGACT AGTTATTAAT AGTAATCAAT 50  
35 TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC 100  
40 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATTG 150  
ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA 200  
45 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCCCAC TTGGCAGTAC 250  
ATCAAGTGTA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT 300  
50 AAATGGCCCG CCTGGCATTG TGCCAGTAC ATGACCTTAT GGGACTTTCC 350  
55 TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC 400  
GGTTTTGGCA GTACATCAAT GGGCGTGGAT AGCGGTTTGA CTCACGGGGA 450  
60 TTTCCAAGTC TCCACCCCAT TGACGTCAAT GGGAGTTTGT TTTGGCACCA 500  
AAATCAACGG GACTTTCCAA AATGTCGTAA CAACTCCGCC CCATTGACGC 550  
65 AAATGGGCGG TAGGCGTGTA CGGTGGGAGG TCTATATAAG CAGAGCTCGT 600

TTAGTGAACC GTCAGATCGC CTGGAGACGC CATCCACGCT GTTTTGACCT 650

5 CCATAGAAGA CACCGGGACC GATCCAGCCT CCGCGGCCGG GAACGGTGCA 700

TTGGAACGCG GATTCCCCGT GCCAAGAGTG ACGTAAGTAC CGCCTATAGA 750

10 GTCTATAGGC CCACCCCTT GGCTTCGTTA GAACGGGCT ACAATTAATA 800

CATAACCTTA TGTATCATAC ACATACGATT TAGGTGACAC TATAGAATAA 850

15 CATCCACTTT GCCTTCTCT CCACAGGTGT CCACTCCCAG GTCCAACGTC 900

ACCTCGGTTC TAAGCTTATC GATATGAAA AGCCTGAACT CACCGCGACG 950

20 TCTGTCGAGA AGTTTCTGAT CGAAAAGTTC GACAGCGTCT CCGACCTGAT 1000

25 GCAGCTCTCG GAGGGCGAAG AATCTCGTGC TTTCAGCTTC GATGTAGGAG 1050

GGCGTGGATA TGTCTGCGG GTAAATAGCT GCGCCGATGG TTTCTACAAA 1100

30 GATCGTTATG TTTATCGGCA CTTTGCATCG GCCGCGCTCC CGATTCCGGA 1150

AGTGCTTGAC ATTGGGGAAT TCAGCGAGAG CCTGACCTAT TGCATCTCCC 1200

35 GCCGTGCACA GGGTGTACG TTGCAACACC TGCCTGAAAC CGAACTGCCC 1250

40 GCTGTTCTGC AGCCGGTCGC GGAGGCCATG GATGCGATCG CTGCGGCCGA 1300

TCCTAGCCAG ACGAGCGGGT TCGGCCCATT CGGACCGCAA GGAATCGGTC 1350

45 AATACACTAC ATGGCGTGAT TTCATATGCG CGATTGCTGA TCCCCATGTG 1400

TATCACTGGC AAACGTGAT GGACGACACC GTCAGTGGT CCGTCGCGCA 1450

50 GGCTCTCGAT GAGCTGATGC TTTGGGCCGA GGAATGCCCC GAAGTCCGGC 1500

55 ACCTCGTGCA CGCGGATTTC GGCTCCAACA ATGTCCTGAC GGACAATGGC 1550

CGCATAACAG CGGTCATTGA CTGGAGCGAG GCGATGTTTC GGGATTCCCA 1600

60 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCCGTGGTTG GCTTGTATGG 1650

AGCAGCAGAC GTACTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCCG 1700

65 CGGCTCCGGG CGTATATGCT CCGCATTGGT CTTGACCAAC TCTATCAGAG 1750



CTTGGTTGAC GGCAATTTTCG ATGATGCAGC TTGGGCGCAG GGTGATGCG 1800

5 ACGCAATCGT CCGATCCGGA GCCGGGACTG TCGGGCGTAC ACAAATCGCC 1850

CGCAGAAGCG CGGCCGTCTG GACCGATGGC TGTGTAGAAG TACTCGCCGA 1900

10 TAGTGGAAC CGACGCCCCA GCACTCGTCC GAGGGCAAAG GAATAGAGTA 1950

GATGCCGACC GAAGGATCCC CGGGGAATTC AATCGATGGC CGCCATGGCC 2000

15 CAACTTGTTT ATTGCAGCTT ATAATGGTTA CAAATAAAGC AATAGCATCA 2050

CAAATTTTAC AAATAAAGCA TTTTTCAC TGCATTCTAG TTGTGGTTTG 2100

20 TCCAAACTCA TCAATGTATC TTATCATGTC TGGATCGATC GGGAATTAAT 2150

TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAAGAGG AACTTGGTTA 2200

GGTACCTTCT GAGGCGGAAA GAACCAGCTG TGGAATGTGT GTCAGTTAGG 2250

30 GTGTGGAAG TCCCCAGGCT CCCCAGCAGG CAGAAGTATG CAAAGCATGC 2300

ATCTCAATTA GTCAGCAACC AGGTGTGGAA AGTCCCCAGG CTCCCCAGCA 2350

GGCAGAAGTA TGCAAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC 2400

40 GCCCCTAACT CCGCCCATCC CGCCCCTAAC TCCGCCAGT TCCGCCATT 2450

CTCCGCCCCA TGGCTGACTA ATTTTCTTA TTTATGCAGA GGCCGAGGCC 2500

45 GCCTCGGCCT CTGAGCTATT CCAGAAGTAG TGAGGAGGCT TTTTGGAGG 2550

CCTAGGCTTT TGCAAAAAGC TAGCTTATCC GGCCGGGAAC GGTGCATTGG 2600

50 AACGCGGATT CCCCCTGCCA AGAGTCAGGT AAGTACCGCC TATAGAGTCT 2650

ATAGGCCAC CCCCTGGCT TCGTTAGAAC GCGGCTACAA TTAATACATA 2700

ACCTTTTGGA TCGATCCTAC TGACACTGAC ATCCACTTTT TCTTTTCTC 2750

60 CACAGGTGTC CACTCCCAGG TCCAACGCA CCTCGGTTTCG CGAAGCTAGC 2800

TTGGGCTGCA TCGATTGAAT TCCACCATGG GATGGTCATG TATCATCCTT 2850

65 TTTCTAGTAG CAACTGCAAC TGGAGTACAT TCAGATATCC AGCTGACCCA 2900

GTCCCCGAGC TCCCTGTCCG CCTCTGTGGG CGATAGGGTC ACCATCACCT 2950

5 GCCGTGCCAG TCAGAGCGTC GATTACGATG GTGATAGCTA CATGAACTGG 3000

TATCAACAGA AACCAGGAAA AGCTCCGAAA CTACTGATTT ACGCGGCCTC 3050

10 GTACCTGGAG TCTGGAGTCC CTTCTCGCTT CTCTGGATCC GGTTCCTGGGA 3100

CGGATTTTAC TCTGACCATC AGCAGTCTGC AGCCGGAAGA CTTGCAACT 3150

15 TATTACTGTC AGCAAAGTCA CGAGGATCCG TACACATTTG GACAGGGTAC 3200

CAAGGTGGAG ATCAAACGAA CTGTGGCTGC ACCATCTGTC TTCATCTTCC 3250

20 CGCCATCTGA TGAGCAGTTG AAATCTGGAA CTGCCTCTGT TGTGTGCCTG 3300

25 CTGAATAACT TCTATCCCAG AGAGGCCAAA GTACAGTGGA AGGTGGATAA 3350

CGCCCTCCAA TCGGGTAACT CCCAGGAGAG TGTCACAGAG CAGGACAGCA 3400

30 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC 3450

TACGAGAAAC ACAAAGTCTA CGCCTGCGAA GTCACCCATC AGGGCCTGAG 3500

35 CTCGCCCCGTC ACAAAGAGCT TCAACAGGGG AGAGTGTTAA GCTTCGATGG 3550

40 CCGCCATGGC CCAACTTGTT TATTGCAGCT TATAATGGTT ACAAATAAAG 3600

CAATAGCATC ACAAATTTCA CAAATAAAGC ATTTTTTTCA CTGCATTCTA 3650

45 GTTGTGGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGGATCGAT 3700

CGGGAATTAA TTCGGCGCAG CACCATGGCC TGAAATAACC TCTGAAAGAG 3750

50 GAACTTGGTT AGGTACCTTC TGAGGCGGAA AGAACCAGCT GTGGAATGTG 3800

55 TGTCAGTTAG GGTGTGGAAA GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT 3850

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65 TTCCGCCCAT TCTCCGCCCC ATGGCTGACT AATTTTTTTT ATTTATGCAG 4050

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5 TTTTTTGGAG GCCTAGGCTT TTGCAAAAAG CTGTTAACAG CTTGGCACTG 4150

GCCGTCGTTT TACAACGTCG TGA CTGGGAA AACCTGGCG TTACCCA ACT 4200

10 TAATCGCCTT GCAGCACATC CCCCCTTCGC CAGCTGGCGT AATAGCGAAG 4250

AGGCCCCGAC CGATCGCCCT TCCCAACAGT TGCCTAGCCT GAATGGCGAA 4300

15 TGGCGCCTGA TGGGTATTT TCTCCTTACG CATCTGTGCG GTATTTTACA 4350

CCGCATACGT CAAAGCAACC ATAGTACGCG CCCTGTAGCG GCGCATTAAG 4400

20 CGCGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG 4450

CCCTAGCGCC CGCTCCTTTC GCTTCTTCC CTTCTTTCT CGCCACGTTT 4500

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30 ATTTAGTGCT TTACGGCACC TCGACCCCAA AAACTTGAT TTGGGTGATG 4600

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ACTCAACCCT ATCTCGGGCT ATTCTTTTGA TTTATAAGGG ATTTTGCCGA 4750

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45 AATTTTAACA AAATATTAAC GTTTACAATT TTATGGTGCA CTCTCAGTAC 4850

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50 CTGGGTCATG GCTGCGCCCC GACACCCGCC AACACCCGCT GACGCGCCCT 4950

GACGGGCTTG TCTGCTCCCG GCATCCGCTT ACAGACAAGC TGTGACCGTC 5000

TCCGGGAGCT GCATGTGTCA GAGGTTTTCA CCGTCATCAC CGAAACGCGC 5050

60 GAGGCAGTAT TCTTGAAGAC GAAAGGGCCT CGTGATACGC CTATTTTTAT 5100

AGGTTAATGT CATGATAATA ATGGTTTCTT AGACGTCAGG TGGCACTTTT 5150

65 CGGGGAAATG TGC GCGGAAC CCCTATTTGT TTATTTTTCT AAATACATTC 5200

AAATATGTAT CCGCTCATGA GACAATAACC CTGATAAATG CTTCAATAAT 5250

ATTGAAAAAG GAAGAGTATG AGTATTCAAC ATTTCCGTGT CGCCCTTATT 5300

5 CCCTTTTTTG CGGCATTTTG CCTTCCTGTT TTTGCTCACC CAGAAACGCT 5350

GGTGAAAGTA AAAGATGCTG AAGATCAGTT GGGTGACGA GTGGGTTACA 5400

10 TCGAACTGGA TCTCAACAGC GGTAAGATCC TTGAGAGTTT TCGCCCCGAA 5450

GAACGTTTTT CAATGATGAG CACTTTTAAA GTTCTGCTAT GTGGCGCGGT 5500

15 ATTATCCCGT GATGACGCCG GGCAAGAGCA ACTCGGTCGC CGCATACACT 5550

20 ATTCTCAGAA TGA CTGTTGGTT GAGTACTCAC CAGTCACAGA AAAGCATCTT 5600

ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG 5650

25 TGATAACACT GCGGCCAACT TACTTCTGAC AACGATCGGA GGACCGAAGG 5700

30 AGCTAACCGC TTTTGTGCAC AACATGGGGG ATCATGTAAC TCGCCTTGAT 5750

CGTTGGGAAC CGGAGCTGAA TGAAGCCATA CCAAACGACG AGCGTGACAC 5800

35 CACGATGCCA GCAGCAATGG CAACAACGTT GCGCAAATA TTA ACTGGCG 5850

AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG 5900

40 GATAAAGTTG CAGGACCACT TCTGCGCTCG GCCCTTCCGG CTGGCTGGTT 5950

45 TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTCGC GGTATCATTG 6000

CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG 6050

50 ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA TCGCTGAGAT 6100

AGGTGCCTCA CTGATTAAGC ATTGGTAACT GTCAGACCAA GTTTACTCAT 6150

ATATACTTTA GATTGATTTA AAACCTTCATT TTAAATTTAA AAGGATCTAG 6200

60 GTGAAGATCC TTTTGTGATA TCTCATGACC AAAATCCCTT AACGTGAGTT 6250

TTCGTTCCAC TGAGCGTCAG ACCCCGTAGA AAAGATCAAA GGATCTTCTT 6300

65 GAGATCCTTT TTTTCTGCGC GTAATCTGCT GCTTGCAAAC AAAAAACCA 6350

CCGCTACCAG CGGTGGTTTG TTTGCCGGAT CAAGAGCTAC CAACTCTTTT 6400  
TCCGAAGGTA ACTGGCTTCA GCAGAGCGCA GATACCAAAT ACTGTCCTTC 6450  
5 TAGTGTAGCC GTAGTTAGGC CACCACTTCA AGAACTCTGT AGCACC GCCT 6500  
ACATACCTCG CTCTGCTAAT CCTGTTACCA GTGGCTGCTG CCAGTGGCGA 6550  
10 TAAGTCGTGT CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG 6600  
TAAGTCGTGT CTTACCGGGT TGGACTCAAG ACGATAGTTA CCGGATAAGG 6600  
15 CGCAGCGGTC GGGCTGAACG GGGGGTTCGT GCACACAGCC CAGCTTGGAG 6650  
CGAACGACCT ACACCGAACT GAGATACCTA CAGCGTGAGC ATTGAGAAAG 6700  
20 CGCCACGCTT CCCGAAGGGA GAAAGGCGGA CAGGTATCCG GTAAGCGGCA 6750  
GGGTCGGAAC AGGAGAGCGC ACGAGGGAGC TTCCAGGGGG AAACGCCTGG 6800  
25 TATCTTTATA GTCCTGTCGG GTTTCGCCAC CTCTGACTTG AGCGTCGATT 6850  
TTTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGAAAAC GCCAGCAACG 6900  
30 CGGCCTTTTT ACGGTTCCCTG GCCTTTTGCT GGCCTTTTGC TCACATGTTC 6950  
TTTCCTGCGT TATCCCCTGA TTCTGTGGAT AACCGTATTA CCGCCTTTGA 7000  
40 GTGAGCTGAT ACCGCTCGCC GCAGCCGAAC GACCGAGCGC AGCGAGTCAG 7050  
TGAGCGAGGA AGCGGAAGAG CGCCCAATAC GCAAACCGCC TCTCCCGCG 7100  
45 CGTTGGCCGA TTCATTAATC CAGCTGGCAC GACAGGTTTC CCGACTGGAA 7150  
AGCGGGCAGT GAGCGCAACG CAATTAATGT GAGTTACCTC ACTCATTAGG 7200  
50 CACCCCAGGC TTTACACTTT ATGCTTCCGG CTCGTATGTT GTGTGGAATT 7250  
GTGAGCGGAT AACAAATTTCA CACAGGAAAC AGCTATGACC ATGATTACGA 7300  
55 ATTA 7305  
60

CLAIMS

1. A DNA construct comprising a transcriptional initiation site, a transcriptional termination site, a selectable gene, a product gene  
5 provided 3' to the selectable gene, a transcriptional regulatory region regulating transcription of both the selectable gene and the product gene, the selectable gene being positioned within an intron having a splice donor site 5' of the intron, which splice donor site regulates expression of the product gene using the transcriptional  
10 regulatory region.
2. The DNA construct of claim 1 wherein the splice donor site comprises an efficient splice donor sequence.
- 15 3. The DNA construct of claim 2 wherein the splice donor site comprises a consensus splice donor sequence.
4. The DNA construct of claim 2 wherein the splice donor site comprises the sequence GACGTAAGT.  
20
5. The DNA construct of claim 1 wherein the selectable gene is an amplifiable gene.
6. The DNA construct of claim 5 wherein the amplifiable gene is DHFR.  
25
7. The DNA construct of claim 1 wherein the transcriptional regulatory region comprises a promoter and an enhancer.
8. A vector comprising the DNA construct of claim 1.  
30
9. The vector of claim 8 wherein the selectable gene of the DNA construct is an amplifiable gene.
10. The vector of claim 8 that is capable of replication in a eukaryotic  
35 host.
11. A eukaryotic host cell comprising the vector of claim 10.
12. A eukaryotic host cell comprising the DNA construct of claim 5.  
40
13. The host cell of claim 11 wherein the vector is introduced into the host cell by electroporation.
14. A eukaryotic host cell comprising the DNA construct of claim 1  
45 integrated into a chromosome of the host cell.

15. The host cell of claim 14 that is a mammalian cell.
16. A method for producing a product of interest comprising culturing the  
host cell of claim 11 so as to express the product gene and  
recovering the product from the host cell culture.
17. The method of claim 16 further comprising recovering the product from  
the culture medium.
18. The method of claim 16 wherein the selectable gene is an amplifiable  
gene and the splice donor site comprises an efficient splice donor  
sequence.
19. A method for producing a product of interest comprising culturing the  
host cell of claim 12 so as to express the product gene in a  
selective medium comprising an amplifying agent for sufficient time  
to allow amplification to occur, and recovering the product.
20. A method for producing eukaryotic cells having multiple copies of a  
product gene comprising transforming eukaryotic cells with the DNA  
construct of claim 5, growing the cells in a selective medium  
comprising an amplifying agent for a sufficient time for  
amplification to occur, and selecting cells having multiple copies  
of the product gene.
21. The method of claim 20 further comprising recovering from the  
selected cells the product of interest.

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FIG. 1A

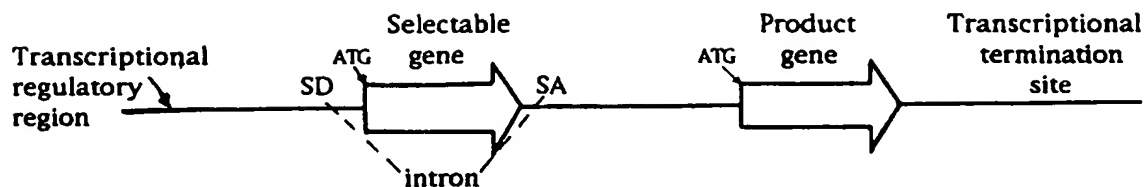


FIG. 1B

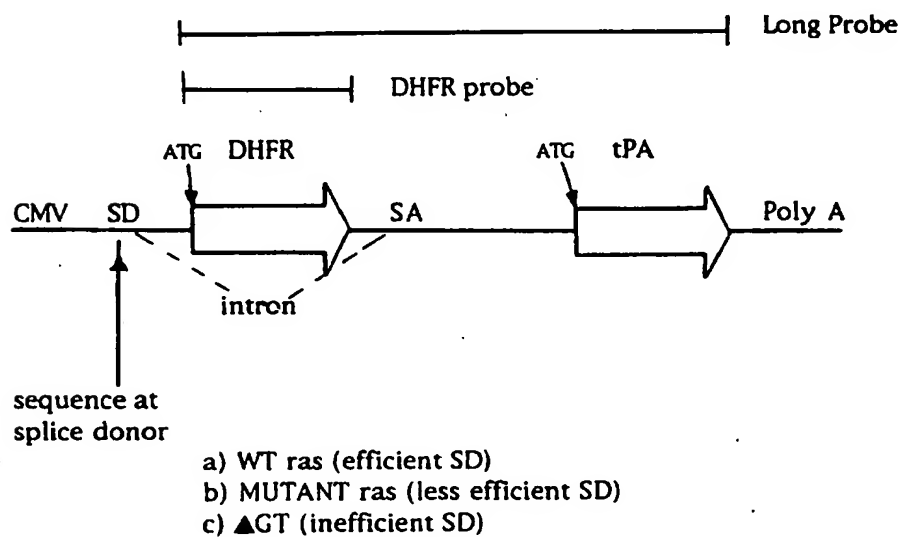
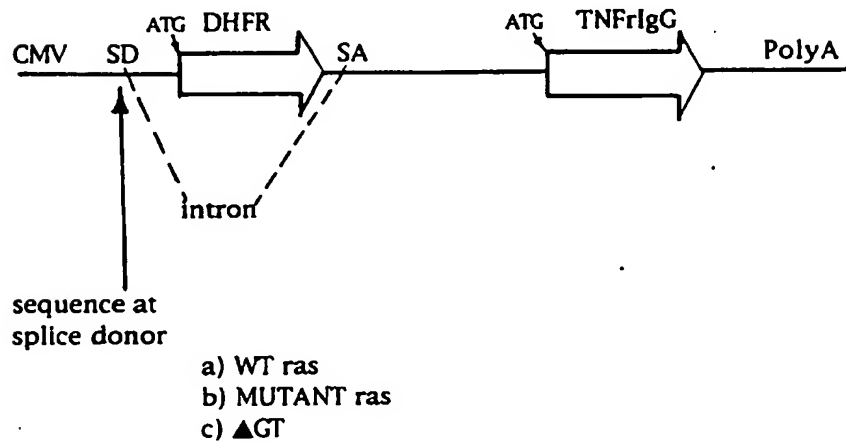


FIG. 1C





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FIG. 1D

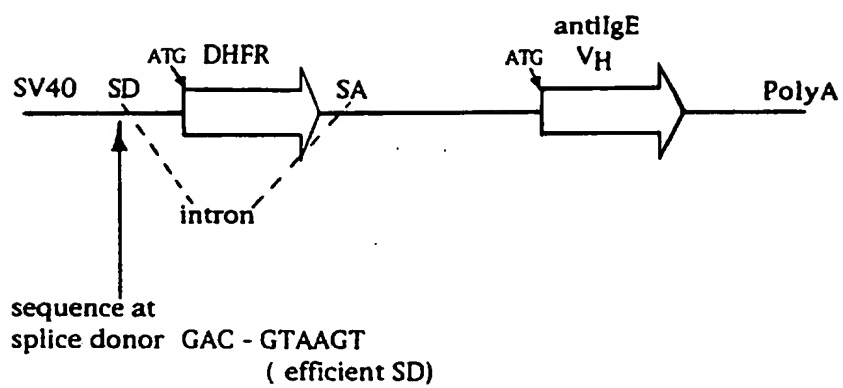
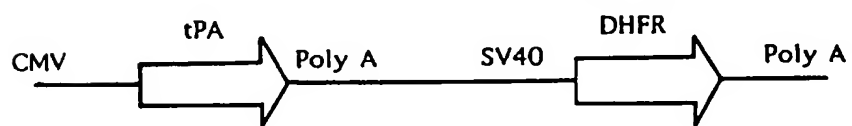


FIG. 2



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## FIG. 3A

```

alul
sstI
sacI
hgiIII
hgiAI/asphi
ecII36II
bsp1286
bsiHKA1
bmyI
banII
taqI
1 TTCGAGCTCG CCGACATTG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC
AAGCTCGAGC GGGCTGTAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CGGGTATATA CCTCAAGCGG CAATGTATTG
          rmaI   tru9I
          maeI   mseI
          speI   aseI/asnI/vspI
          bslI
          aciI maeIII
          bsh1236I
          fnuDII/mvnI
          bstUI
          chaI

          scrFI
          mvaI
          ecorII
          dsav
          aciI
          bglI bstNI
          sau96I
          haeIII/palI aciI
          asuI apyI(dcm+)
101 TTACGGTAAA TGGCCCGCCT GGCTGACCGC CCAACGACCC CCGCCCATG ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA
AATGCCATT ACCGGCGGA CCGACTGGC GGTGCTGGG GCGGGGTAAC TGCAGTTATT ACTGCATACA AGGTATCAT TCGGTTATC CTGAAAGGT
          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
          bglI
          rsaI   csp6I
          ndeI   csp6I
          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
201 TTGACGTCAA TGGTGGAGT ATTACGGTA AACTGCCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGAGGT CAATGACGGT
AAGTGCAGTT ACCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGGT TCATCGCGGG GATAACTGCA GTTACTGCCA
          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
          bglI
          rsaI   csp6I
          ndeI   csp6I
          maeII
          hinII/acyI
          ahaII/bsaHI
          aatII
          styI
          nlaIII
          ncoI
          dsaI hphI aciI
          bsaJI sfaNI
301 AAATGCCCG CCGGCATTA TGCCCGATAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATGC
TTTACCGGGC GGACCGTAAT ACGGTCATG TACTGGAATA CCCTGAAAGG ATGAACCGTC ATGTAGATGC ATAATCAGTA GCGATAATGG TACCACATCG
          scrFI
          mvaI
          ecorII
          aciI
          bglI dsav
          sau96I bstNI
          haeIII/palI
          asuI apyI(dcm+) bsrI nlaIII
          rsaI   csp6I
          maeII
          snaBI
          csp6I   bsaAI
          rsaI   csp6I
          dsaI hphI aciI
          bsaJI sfaNI

```

SUBSTITUTE SHEET (RULE 26)

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## FIG. 3C

```

      tfil
      acil
      thai hinfI
      fnuDII/mvnI
      bstUI
      bsh1236I
701 TTGGAACGCG GATTCGCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA CGGATAATCT CGCTATTCTC CTAAATAGG GCGACGGTA GTACCAAGCT GGTAACTTGA
      AACCTTGGC CTAAGGGCA CGGTTCTCAC GACATTCTATG GCGGATAATCT CGCTATTCTC CTAAATAGG GCGACGGTA GTACCAAGCT GGTAACTTGA
      fnu4HI
      bbvI
      nspBII
      acil
      nlaIII
      taqI
      fnuDII/mvnI
      bstUI
      bsh1236I
      mlui
      bsrBI
      acil
      xmnI
      rsal
      csp6I
      mnlI
      ddel asp700
      scaI
801 GCATCGTCG CGTGTCCTCA AATATGGGA TTGGCAAGAA CGGAGACCTA CCTGCGCTC CGCTCAGGAA CGCGTTCAAG TACTTCCAAA GAATGACCAC
      CGTAGCAGCG GCACAGGGT TTATACCCCT TTATACCCCT GCCTCTGGAT GGGACGGGAG GCGAGTCTCTT GCGCAAGTTC ATGAAGGTTT CTTACTGGTG
      scrFI
      mvaI
      ecorII
      dsav
      bstNI
      apyI[dcn+]
      sexAI
      tfil
      mboII
      taqI
      msel
      tru9I
      msel
901 AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAA CCTGGTTCTC CATTCCTGAG AAGATCGAC CTTTAAAGGA CAGAATTAAT
      TTGGAGAAGT CACCTTCCAT TTGTCITTAG CCACTAATAC CCATCCTTTT GGACCAAGAG GTAAGGACTC TTCTTAGCTG GAAATTTCTT GTCTTAATTA
      aluI
      sstI
      sacI
      hgiJII
      hgiAI/asphi
      ec1136II
      bsp1286
      bsiHKA
      bmyI
      banII
      bslI
      mnlI
1001 ATAGTTCTCA GTAGAGACT CAAGAACCA CCACGAGGAG CTCATTTTCT TGCCAAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG
      TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCTCT GAGTAAAGA ACGGTTTCA AACCTACTAC GGAATTTCTG ATAACCTTGT GGCCTTAACC
      ddel
      bstXI
      foki
      sfaNI
      msel
      tru9I
      mspI
      hpaII
      bsaBI

```

## FIG. 3D

[illegible]

FIG. 3E

1501 CACTATAGAA TAACATCCAC TTGCTCTTTC TCTCCACAGG TGTCACTCCA GGTCAACTGC ACCTCGGTTT TAAGCTTGGG CTGCAGGTGC CCGTGAATTT  
GTGATATCTT ATTGTAGGTG AAACGGAAAG AGAGGTGTCC ACAGTGTGCT CAGTGTGAGT TGGAGCCCAAG ATTGGAACCC GACGTCCAGC GGCACCTTAA

1601 AAGGGACGCT GTGAAGCAAT CATGGATGCA ATGAAGAGAG GGTCTGTCTG TGTGTCTGCTG CTGTGTGGAG CAGTCTTCTG TTGCCCCAGC CAGGAATCC  
TTCCCTGCGA CACTTCGTTA GTACCTACGT TACTTCTCTC CCGAGACGAC ACACGACCTC GACACACCTC GTCAGAAGCA AAGCGGTGC GTCTTTTAGG

1701 ATGCCCGATT CAGACAGGA GCCAGATCTT ACCAGTGTAT CTGCAGAGAT GAAAAACGC AGATGATATA CCAGCAACAT CAGTCATGGC TGCGCCCTGT  
TAGCGGGCTAA GTCTTCTCCT CGGTCTAGAA TGGTCTACTA GACGTCTCTA CTTTGTGCG TCTACTATAT GGTCGTGTA GTCAGTACCG ACGCGGGACA

scfI mvaI ecorII dsav bstNI apyI[dcM+] mnII bsaJI hincII/hindII maeIII hincII/hindII gsuI/bpmI foki nlaIII hgaI

bspMI scfI pstI aluI hindIII fnu4HI bbsI mboII bpuAI bbsI

scrFI mvaI ecorII dsav bstNI apyI[dcM+] nlaIII

hgaI/aspHI bsp1286 bsiHKA I bmyI nlaIII hhaI/cfoI

hinfI mboII bglII dnfI[dam+] dnfII[dam+] mboI/ndeII[dam-] sau3AI mboI/ndeII[dam-] bsgI

nlaIV dnfI[dam+] bstYI/xhoII dnfII[dam-] dnfIII[dam-] mboII

sspi dsav cauli

ncII mspI hpaII

alwNI ddeI draIII bmyI bsp1286

styI bsaJI

truuI msel apoI

1801 GCTCAGAAGC AACCGGGTGG AATATTGCTG GTGCAACAGT GCAGGGGCAC AGTGCCACTC AGTGCCCTGTC AAAAGTTGCA GCGAGCCCAAG GTGTTTCAAC  
CGAGTCTTCG TTGGCCCCACC TTATAACGAC CAGTTGTCA CCGTCCCGTG TCACGGGTGAG TCACGGACAG TTTTCAACGT CGCTCGGTTC CACAAAGTTG

**FIG. 3**

**FIG. 3F**

bspMI	sau96I	pmlI
nlaIV	haeIII/palI	eco72I
hgICl	asuI	sau96I
bani	xsaI	asuI maeII
bsp1286	ecoO109I/draII	scrFI
bmyI	alwNI csp6I ddeI	mvaI haeIII/palI
GGGGGCACCT	GCCAGCAGGC CTTGTACTTC TCAGATTTCG CCCGAAGGA TTTGCTGGG AGTCTCTGGA AATAGATACC AGGCCACGT	ecoRII
CCCCCGTGGA	CGGTCTCCG GGACATGAAG AGTCTAAAGC ACACGGTCAC GGGGCTTCCT AAACGACCCT TCACGACACT TTATCTATGG TCCCGTGCA	dsav
		bstNI bsaAI
		bsaJI bbrPI
		apyI(dcm+)

[illegible]

**FIG. 3G**

FIG. 3G

2201 fnu4HI  
bbvI  
scfI  
pstI  
bsgI  
rsal  
alul  
asp6I ddel  
aciI  
AAGCGGGGA AGTACAGCTC AGAGTTCTGC AGCACCCTG CCGTCTCTGA GGGAAACAGT GACTGCTACT TTGGGAATGG GTACGCCTAC CGTGGCAGCG  
TTCCGCCCTT TCATGTCGAG TCTCAAGACG TCGTGGGGAC GGACGAGACT CCCTTTGTCA CTGACGATGA AACCTTACC CAGTCGGATG GCACCGTGGC  
scrFI  
pfIMI  
mvaI  
ecorII  
dsav  
bstNI  
bslI  
apyl[dcml+] haelIII/palI  
bsp1286 sau96I  
bmyI alwNI asuI  
bsrI bsaJI bsrI  
2301 ACAGCCTCAC CGAGTCGGGT GCCTCCTGCC TCCCGTGGAA TTCCATGATC CTGATATAGGA AGGTTTACAC AGCAGAGAAC CCCAGTGCCC AGGCACCTGGG  
TGTCGGAGTG GCTCAGCCCCA CGGAGGACGG AGGCACCTT AAGGTACTAG GACTATCCGT TCCAAATGTG TCGTGTCTTG GGGTCACGGG TCGGTGACCC  
pleI nlaIV  
hphi hinfI hgiCI  
mnli bcrI bani mnli mnli bsaJI nlaIII  
ecorI  
dsal apoI  
2401 fnu4HI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII  
sfanI  
mnli sapi  
fnu4HI bsmAI  
aciI ddel  
rsal  
bspMI haelIII/palI csp6I  
CCTCTCTGCT CCACCTGCGG CCTGAGACAG TACAGCCAGC CTCAGTTTCG CATCAAAGGA GGGCTCTTCG CCGACATCGC CTCCCACCCC TGGCAGGCTG  
GGGAGGACGA GGTGGACGCC GGACTCTGTC ATGTGGGTGC GAGTCAAAGC GTAGTTTCT CCCGAGAAGC GGCTGTAGCG GAGGGTGGG ACCGTCCGAC  
scrFI  
mvaI  
ecorII  
dsav  
bstNI  
bsaJI  
apyl[dcml+]  
CGACCCGTTT GTATTAATGA CGGCCCTTAGG ACTACCCCTA CCGTTCGGGA CCACGGTGCA CGACTTCTTG GCGTCCGACT GCACCCCTCAT GACACTACAC  
mboII  
earI/ksp632I  
hgiJII  
bsp1286  
bmyI  
banII





**FIG. 31**

FIG. 3I

2901

3001

3101

11 / 81

FIG. 3J

```
scrFI      mspI      hpaII      xmaI/pspAI      smaI      scrFI      nciI      nlaIV      ecoO109I/draII      aluI sau96I      pvuII asuI      sfaNI      nspBII haeIII/palI      fokiI      avai      bspI407I      bstEII      fmaI      hinfI      nsfPI      nspHI      maeIII nlaIII      CAACTACCTA GACTGGATTG GTGACAACAT      ACCACCCGTA GTAGTCGACC CCGGACCCGA CACCTGTCTT CCTACAGGCG CCACACATGT GGTTCGAATG GTTGATGGAT CTGACCTAAG CACTGTGTGA      3201 TGGTGGGCAT CATCAGCTGG GGCCTGGGCT GTGGACAGAA GGATGTCGCG GGTTGTGTACA CCAAGGTTC      3301 GCGACCGTGA CCAGGAACAC CCGACTCCTC AAAAGCAAT GAGATCCCGC CTCTCTCTCT TCAGAGACA CTGCAAGGC GCAGTCTTC TCTACAGACT      CGCTGGCACT GGTCTTG TGCTGAGGAG TTTTCGTTA CTCTAGGCGG GAGAAGAAGA AGTCTCTGT GACGTTTCCG CGTCACGAAG AGATGTCTGA      3401 TCTCCAGACC CACCACACC CAGAAGCGG ACAGACCT ACAGGAGAGG GAAGATGCA TTTTCCAG TACTTCCCAT TTTGGAAGTT TTCAGGACTT      AGAGGTCTGG GTGGTGTCG GTCTTGCCC TGCTCTGGA TGCTCTCTCC CTCTCTACGT AAAAGGCTCT ATGAAGGTA AAACCTTCAA AAGTCTGAA      tthlIII/aspI      3501 GGTCTGATTT CAGGATACTC TGTCAGATGG GAAGACATGA ATGCACACTA GCCTCTCCAG GAATGCTCC TCCCTGGCA GAAAGTGGGG GAATTCATC      CCAGACTAAA GTCCTATGAG ACAGTCTACC CTCTCTGACT TACGTGTGAT CGGAGAGGTC CTTACGAGG AGGACCCGT CTTACACCCC CTTAAGTTAG      apoI      clalI/bsp106      ecoRI      taqI
```

**FIG. 3K**

SUBSTITUTE SHEET (RULE 26)

[illegible]

## FIG. 3M

hinPI  
 hhaI/cfoI  
 nlaIV  
 nari  
 kasi  
 hinII/acyI  
 hgiCI  
 haeII  
 bani  
 ahaII/bsaHI  
 bglI  
 4301 CGAAGAGGCC CGCACCAGATC GCCCTTCCCA ACAGTTGCGT AGCCTGAATG GCGAATGCGC CCTGATGCGG TATTTCTCC TTACGCATCT GTGCGGTATT  
 GCTTCTCCGG CGGTGGCTAG CGGGAAGGCT TGTCAACGCA TCGGACTTAC CGCTTACCGC GGAATAGAGG AATAAGAGG AATGCTAGA CAGCCATAA  
 sau3AI  
 sau96I mboI/ndeII(dam-)  
 haeII/palI  
 asuI  
 mnlI  
 mboII aciI pvuI/bspCI  
 earI/ksp63II mcrI  
 4401 TCACACCGCA TAGCTCAAAG CAACCATAGT ACGCGCCCTG TAGCGCGCA TTAAGCGCG CGGTGCTGGT GGTTACGCGC AGCGTGACCG CTACACTTGC  
 AGTGTCGCGT ATGCAGTTTC GTTGGTATCA TCGCGGGGAC ATCGCGCGCT AATCGCGCT GCGCACACCA CCAATGCGG TCGCACTGCG GATGTGAACG  
 aciI maeII  
 hinPI  
 hhaI/cfoI  
 rnaI  
 hinPI haeII  
 hhaI/cfoI bsrBI  
 haeII maeI aciI  
 4501 CAGCGCCCTA GCGCCGCTC CTTTCCTTCC TTTCTCGCCA CGTTCCGCGG CTTTCCCGGT CAAGCTCTAA ATCGGGGCT CCCTTTAGGG  
 GTCGCGGAT CGCGGCGAG GAAAGCGAA GAAGGGAAGG AAGAGCGGT GCAAGCGCT GCAAGCGCC GAAAGCGCA GTTCGAGAT TAGCCCCGA GCGAAATCCC  
 nlaIV  
 hgiCI taqI  
 bani mnlI  
 4601 TTCCGATTTA GTGCTTTACG GCACCTCGAC CCCAAAAC TTGATTTGGG TGATGTTCA CGTAGTGGC CATCGCCCTG ATAGACGTT TTGCGCCCTT  
 AAGGCTAAT CACGAAATGC CGTGGAGCTG GGGTTTTTG AACTAAACC ACTACCAAGT GCATCACCG GCATCGGAC TATCTGCAA AAGCGGGA  
 maeII pleI  
 drdI hinfI maeII  
 4701 TGACGTGGA GTCCAGTTT TTTAATAGTG GACTCTTGT CCAAACTGA ACAACTCA ACCCTATCT GGGCTATTCT TTGATTTAT AAGGATTTT  
 ACTGCAACCT CAGGTGCAAG AAATTATCAC CTGAGAACA GGTGTGACCT TGTGTGAGT TGGATAGAG CCCGATAAGA AAATAAATA TTCCCTAAA

[illegible]

FIG. 30

```

nlaIV
aciI
thai
fnuDII/mvnl
bstUI
bsh1236I
hinPI
hhaI/cfoI

5201 CTTTTCGGG AAATGTGCGC GGAACCCCTA TTTGTTTATT TTTCTAATA CATTCAATA TGTATCCGCT CATGAGACAA TAACCTTGAT AAATGCTTCA
GAAAAGCCCC TTTACAGCGC CCTTGGGGAT AAACAATAA AAAGATTTAT GTAAAGTTTAT ACATAGCGCA GTACTCTGTT ATTGGGACTA TTTACGAAGT

mboII
earI/ksp632I
sspI
55301 ATAATATTGA AAAAGGAAGA GTATCAGTAT TCAACATTTC CGTGTCGCCC TTATTCCCTT TTTTGCGGCA TTTTGCCCTC CTGTTTTTGC TCACCCAGAA
TATTATAACT TTTTCTCTCT CATACTCATA AGTGTAAAG GCACAGCGG AATAAGCGAA AAAACGCCGT AAAACGGAAG GACAAAAACG AGTGGTCTT

hgiAI/aspHI
bsp1286
sau3AI
mboI/ndeII[dam-]
dpmI[dam+] bmyI
dpmII[dam-]
eco57I
apalI/snoI
hphI
sfaNI mboII[dam-] alw44I/snoI maeIII taqI alwI[dam-] aciI bstYI/xhoII
5401 ACCTGTGTA AGTAAAGA TGCTGAAGAT CAGTTGGTG CACGAGTGG TTACATCGAA CTGATCTCA ACAGCGGTAA GATCCTTGAG AGTTTTGCGC
TGGGACCACT TTCATTTTCT ACGACTTCTA GTCACCCAC GTGCTCACCC AATGTAGCTT GACCTAGAGT TGTGCGCAAT CTAGGAACCTC TCAAAAGCGG

maeII
psp1406I
xmnI
asp700
mboII
5501 CCGAAGACG TTTTCCAATG ATGAGCACTT TTAAGTTCT GCTATGTGCG GCGGTATTAT CCGGTGATGA CGCGCGGCAA GAGCAACTCG GTCGCGCAT
GGCTTCTTGC AAAAGGTTAC TACTCGTGAA AATTTCAAGA CGATACACCG CGCCATAATA GGGCACTACT GCGGCCCGTT CTCGTTGAGC CAGCGCGCTA

rcal
bspHI
bsrBI bsmAI
aciI nlaIII
fnu4HI
aciI
sau3AI nspBII
sau3AI mboI/ndeII[dam-]
dpmI[dam+]
bstYI/xhoII
bsrI dpmII[dam-]
alwI[dam-]
aciI bstYI/xhoII
scrFI
nciI
mspI
hpaII
dsv
hinII/acyI
hgaI caulI
hhaII/bsaHI
bcgI mcrI fnu4HI
aciI
fnu4HI
bbvI
nlaIII
fnu4HI

```



FIG. 3P

5701 ATGAGTGATA ACACTGCGC CAACCTACTT CTGACAACGA TCGGAGGACC GAAGGAGCTA ACCGCTTTT TGCACAACAT GGGGGATCAT GTAACCTCGCC  
TACTCACTAT TGTGAGCGCG GTTGAATGAA GACTGTGTCT AGCCTCTCTG CTCTCTCGAT TGGCGAATAA ACGTCTGTGA CCCCTTAGTA CATGAGCGCG

5801 TTGATCGTTG GGAACCGGAG CTGAATGAAG CCATACCAAA CGACGAGCGT GACACACGA TGCCAGCAGC AATGGCAACA AGCTTGGCA AACTATTAAAC  
AACTAGCAAC CCTTGGCCTC GACTTACTTC GGTATGGTTT GCTGCTCGCA CTGTGTGCT ACGTCTGCTG TTACCGTTGT TGAACCGGT TTGATAATTG

5901 TGGCGAACTA CTTACTCTAG CTTCCCGGCA ACAATTATA GACTGGATGG AGGCGGATAA AGTTGCAGGA CCACCTCTGC GCTCGGCCCT TCCGGCTGGC  
ACCGCTTGAT GAATGAGATG GAAGGGCCGT TGTAAATTAT CTGACCTACC TCCGCTATT TCAACGTCTT GGTGAAGACG CGAGCCGGA AGGCCGACCG

6001 TGGTTTATTG CTGATAAATC TGGAGCCGGT GAGCGTGGGT CTCGCGGTAT CATTCAGCA CTGGGGCCAG ATGGTAAGCC CTCCCGTATC GTAGTTATCT  
ACCAATAAC GACTATTATG ACCTCGGCA CTGCGACCCA GAGCGCCATA GTAACGTCTG TACCCTCGT GAGGCGATAG CATCAATAGA

6101 ACACGACGGG GAGTCAGGCA ACTATGGAT AACGAAATAG ACAGATCGT GAGATAGGTG CCTCACTGAT TAAGCATTTG TAACTGTGAC ACCAAGTTTA  
TGTGCTGCC CTAAGTCCGT TGATACCTAC TTGCTTTATC TGTCTAGGCA CTCTATCCAC GGAGTACTA ATTGTAACC ATTGACAGTC TGGTTCAAT

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 sau96I  
5801 sau3AI  
5901 sau3AI  
6001 sau3AI  
6101 sau3AI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001 pnuI/bspCI  
6101 pnuI/bspCI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mcrI mnlI  
5801 mcrI mnlI  
5901 mcrI mnlI  
6001 mcrI mnlI  
6101 mcrI mnlI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001 pnuI/bspCI  
6101 pnuI/bspCI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 mcrI mnlI  
5801 mcrI mnlI  
5901 mcrI mnlI  
6001 mcrI mnlI  
6101 mcrI mnlI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001 pnuI/bspCI  
6101 pnuI/bspCI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mcrI mnlI  
5801 mcrI mnlI  
5901 mcrI mnlI  
6001 mcrI mnlI  
6101 mcrI mnlI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001 pnuI/bspCI  
6101 pnuI/bspCI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 mcrI mnlI  
5801 mcrI mnlI  
5901 mcrI mnlI  
6001 mcrI mnlI  
6101 mcrI mnlI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001 pnuI/bspCI  
6101 pnuI/bspCI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mcrI mnlI  
5801 mcrI mnlI  
5901 mcrI mnlI  
6001 mcrI mnlI  
6101 mcrI mnlI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001 pnuI/bspCI  
6101 pnuI/bspCI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 mcrI mnlI  
5801 mcrI mnlI  
5901 mcrI mnlI  
6001 mcrI mnlI  
6101 mcrI mnlI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001 pnuI/bspCI  
6101 pnuI/bspCI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mcrI mnlI  
5801 mcrI mnlI  
5901 mcrI mnlI  
6001 mcrI mnlI  
6101 mcrI mnlI

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 haeIII/palI  
5801 haeIII/palI  
5901 haeIII/palI  
6001 haeIII/palI  
6101 haeIII/palI

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 mboI/ndeII[dam-]  
5801 mboI/ndeII[dam-]  
5901 mboI/ndeII[dam-]  
6001 mboI/ndeII[dam-]  
6101 mboI/ndeII[dam-]

Genetic markers and their positions are indicated on the right side of the sequence blocks:

5701 dpmI[dam+]  
5801 dpmI[dam+]  
5901 dpmI[dam+]  
6001 dpmI[dam+]  
6101 dpmI[dam+]

Genetic markers and their positions are indicated on the left side of the sequence blocks:

5701 pnuI/bspCI  
5801 pnuI/bspCI  
5901 pnuI/bspCI  
6001

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## FIG. 3Q

```

        rmaI      sau3AI
sau3AI hphI      mboI/ndeII[dam-]
mboI/ndeII[dam-]
dpmI[dam+]      dpmI[dam+]
dpmII[dam+]      dpmII[dam-]
tru9I bstYI/xhoII alwI[dam-]      nlaIII      maeII
mseI      tru9I mseI alwI[dam-] bstYI/xhoII      rcaI      tru9I
ahaIII/draI mseI ahaIII/draI maeI mboII[dam-]      bspHI      mseI
6201 CTCATATATA CTTTAGATTG ATTTAAAGGA TCTAGGTGAA GATCCTTTT GATAATCTCA TGACCAAAAT CCCTTAACGT
GAGTATATAT GAAATCTAC TAAATTTTCT AGATCCACTT CTAGGAAAAA CTAATTAGAGT ACTGGTTTGA GGGAAATTGCA

        sau3AI
mboI/ndeII[dam-]
dpmI[dam+]      sau3AI
dpmII[dam-] mboI/ndeII[dam-]      thai
bstYI/xhoII dpmI[dam+]      fnuDII/mvnI
sau3AI alwI[dam-] dpmII[dam-]      bstUI
mboI/ndeII[dam-] alwI[dam-]      bsh1236I
dpmI[dam+] mboII[dam-]      hinPI      fnu4HI
dpmII[dam-] bstYI/xhoII      hhai/cfoI      bbvI
6301 GAGTTTTCGT TCCACTGAGC GTCAGACCCC GTAGAAAAGA TCAAGGATC TTCTTGAGAT CCTTTTTC TGCGCGTAAT CTGCTGCTTG CAAACAAAAA
CTCAAAAAGCA AGGTGACTCG CAGTCTGGGG CATCTTTTCT AGTTTCCTAG AAGAACTCTA GGAAAAAAG ACGCCGATTA GACGACGAAC GTTTGTTTTT

        sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam+]
alwI[dam-]
mspI
aciI      nspBII      hpaII      aluI      bsrI      hinPI
aciI      AACCAACCGT ACCAGCGGTG GTTTGTTTGC CGGATCAAGA GCTACCAACT CTTTTTCCGA AGGTAACCTGG CTTTCAGCAGA GCGCAGATAC CAAATFACTGT
6401 TTGGTGGCGA TGGTCGCCAC CAAACAAACG GCTAGTTCT CGATGTTGA GAAAAAGGCT TCCATTGACC GAAGTCGTCT CGCGTCTATG GTTTATGACA

        haeIII/palI
rmaI      haeI      bslI      maeI      scfI      aciI      mnlI      bsrI      fnu4HI
maeI      TAGCCACCA CTTCAAGAAC TCTGTAGCAC CGCCTACATA CCTCGCTCTG CTAATCTCTGT TACCAGTGGC TGCTGCCAGT
6501 CCTTCTAGTG TAGCCGATAG ATCCGGTGGT GAATTTCTTG AGACATCGTG GCGGATGTAT GGAGCGAGAC GATTAGGACA ATGTCACCG ACGACGGTCA
GGAAGATCAC ATCGGCATCA

```

[illegible]

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## FIG. 3S

```

          thai
          fnuDII/mvnI
          bstUI
          bsh1236I
          hinPI
          hhaI/cfoI
          thai
          fnuDII/mvnI
          bstUI haeIII/palI          aluI
          bsh1236I          tru9I pvuII
          bslI eaeI tfiI aseI/asnI/vspI
          aciI cfrI hinfI mseI nspBII          bsrI
          CCGCGCGGTG GCGGCGTTCAT TAATCCAGCT GGCACGACAG GTTCCCGAC
          CAGTCACTCG CTCCTTCGCC TTCTCGCGG TTATCGGTTT GCGGAGAGG GCGCGCAAC CGGCTAAGTA ATTAGGTGA CCGTGCTGTC CAAAGGGCTG

          sapi hinPI
          mboII hhaI/cfoI
          earI/ksp632I
          mnlI aciI haeII
          GAGGAAGCGG AAGAGCGCCC AATACGCAAA CCGCCTCTCC
          CAGTCACTCG CTCCTTCGCC TTCTCGCGG TTATCGGTTT GCGGAGAGG GCGCGCAAC CGGCTAAGTA ATTAGGTGA CCGTGCTGTC CAAAGGGCTG

          scrFI
          mvai
          ecorII
          dsav
          nlaIV bstNI
          hgICl apyI[dcM+]
          bani bsaJI
          TTAGGCACCC CAGGCTTTAC ACTTTATGCT TCCGGCTCGT ATGTTGTGTG
          AATCCGTGGG GTCCGAAATG TGAATACGA AGGCCGAGCA TACAACACAC
          tru9I
          mseI
          aseI/asnI/vspI
          xmnI
          asp700
          aluI nlaIII
          GAAACAGCTA TGACCATGAT TACGAATTAA
          CTTTGTGAT ACTGGTACTA ATGCTTAAT

          aciI
          bsrBI
          TTTACACAG GAAACAGCTA TGACCATGAT TACGAATTAA
          GAAACAGCTA TTTACACAG GAAACAGCTA TGACCATGAT TACGAATTAA
          CTTTGTGAT ACTGGTACTA ATGCTTAAT

          >length: 7360

```

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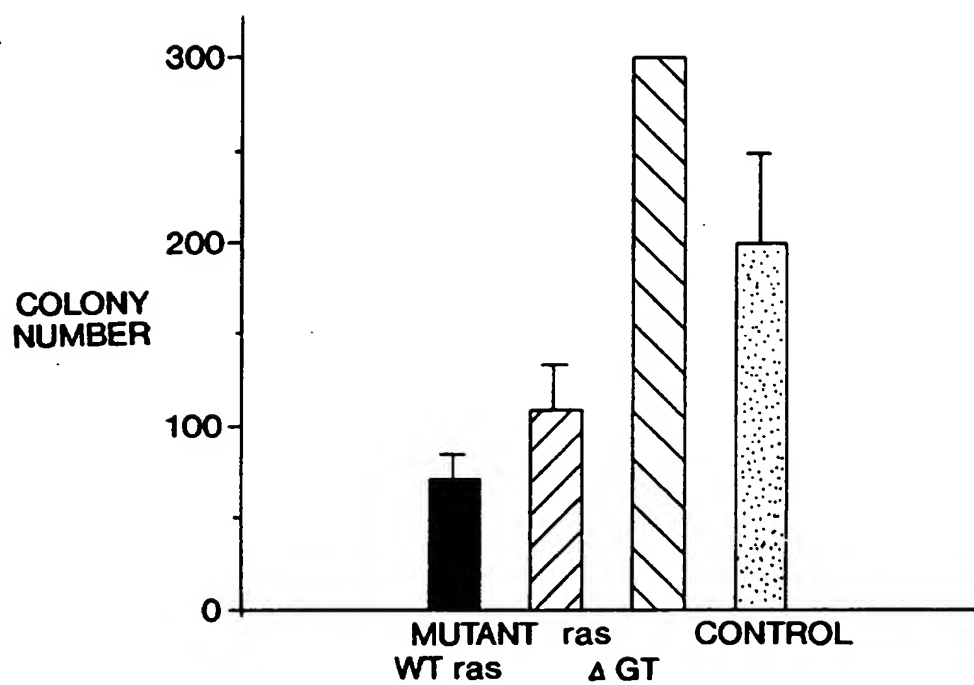


FIG. 4

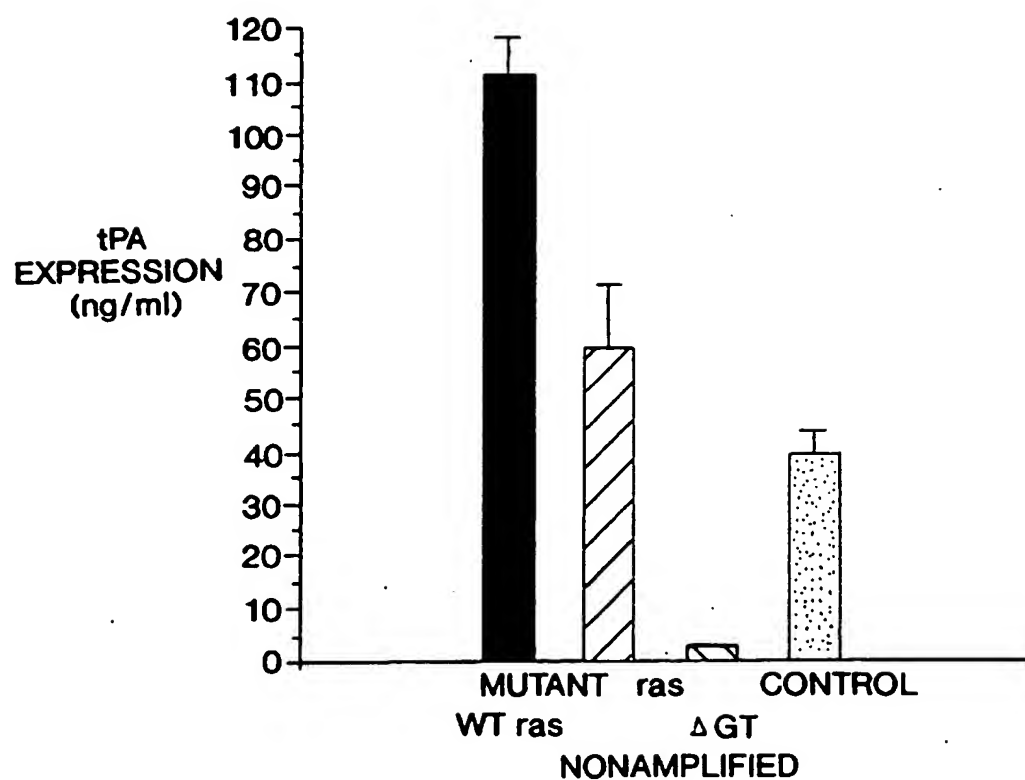


FIG. 5A

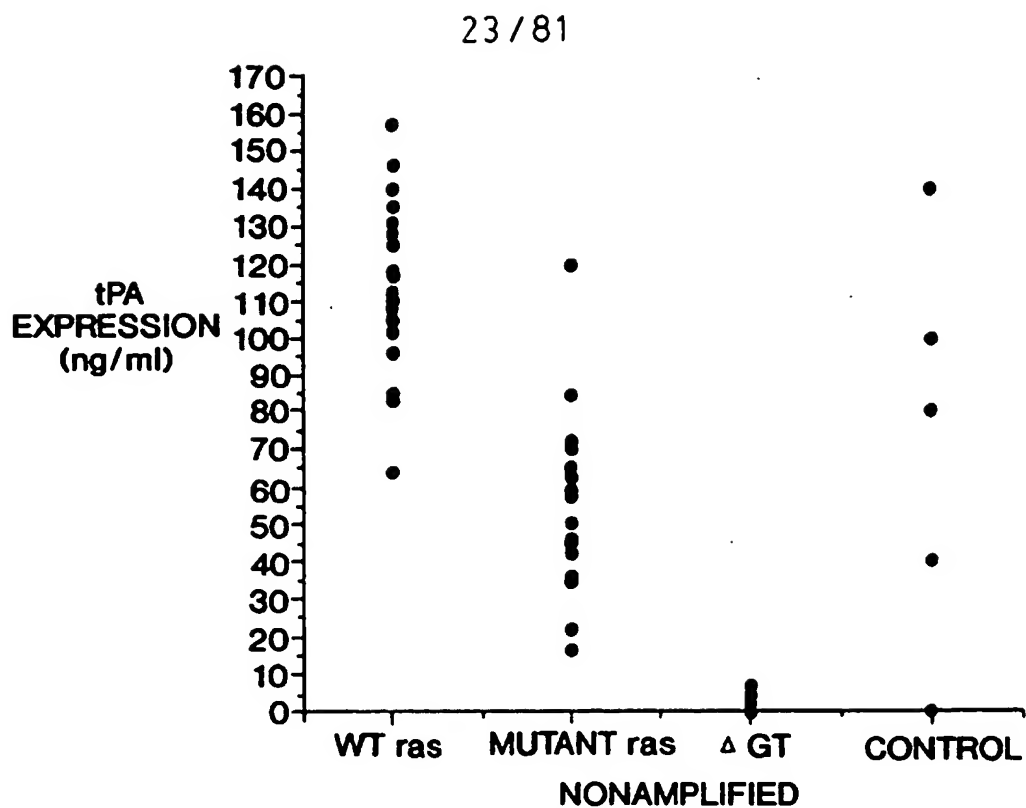


FIG. 5B

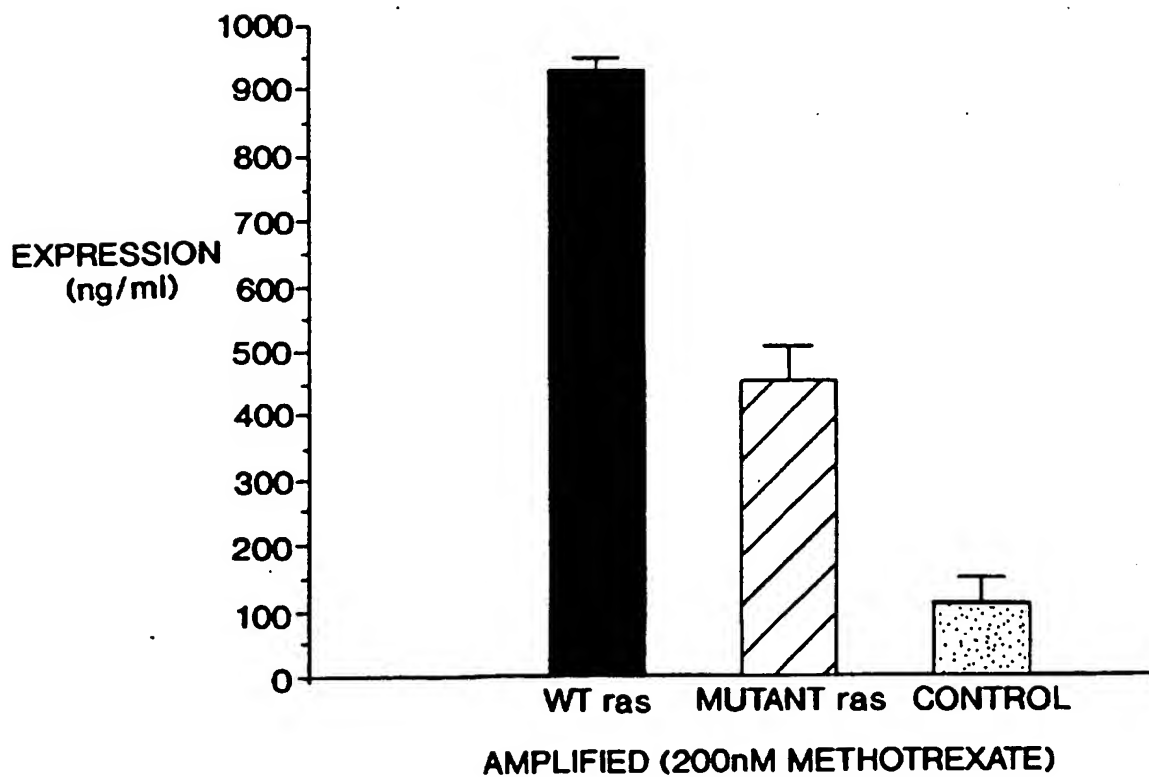


FIG. 5C

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## FIG. 6A

```

1  TTCCAGCTCG CCGACATTG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC
   AAGCTCGAGC GGGCTGTAAC TAATAACTGA TCAATTAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CCGGTATATA CCTCAAGGCG CAATGTATTG
   taqI                                     rmaI   tru9I   maeI   speI   asel/asni/vspI   bslI   aciI   maeIII
   sstI                                     maeI                                     bsh1236I
   sacI                                     maeI                                     bstUI
   hgiJII                                  fnuDII/mvnI
   hgiAI/aspHI
   ecl136II
   bsp1286
   bsiHKA1
   bmyI
   banII
   scrFI
   mvaI
   ecorII
   dsav
   aciI
   bglI bstNI
   sau96I
   haeIII/palI   aciI
   asuI apyI(dcm+)
   maeII
   hinII/acyI
   ahaII/bsaHI
   aatII
   maeIII
   101 TTACGGTAA TGGCCCGCT GCGTACCGC CCAACGACCC CCGCCCATTTG AGTCAATTA TGACGTATGT TCCCATAGTA ACGCCAATAG GGACTTTCCA
   AATGCCATTT ACCGGCGGA CCGACTGGCG GGTGTCTGGG GCGCGGTAAC TGCAGTTATT ACTGCATACA AGGTATCAT TCGGTTATC CTGAAAGGT
   maeII
   hinII/acyI
   ahaII/bsaHI
   aatII
   rsaI
   csp6I
   bglI
   ndeI
   maeII
   201 TTGACGTCAA TGGGTGGAGT ATTTACGGTA AACTGCCAC TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCT CTATTGACGT CAATGACGGT
   AACTGCAGTT ACCACCTCA TAATGCCAT TTGACGGGTG AACCGTCATG TAGTTCACAT AGTATACGGT TCATGGGGG GATAACTGCA GTTACTGCCA
   scrFI
   mvaI
   ecorII
   aciI
   bglI dsav
   sau96I bstNI
   haeIII/palI   rsaI
   asuI apyI(dcm+)   bsrI nlaIII
   maeII
   hinII/acyI
   ahaII/bsaHI
   aatII
   rsaI
   csp6I
   bglI
   ndeI
   maeII
   301 AAATGGCCG CCTGGCATT TGCACAGTAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTATGC
   TTTACCGGCG GACCGTAAT ACGGTCATG TACTGGATA CCCTGAAAGG ATGAACCGTC ATGTAGTGC ATATACAGTA GCGATAATGG TACCACCTAGC
   nlaIII
   styI
   ncoI
   dsaI hphI aciI
   bsaJI   sfaNI

```

**FIG. 6B**

401	GGTTTTGGCA	GTACATCAAT	GGCGTGGAT	AGCGGTTTGA	CTCACGGGA	TTTCCAAGTC	TCCACCCAT	TGAGCTCAAT	GGGAGTTTGT	TTTGGCACCA
	CCAAAACCGT	CATGTAGTTA	CCGCACCTA	TCGCCAACT	GAGTCCCT	AAAGTTTCA	AGTGGGTA	ACTGCAGTTA	CCCTCAAACA	AAACCGTGGT
	rsal	pleI	aciI	hinfi	bsmAI					
	mspI									
	alul									
	sstI									
	sacI									
	hgiI									
	hgiAI/aspHI									
	ecII36II									
	bspI286									
	bsiHKAII									
	bmyI									
	banII									
	maeII									
	hinII/acyI									
	ahaII/bsaHI									
	aatII									
	rsal									
	mspI									
	alul									
	sstI									
	sacI									
	hgiI									
	hgiAI/aspHI									
	ecII36II									
	bspI286									
	bsiHKAII									
	bmyI									
	banII									
	maeII									
	hinII/acyI									
	ahaII/bsaHI									
	aatII									
	rsal									
	mspI									
	alul									
	sstI									
	sacI									
	hgiI									
	hgiAI/aspHI									
	ecII36II									
	bspI286									
	bsiHKAII									
	bmyI									
	banII									
	maeII									
	hinII/acyI									
	ahaII/bsaHI									
	aatII									
	rsal									
	mspI									
	alul									
	sstI									
	sacI									
	hgiI									
	hgiAI/aspHI									
	ecII36II									
	bspI286									
	bsiHKAII									
	bmyI									
	banII									
	maeII									
	hinII/acyI									
	ahaII/bsaHI									
	aatII									
	rsal									
	mspI									
	alul									
	sstI									
	sacI									
	hgiI									
	hgiAI/aspHI									
	ecII36II									
	bspI286									
	bsiHKAII									



## FIG. 6C

tfii  
 acii  
 thai hinfI  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 701 TTGGAACGCG GATTCGCCGT GCCAAGAGTG CTGTAAGTAC CGCCTATAGA CGGCTATCC GATTTATCC CGCTGCCAT CATGGTTCGA CCATTGAACT  
 AACCTTGCGC CTAAGGGCA CGGTCTCTAC GACATTCATG CGGATATCT CGCTATTCTC CTAATAATAGG CCGCTACGTA GTACCAAGCT GGTAACTTGA  
 fnu4HI  
 bbvI  
 nspBII  
 acii  
 nlaIII  
 taqI  
 thai  
 fnuDII/mvni  
 bstUI  
 bsh1236I  
 mluI  
 bsrBI  
 acii  
 xmiI  
 rsai  
 csp6I  
 801 GCATCGTCCG CGTGTCCTCA AATATGGGGA TTGGCAAGAA CGGAGACCTA CCCTGCCCTC CGCTCAGGAA CGGTTTCAAG TACTTCCAA GAATGACCAC  
 CGTAGCAGCG GCACAGGCTT TTATACCCCT AACCGTCTT GCCTCTGGAT GGGACGGGAG CGGAGTCTT CGGCAAGTTC ATGAAGCTT CTTACTGGTG  
 pflMI  
 bslI  
 bsmAI  
 bsai  
 mnlI  
 ddel  
 asp700  
 scaI  
 scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 apyI(dcm+)  
 sexAI  
 901 AACCTCTTCA GTGGAAGGTA AACAGAATCT GGTGATTATG GGTAGGAAA CCTGGTTCTC CATTCCTGAG AAGATCGAC CTTTAAAGGA CAGAATTAAT  
 TTGGAGAAGT CACCTTCCAT TTGTCTTAGA CCACCTAATAC CCATCTTTT GGACCAAGAG GTAGGACTC TTCTTAGCTG GAATTTCTT GTCTTAATTA  
 tfil  
 hinfI  
 alwNI  
 hphI  
 tfil  
 mboII  
 taqi  
 msei  
 tru9I  
 msei  
 ahaIII/draI  
 asel/asnI/vspI  
 aluI  
 sstI  
 sacI  
 hgiJII  
 hgiAI/aspHI  
 ecl136II  
 bsp1286  
 bsiHKA  
 bmyI  
 banII  
 1001 ATAGTTCTCA GTAGAGAAGT CAAAGAACCA CCACGAGGAG CTCATTTTCT TGCCAAAGT TTGGATGATG CCTTAAGACT TATTGAACAA CCGGAATTGG  
 TATCAAGAGT CATCTCTTGA GTTCTTGGT GGTGCTCTC GAGTAAAGA ACGGTTTCA AACCTACTAC GGAATTTCTG ATAATTGTT GGCCTTAACC  
 ddel  
 bslI  
 mnlI  
 bstXI  
 foki  
 sfaNI  
 msei  
 afliI/bfri  
 bsaNI  
 mspI  
 hpaII

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## FIG. 6D

1101 CAAGTAAAGT AGACATGGTT TGGATAGTCG GAGGCAGTTC TGTATTACCAAG ACCTATCAGC CTCCGTCAAG ACAATGGTC CTCCGGTACT TAGTGTGTCG GGTGGATCT GAGAACACT GTTCCTAGTA  
 accI nlaIII mnlI haeIII/palI haeI  
 scrFI mvaI mvaI mvaI  
 ecorII ecorII ecorII  
 dsav tfII dsav pleI  
 bstNI nlaIII bstNI ddeI  
 apyI(dcm+) hinfi apyI(dcm+) hinfi maeIII aluI(dam-) dpnII(dam-)  
 mboI/ndeII(dam-) mboI/ndeII(dam-)  
 dpnI(dam+) dpnI(dam+)

1201 GCAGGAATTT GAAAGTGACA CGTTTTTCCC AGAATTTGAT TTGGGGAAAT ATAACTCTT CCCAGATAC CCAGCGCTCC TCTCTGAGT CCAGGAGGAA  
 apoI maeIII aflIII mnlI  
 maeIII maeIII  
 scrFI mvaI ecorII dsav  
 bstNI bstNI bstNI  
 apyI(dcm+) mnlI bstNI  
 bsaJI hgaI ddeI apyI(dcm+)

1301 AAAGGCATCA AGTATAAGTT TGAAGTCTAC GAGAAGAAAG ACTAACAGGA AGATGCTTTC AAGTTCTCTG CTCCCTCCT AAAGCTATGC ATTTTATATA  
 sfanI accI mboII mboII  
 scrFI mvaI mvaI  
 ecorII ecorII ecorII  
 dsav pleI pleI  
 bstNI bstNI bstNI  
 apyI(dcm+) mnlI bstNI  
 bsaJI hgaI ddeI apyI(dcm+)

1401 GACCATGGGA CTTTGTGCTG CTTAGACCC CTTGGCTTC GTTAGAACGC GGTACAATT AATACATAAC CTTATGTATC ATACACATAG ATTTAGGTGA  
 nlaIII styI ncoI dsai bsaJI  
 fnu4HI acII thal fnuDII/mvmI tru9I  
 bstUI msei  
 bsh1236I asel/asni/vsPI  
 maeIII hphI

CTGGTACCCT GAAACGACC GAAATCTGG GGAACCGAAG CAATCTTGG CCGATGTTAA TTATGTATTG GAATACATAG TATGTGTATC TAAATCCACT



FIG. 6F

[illegible]

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## FIG. 6H

```

          eam11051
          sau96I
          scrFI
          mvaI      avall
          ecorII
          dsav
          bstNI     asuI      mboII mboII
          bsajI     mnlI      bpuAI earI/ksp632I   styI
          apyI(dcm+) bbsI mnlI      bsajI
          2401 CTTGTGACAC ACTCCCCCA TGCCACACCGT GCCCAGCACC TGAACCTCCTG GGAGGACCGT CAGTCTTCTCT CTTCCCCCA AAACCCCAAGG ATACCTTAT
          GAACACTGTG TGGAGGGGGT ACGGTGCGCA CGGTGCGTGG ACTTGAGGAC CCTCCTCGCA GTCAAGGA GAAGGGGGGT TTTGGGTTC TATGGGAATA

          sau96I
          nlaIV
          avall
          asuI
          mspI
          hpaII
          scrFI
          nciI
          dsav
          cauII
          2501 GATTTCCCGG ACCCTGAGG TCACGTGCGT GGTGTGGGAC GTGAGCCACG AAGACCCCGA GGTCCAGTTC AAGTGGTACG TGGACGGCGT GGAGGTGCAT
          CTAAGGGCC TGGGGACTCC AGTGCACGCA CCACCACCTG CACTCGGTGC TTCTGGGGCT CCAGTCAAG TTCACCATGC ACCTGCCGCA CCTCCACGTA

          acil
          thal
          fnuDII/mvnI
          bstUI
          bsh1236I
          sacII/stII
          nspBII
          kspI
          dsal
          bsajI
          aciI
          fnu4HI mnlI
          2601 AATGCCAAGA CAAGCCCGG GGAGGAGCAG TTCAACAGCA CGTTCCGCGT GGTACCGTCC CTCACCGTCC TGCACACGGA CTGGCTGAAC GGCAAGGAGT
          TTACGGTCT GTTTCGGCGC CCTCCTCGTC AAGTTGTGCT GCAAGGCACA CCAGTGCAG GAGTGCAGG ACCTGGTCTC GACCGACTTG CCGTTCCTCA

          scrFI
          mvaI bsrI
          ecorII
          dsav
          bstNI
          mnlI     ecoNI
          hgaI     hphI     bslI     apyI(dcm+)
          rsaI
          csp6I

```

**FIG. 61**

[illegible]

**FIG. 6J**

[illegible]



## FIG. 6K

nlaIV  
 scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 apyI(dcm+)  
 bsajI  
 ppulOI  
 nsII/avaIII  
 nlaIII  
 sphi  
 nspi sfaNI  
 nspHI  
 bsajI  
 nlaIV  
 scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 apyI(dcm+)  
 bsajI  
 3401 GAAAGTCCCC AGGCTCCCCA GCAGGCAGAA GTATGCAAG CATGCATCTC AATTAGTCAG CAACCAGGTG TGGAAAGTCC CCAGGCTCCC CAGCAGGCAG  
 CTTTCAGGGG TCCGAGGGGT CGTCCGTCTT CATACGTTT CATACGTAG TTAATCAGTC GTTGGTCCAC ACCTTTCAGG GGTCCGAGGG GTCTGTCGTC

sfaNI  
 ppulOI  
 nsII/avaIII  
 nlaIII  
 sphi  
 nspi  
 nspHI  
 3501 AAGTATGCAA AGCATGCATC TCAATTAGTC AGCAACATA GTCCGCCCC TAACTCCGC CTAATCCGC CCAGTTCGC CCATTCTCCG  
 TTCATACGTT TCCTACGTAG AGTTAATCAG TCGTTGTAT CAGGCGGGG ATTGAGCGG GTAGCGGGG GATTAGGGG GGTCAAGGGG GGTAGAGGC

nlaIII  
 styI  
 ncoI  
 dsal  
 bsajI  
 fnu4HI  
 sfil mnlI  
 haeIII/palI  
 bsajI bglI  
 haeIII/palI bsajI mnlI aluI  
 mnlI mnlI acII haeIII/palI  
 3601 CCCATGGCT GACTAATTTT TTTTATTTAT GCAGAGCCG AGGCGCCTC GGCCTCTGAG CTATTCCAGA AGTAGTGAGG AGGCTTTTTT GGAGGCTAG  
 GGGGTACCGA CTGATTAAA AAAATAAATA CGTCTCCGC TCCGCGGGG CCGGAGACTC GATAAGGTCT TCATCACTCC TCCGAAAAAA CCTCCGATC

nlaIII  
 styI  
 bsajI  
 blnI  
 avrII  
 haeIII/palI  
 stuI  
 haeI  
 mnlI maeI  
 3701 GCTTTTGCAA AAAGCTGTTA ACAGCTTGGC ACTGGCCGTC GTTTTACAAC GTCGTGACTG GGAACACCTT GGCGTTACCC AACTTAATCG CCTTGACGCA  
 CGAAACGTT TTTCGACAAT TGTCGAACCG TGACCGGCGAG CAAATGTTG CAGCACTGAC CCTTTTGGGA CCGCAATGGG TTGAATTAGC GGAACGTCGT

scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 apyI(dcm+)  
 bsajI  
 maeIII  
 maeIII  
 bsajI  
 apyI(dcm+)  
 tru9I  
 msel  
 fnu4HI  
 bbvI

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**FIG. 6L**

[illegible]

[illegible]

## FIG. 6N

4701 AATAATGGTT TCTTACAGCT CAGGTGGCAC TTTTCGGGGA AATGTGGCGG GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT GTATCCGCTC  
 TTATTACCAA AGAATCTGCA GTCCACCGTG AAAAGCCCTT TTACACGCGC CTGGGGGATA AACAAATAAA AAGATTATG TAAGTTTATA CATAGCGGAG  
 rcaI  
 bspHI  
 bsrBI  
 aciI nlaIII  
 fnu4HI  
 4801 ATGAGACAAAT AACCTTGATA AATGCTTCAA AATGATGATA AAGGAAGAG TATGAGTATT CAACATTTC GTGTGGCCTT TATTCCTTT TTTGGCGCAT  
 TACTCTCTTA TTGGACTAT TTACGAAGTT ATTATAACTT TTTCTCTTCT ATACTCATAA GTGTAAAGG CACAGCGGA ATAAGGAAA AACGCCGTA  
 hgiAI/aspHI  
 bsp1286  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+) bmyI  
 dpnII(dam-)  
 eco57I  
 apaLI/snoI  
 bsrI dpnII(dam-)  
 alwI(dam-)  
 4901 TTTGCCCTCC TGTTTTGGCT CACCCAGAAA CGCTGGTGAA AGTAAAGAT AGTGAAGATC AGTTGGGTGC ACGAGTGGT TACATCGAAC TGGATCTCAA  
 AACCGGAAGG ACAAACAAGA GTGGGTCTTT CGGACCACCT TCATTTTCTA CGACTTCTAG TCAACCCACG TGCTCACCCA ATGTAGCTTG ACCTAGAGTT  
 hgiAI/aspHI  
 bsp1286  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+) bmyI  
 dpnII(dam-)  
 eco57I  
 apaLI/snoI  
 bsrI dpnII(dam-)  
 alwI(dam-)  
 5001 CAGCGGTAAG ATCCTTGAGA GTTTTCGCCG CAAAGAACGT TTTCCAAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC CCGTGAAGAC  
 GTCGCCATTC TAGGAATCT CAAAGCGGG GCTTCTTGCA AAAGGTTACT ACTCGTGAAA ATTTCAGAC GATACACCGC GCCATAATAG GGCACACTAG  
 aciI  
 thai  
 fnuDII/mvnI  
 bstUI  
 bsh1236I  
 hinPI  
 hhaI/cfoI  
 5101 GCCGGGCAAG AGCAACTCGG TCGCCGCGATA CACTATTCTC AGAATGACTT GGTGAGTAC TCACCAGTCA CAGAAAGCA TCTTACGGAT GGCATGACAG  
 CGGCCCGTTC TCGTTAGACC AGCGGCGGTAT GTGATAAGAG TCTTACTGAA CCAACTCATG AGTGGTCACT GTCITTTCTG AGAATGCCTA CCGTACTCTC  
 rsal  
 csp6I  
 bsrI  
 scaI  
 hphI  
 maeIII  
 sfaNI  
 foki  
 nlaIII  
 cauII  
 bclI  
 mcrI  
 fnu4HI  
 ddeI  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 bclI  
 mcrI  
 fnu4HI  
 ddeI  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav

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## FIG. 60

sau96I  
 avalI  
 sau3AI asuI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI mnlI  
 aluI aciI  
 haeIII/palI  
 eaeI  
 cfrI  
 fnu4HI  
 aciI  
 nlaIII  
 fnu4HI  
 bbvI  
 maeIII  
 nlaIII  
 sau3AI mspI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 nlaIII aluI(dam-)  
 bsaWI  
 5201 GCACAAACATG GGGGATCATG TAACTCGCCT TATCTGTGG GAACGGAGC TGAATGAAGC CATACCAAC GACGAGCGTG ACACACCGAT GCCAGCAGCA  
 CGTGTGTGAC CCCCTAGTAC ATTGAGCGGA ACTAGCAACC CTGTGCTCG ACTTACTTCG GTATGGTTG CTGCTCGCAC TGTGGTGCTA CGTCTGCTGT  
 fnu4HI  
 bbvI  
 maeIII  
 nlaIII  
 sau3AI mspI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 nlaIII aluI(dam-)  
 bsaWI  
 5301 GCACAAACATG GGGGATCATG TAACTCGCCT TATCTGTGG GAACGGAGC TGAATGAAGC CATACCAAC GACGAGCGTG ACACACCGAT GCCAGCAGCA  
 CGTGTGTGAC CCCCTAGTAC ATTGAGCGGA ACTAGCAACC CTGTGCTCG ACTTACTTCG GTATGGTTG CTGCTCGCAC TGTGGTGCTA CGTCTGCTGT  
 hinPI  
 mstI  
 aviII/fspI bsrI  
 maeI hhaI/cfoI tru9I mseI  
 psp1406I  
 5401 ATGGCAACAA CGTTGCGCAA ACTATTAACT GCGCAACTAC TTACTCTAGC TTCCCGGCAA CAATTATAG ACTGGATGA GCGGATATA GTTGACGAC  
 TACCGTTGTT GCAACGCGTT TGATAATTGA CCGCTTGATG AATGAGATCG AAGGCGCGTT GTTAATTATC TGACCTACCT CGCCTATTT CAACGTCCTG  
 bglI  
 sau96I  
 haeIII/palI  
 hinPI asuI mspI  
 hhaI/cfoI hpaII  
 5501 CACTTCTGCG CTGCGCCCTT CCGGCTGGCT GGTATTATGC TGATAAATCT GGAGCCGGTG AGCGTGGTTC TCGCGGTATC ATTGACGAC TGGGGCCAGA  
 GTGAAGACGC GAGCCGGGAA GCGCGACCGA CCAATAACG ACTATTAGA CTCGGCCAC TCGCACCCAG AGCGCATAG TAACGTCGTG ACCCGGTCT  
 ddeI  
 sau3AI nlaIV  
 mboI/ndeII(dam-) mnlI  
 dpnI(dam+) hgiCI  
 dpnII(dam-) bniI  
 5601 TGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGG AGTCAGGCAA CTATGGATGA ACGAATAGA CAGATCGCTG AGATAGGTGC CTCACGTATT  
 ACCATTGCGG AGGCATAGC ATCAATAGAT GTGCTGCCCC TCAGTCCGTT GATACCTACT TGCTTTATCT GTCTAGCGAC TCATCCACG GAGTGACTAA

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## FIG. 6P

```

maeIII
5701 AAGCATTGGT AACGTGCAGA CCAAGTTTAC TCATATATAC TTTAGATTGA TTTAAAGACTT CATTTTAAAT TTAAGAGAT CTAGGTGAAG ATCCTTTTGG
TTCGTAACCA TTGACAGTCT GGTTCAAATG AGTATATATG AATCTAACT AAATTTTGA GTAAATAATTA AATTTTCTA GATCCACTTC TAGGAAAAAC

      rmaI      sau3AI
sau3AI hphI mboI/ndeII(dam-)
mboI/ndeII(dam-)
dpmI(dam+) dpmI(dam-)
dpmII(dam-) dpmII(dam-)
      ahaIII/draI maeI alwI(dam-)
tru9I msei ahaIII/draI msei alwI(dam-) bstYI/xhoII
ahaIII/draI msei alwI(dam-) mboII(dam-)
5801 ATAATCTCAT GACCAAAATC CCTTAACGTG AGTTTTCGTT CCACTGAGCG TCAGACCCCG TAGAAAAGAT CAAGGATCT TCTTGAGATC CTTTTTTTCT
TATTAGAGTA CTGGTTTATG GGAATTGCAC TCAAAAGCAA GGTGACTCGC AGTCTGGGGC ATCTTTTCTA GTTTCCTAGA AGAATCTAG GAAAAAAGA

      nlaIII      maeII      hgaI
rcaI      tru9I      ddeI
bspHI      msei

      sau3AI
mboI/ndeII(dam-)
dpmI(dam+) sau3AI
dpmII(dam-) mboI/ndeII(dam-)
bstYI/xhoII dpmI(dam+)
sau3AI alwI(dam-) dpmII(dam-)
mboI/ndeII(dam-) alwI(dam-)
dpmI(dam+) mboII(dam-)
dpmII(dam-) bstYI/xhoII

      sau3AI
mboI/ndeII(dam-)
dpmI(dam+)
dpmII(dam-)
alwI(dam-)

      mspI      aciI      nspBII      aciI      accACCGCTA      AACCAAAAAA      ACCACCGCTA      CCAGCGGTGG      TTTGTTTGGC      GGATCAAGAG      CTACCAACTC      TTTTCCGAA      GGTAAGTGGC
bsrI      maeIII      eco57I
5901 GCGCGTAATC TGCTGCTTGC AACCAAAAAA ACCACCGCTA CCAGCGGTGG TTTGTTTGGC GGATCAAGAG CTACCAACTC TTTTCCGAA GGTAAGTGGC
CGCGCATTAG ACGACGAACG TTTGTTTTTT TGGTGGCGAT GGTGCGCCACC AACCAACCG CCTAGTTCTC GATGTTGAG AAAAAGGCTT CCATTGACCG

      hinPI      hhaI/cfoI      rmaI      maeI      bslI      haeI      haeIII/palI
6001 TTCAGCAGAG CGCAGATACC AAATACTGTC CTCTAGTGT AGCCGTAGTT AGCCACCCAC TTCAAGAACT CTGTAGCACC GCCTACATAC CTCGCTCTGC
AAGTCGTCTC GCGTCTATGG TTTATGACAG GAAGATCACA TCGGCATCAA TCCGCTGCTG AAGTTCTTGA GACATCTGG CGGATGTATG GAGCGAGACG

```

**FIG. 6Q**

[illegible]

```
>length: 6889
```



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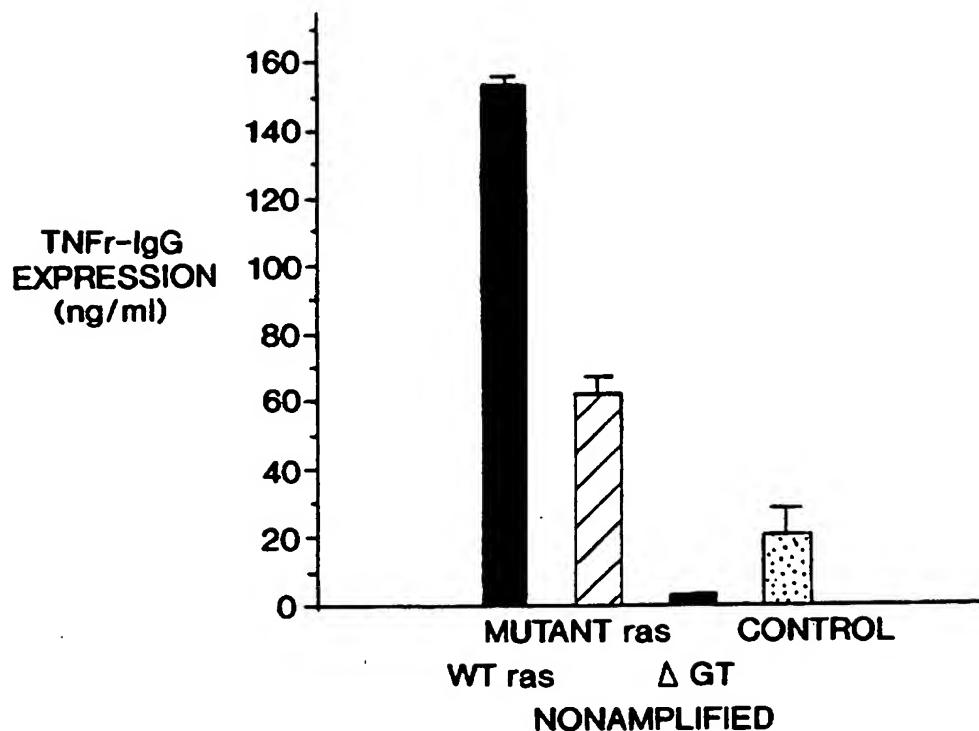


FIG. 7A

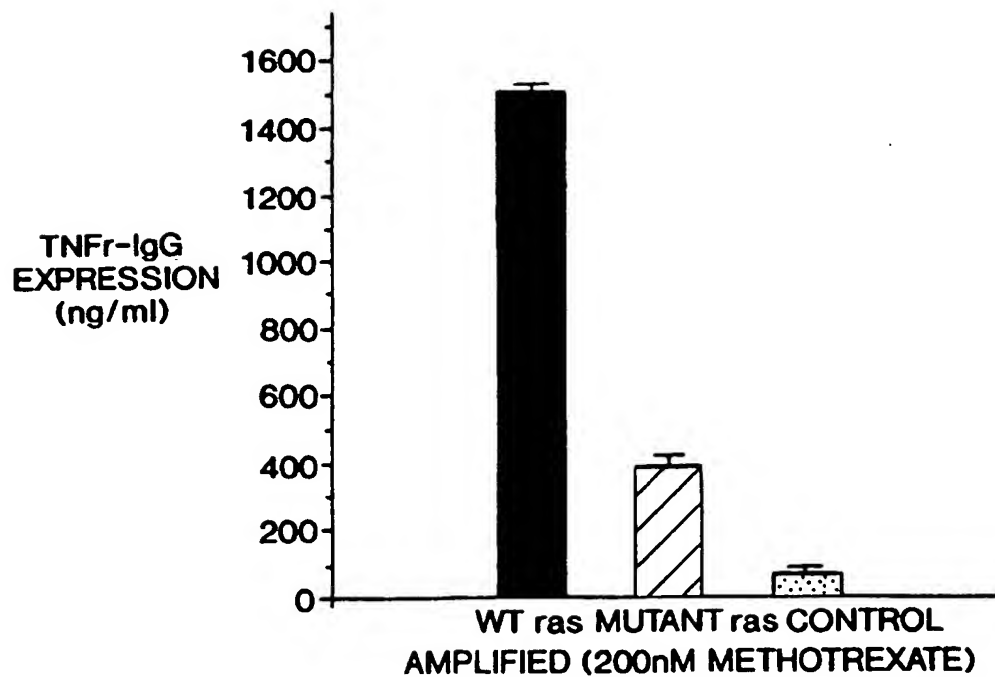


FIG. 7B

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FIG. 8

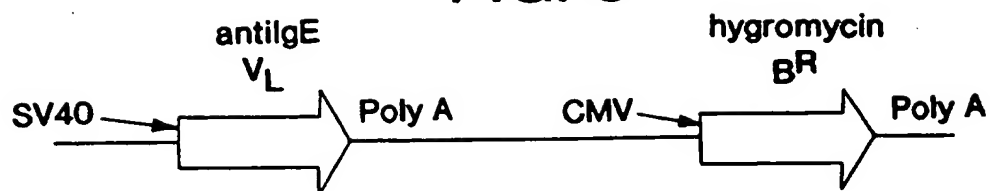
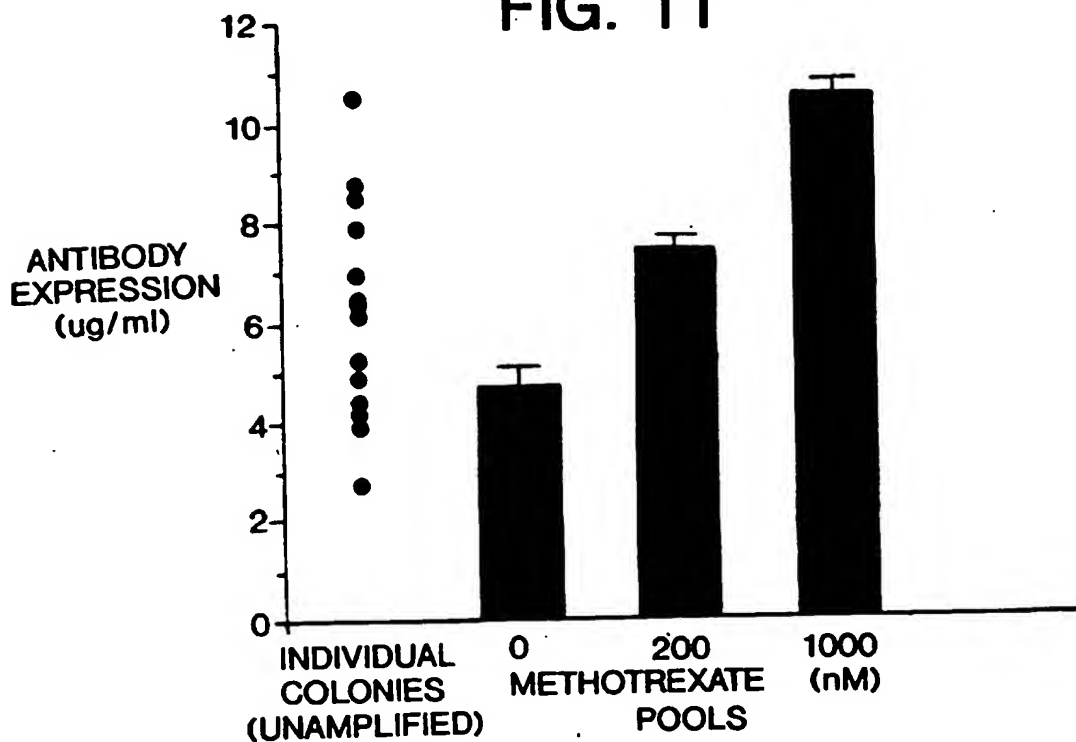


FIG. 11



## FIG. 9A

aluI  
 sstI  
 sacI  
 hgiI  
 hgiI/aspHI  
 ecli36II  
 bsp1286  
 bsiHRAI  
 bmyI  
 banII  
 taqI  
 sau3AI  
 mboI/ndeII(dam-)  
 dpmI(dam+)  
 pvuI/bspcI  
 pleI dpmII(dam-)  
 hinfI taqI(dam-)  
 rmaI mcrI nspBII  
 maeI taqI(dam-)  
 1 TCGAGCTCG CCCGACATTG ATTATTGACT AGAGTCGATC GACAGCTGTG GAATGTGTGT CAGTTAGGCT GTGGAAGTC CCCAGGCTCC CCAGCAGGCA  
 AAGCTCGAGC GGGCTGTAACTA TAATAACTGA TCTCAGCTAG CTGTGACAC CTTACACACA GTCAATCCCA CACCTTTTCAG GGGTCCGAGG GGTGCTCGT  
 nlaIV  
 sfaNI  
 scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 apyI(dcm+)  
 sexAI  
 101 GAAGTATGCA AAGCATGCAT CTCAATTAGT CAGCAACGAG GTGTGGAAG TCCCGAGGCT CCCAGCAGG CAGAAGTATG CAAGCATGTC ATCTCAATTA  
 CTTTCATACGT TTCGTACGTA GAGTTAATCA GTCGTTGGTC CACACCTTTC AGGGTCCGA GGGGTGCTCC GTCTTCATAC GTTTCGTACG TAGAGTTAAT  
 nlaIII  
 styI  
 ncoI  
 bslI dsai  
 aciI bsaJI  
 201 GTCAGCAACC ATAGTCCCG CCCTAACTCC GCCATCCCG CCCTAACTC CGCCAGTTC CGCCCATTTCT CGCCCCCATG GCTGACTAAT TTTTTTATT  
 CAGTCGTTGG TATCAGGGCG GGGATTGAGG CGGGTAGGCG GGGGATTGAG GGGGTGAAGA GCGGGGTAC CGACTGATTA AAAAAATAA  
 rmaI  
 styI  
 bsaJI  
 blnI  
 avrII  
 haeIII/pali  
 stuI  
 haeI  
 mnlI  
 mnlI  
 301 TATGCAGAGG CCGAGGCCG CTCGGCTCT GAGTATTCC AGAAGTACTG AGGAGGCTTT TTTGGAGGCC TAGGCTTTTG CAAAAGCTA GCTTATCCGG  
 ATACGTCCTCC GGCTCCGGCG GAGCCGGAGA CTCGATAGG TCTTCATCAC TCCTCCGAAA AAACCTCCGG ATCCGAAAAC GTTTTTCAT CGAATAGGCC  
 haeIII/pali  
 mcrI  
 eagi/xmaIII/ecI XI  
 eaeI  
 cfrI  
 mspI  
 hpaII  
 aluI  
 rmaI  
 maeI  
 nheI  
 aluI

**FIG. 9B**

[illegible]

## FIG. 9C

801 ACAACCGGAA TTGGCAAGTA AAGTAGACAT GGTTTGGATA GTGCGAGGCA GTTCTGTTTA CCAGGAGCC ATGAATCAAC CAGGCCACCT TAGACTCTTT  
 TGTGGCCCTT AACCGTTCAT TTCATCTGTA CCAACCTAT CAGCCTCCGT CAAGACAAAT GGTCTTCGG TACTTAGTTG GTCCGGTGA ATCTGAGAA

mspI hpaII bsaBI  
 accI nlaIII mnlI  
 scrFI mvaI ecorII dsav bstNI nlaIII bstNI ddeI pIeI  
 haeIII/paI haeI  
 scrFI mvaI ecorII dsav bstNI nlaIII bstNI ddeI pIeI  
 apyI(dcm+) hinfI apyI(dcm+) hinfI

901 GTGACAAGGA TCATGAGGA ATTGAAAGT GACACGTTT TCCAGAAAT TGATTGGGG AAATATAAC CTCTCCAGA ATACCCAGC GTCTCTCTG  
 CACTGTTCTT AGTACGCTT TAACCTTTCA CTGTGCAAA AGGCTCTTTA ACTAACCCTT TTTATATTG GAGAGGTCT TATGGGTCCG CAGGAGAGC

nlaIII sau3AI mboI/ndeII(dam-) maeII  
 dpnI(dam+) aflIII  
 maeIII alwI(dam-) apoI maeIII  
 scrFI mvaI ecorII dsav bstNI apyI(dcm+)

1001 AGGTCCAGGA GGAAAAGGC ATCAAGTATA AGTTTGAAGT CTACGAGAG AAAGACTAAC AGGAGATGC TTTCAAGTTC TCTGCTCCC TCCTAAGCT  
 TCCAGGTCTT CCTTTTCCG TAGTTCATAT TCAAACTTCA GAGCTCTTC TTTCTGATTG TCCTTCTAGC AAGTTCAAG AGAGAGGGG AGGATTTCGA

asuI mnlI sfaNI accI mboII mnlI aluI  
 avaiI  
 scrFI mvaI ecorII dsav bstNI apyI(dcm+) mnlI  
 bstNI ecorII dsav apyI(dcm+) bsaJI bslI ddeI

1101 ATGCATTTT ATAAGACCAT GGGACTTTT CTGGCTTTAG ATCCCTTTG CTTCGTTAGA ACCGAGCTAC AATTATACA TAACCTTATG TATCATACAC  
 TAGTAAAAA TATTCTGTA CCTGAAAAC GACCGAATC TAGGGAAACC GAAGCAATCT TGGGTGATG TTAATTATG ATTGAATAC ATAGTATG

ppulOI nsII/avaIII bsaJI  
 nlaIII styI ncoI dsai  
 sau3AI mboI/ndeII(dam-) dpnI(dam+) dpnII(dam-) alwI(dam-) bstYI/xhoII  
 styI bsaJI  
 aluI fnu4HI bbsI  
 asei/asnI/vspI

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## FIG. 9D

sau96I  
 avall  
 asuI  
 scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 apyI(dcm+)

maeIII  
 hphI  
 1201 ATACGATTTA GGTGACACTA TAGATAACAT CCACCTTTGCC TTCTCTCTCCA CAGGTGTCCA CTCCAGGTC CAACCTGCACC TCGGTTCTAT CGATTGAATT  
 TATGCTAAT CCACTGTGAT ATCTATTGTA GGTGAACGG AAGAGAGGT GTCCACAGGT GAGGTCCAG GTTGACGTGG AGCCAAGATA GCTAACTTAA

foki  
 scfI  
 bslI  
 bsaJI  
 mmlI  
 bsaJI  
 taqI  
 apoI  
 clal/bsp106  
 ecorI

nlaIII  
 styI  
 pflMI  
 ncoI  
 dsal  
 bslI  
 foki  
 bsaJI  
 1301 CCACCATGGG ATGGTCATGT ATCATCCTTT TTCTAGTAGC AACTGCAACT GGAGTACATT CAGAAGTTCA GCTGGTGGAG TCTGGCGGTG GCCTGGTGCA  
 GGTGGTACCC TACCAGTACA TAGTAGGAAA AAGATCATCG TTGACGTTGA CCTCATGTAA GTCTTCAAGT CGACCACCTC AGACCGCCAC CGGACCACGT

scrFI  
 mvaI  
 ecorII  
 dsav  
 bstNI  
 fnu4HI  
 apyI(dcm+)

aluI  
 pvuII  
 nspBII  
 haeI  
 hinfI  
 pleI  
 bbvI

rsaI  
 gsuI/bpmI  
 bsrI  
 csp6I  
 rmaI  
 maeI

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**FIG. 9E**

[illegible]





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## FIG. 9G

```

scrFI      hinPI      nlaIV      hgiAI/aspHI      mspI      hpaII
mvaI      nlaIV      nari      bsp1286      bsiHKAII      scrFI
ecoRII      kasi      hinII/acyI      hgiCI      haeII      bmyI
dsav      hphi      mspI      hpaII      cfr10I      fnu4HI
bstNI      bsaWI      tth111I/aspI      bsaHI      aciI      apaLI/snoI      dsav      nciI
bsII      bsaWI      tth111I/aspI      bsaHI      aciI      apaLI/snoI      dsav      nciI
apyl(dcm+)      bsaWI      tth111I/aspI      bsaHI      aciI      apaLI/snoI      dsav      nciI
fnu4HI      bsaWI      tth111I/aspI      bsaHI      aciI      apaLI/snoI      dsav      nciI
bbvI      bsaWI      tth111I/aspI      bsaHI      aciI      apaLI/snoI      dsav      nciI
1801 GGCTGCTGG TCAAGGACTA CTTCCCGAA CCGGTGACGG TGTCTGGAA CTCAGGCCCT CTGACGACCG GGTGACAC CTTCCCGCT GTCTACAGT
CCGACGACC AGTCTCTGAT GAAGGGGCTT GGCCTGCTT ACAGCACCTT GAGTCCGGG GACTGCTCG CCACGCTGTG GAAGGGCCGA CAGGATGTCA

ddei pleI      nlaIV      hgiCI      tfiI      hinfI      styI
mnlI      hinfI      fnu4HI      hgiCI      bsaI      maeII      bsaJI
eco8II      mnlI      fnu4HI      maeIII      bsp1286      rnaI      bsp1286      bmyI
bsu36I/mstII/sauI ddei bbvI hphi bmyI maeI aluI bmyI
1901 CCTCAGGACT CTACTCCCTC AGCAGGCTGG TCACTGTGCC CTCTAGCAGC TTGGGCACCC AGACTTACAT CTGCAAGCTG AATCACAAGC CCAGCAACAC
GGAGTCTCTGA GATGAGGGAG TCGTGCACCC ACTGACACCG GAGATGCTG AACCGTGGG TCTGGATGTA GAGTGTGCAC TTAGTGTTCG GGTGCTTGTG

eaml105I
sau96I

scrFI      mvaI      avall
ecoRII      dsav
bstNI      asuI      mboII mboII
bsaJI      nlaIV      bpuAI earI/ksp632I
apyl(dcm+)      bbsI mmlI
GGGACCGTC AGTCTTCTC
CTTGAGGACC CCCCTGGCAG TCAGAAGGAG

```

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## FIG. 9H

```

sau96I
nlaIV
avaII
mspi
scrFI
ncii
sau3AI hpaII
mboI/ndeII(dam-)
dpmI(dam+)
nlaIII
rcal
bspHI(dam-)
mnlI dpmII(dam-)
styI
bsaJI
2101 TTCCCCCCAA AACCCAAAGGA CACCCTCATG ATCTCCCGGA CCCCTGAGGT CACATGCCGTG GTGGTGGACG TGAGCCACGA AGACCCCTGAG GTCAAGTTCA
AAGGGGGGTT TTGGGTTTCT GTGGGAGTAC TAGAGGGCCT GGGGACTCCA GTGTACGCAC CACCACCTGC ACTCGGTGCT TCTGGGACTC CAGTTCAAGT
drdi mnlI
mboII ddeI
bpuAI eco8II
bbsI bsu36I/mstII/sauI
maeII
acil
thai
fnuDI/mvnl
bstUI
bsh1236I
sacII/stII
nspBII
kspl
dsal
bsaJI
maeII
rsal
csp6I
bsrI bsaI
mnlI
fnu4HI mnlI csp6I bsaI mnlI hphI bslI
2201 ACTGGTAGGT GGACGGCGTG GAGGTGCATA ATGCCAAGAC AAGCCGCGG GAGGAGCAGT ACAACAGCAC GTACCGTGTG GTACGGTCC TCACCGTCT'
TGACCATGCA CCTGCCGCAC CTCCACGTAT TACGGTTCTG TTTCGGCGCC CTCTCGTCA TGTTGTCTG CATGGCACAC CAGTCCGAGG AGTGGCAGGA
rsal
csp6I
bsrI bsaI
mnlI
fnu4HI bsmAI
bsaI
rsal
csp6I
2301 GCACCAAGCAGT TGGCTGAATG GCAAGGAGTA CAAGTGCAAG GTCTCCAACA AAGCCCTCCC AGCCCCCATC GAGAAACCA TCTCCAAAGC CAAAGGCGAG
CGTGGTCTCTG ACCGACTTAC CGTTCCTCAT GTTCACGTTT CAGAGTTGT TTCCGGAGGG TCGGGGGTAG CTCTTTTGT AGAGGTTTCG GTTTCCCGTC
fnu4HI
bbvI

```

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## FIG. 9I

```

scrFI      nciI      mspI      hpaII      dsav      caulI      xmaI/pspAI
smal      scrFI      nciI      dsav      caulI      foki      mboII
bsaJI      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI
avaI      bspI407I      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI
2401 CCCCAGAAC CACAGGTGA CACCTGGCC CCATCCCGG AAGAGATGAC CAAGAACCAG GTCAGCCTGA CCTGCTTGGT CAAAGGCTTC TATCCAGCG
GGGGCTCTTG GTGTCCACAT GTGGGACGGG GGTAGGCCCC TTCTCTACTG GTTCTTGCTC CAGTCGACT GACGGGACCA GTTCCGAG ATAGGGTCCG

dsal      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI
fnu4HI      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI
mspI      fnu4HI      bsaJI      bsaJI      bsaJI      bsaJI      bsaJI
2501 ACATGCGGT GGAGTGGAG AGCAATGGG AGCGGAGAA CAACTACAG ACCAGCCTC CCGTGTGGA CTGCGAGGC TCCTCTTCC TCTACAGCA
TGTAGCGCA CCTCACCTC TGTATCCG TCGGCTCTT GTTGATGTC TGTGCGGAG GGCACGACT GAGCTGCC AGGAAGAGG AGATGCTGT

pleI      hinfI      nlaIV      mboII      scfI      aluI
mnlI      nlaIII      nlaIII      nlaIII      nlaIII      nlaIII
bpuAI      bbsI      maeII      xmnI      mboII      nlaIII
asp700      nlaIII      nlaIII      nlaIII      nlaIII      nlaIII
2601 GCTCACCGT GACAAGACA GTGGCAGCA GGGGAACGTC TTCTCATGCT CCGTATGCA TGAGGCTCTG CACAACCACT ACACGAGAA GAGCTCTCC
CGAGTGGCAC CTGTTCTCGT CCACCGTCGT CCCCTGCGT AAGAGTACGA GGCACGACT ACTCCGAGAC GTGTTGGTGA TGTGCTCTT CTCGGAGAGG

scrFI      nciI      mspI      hpaII      dsav      caulI      bsmAI
2701 CTGCTCTCGG GTAAATGAGT GCGACGGCC TAGATCGAC CTGACAGAGC TTGGCCGCCA TGGCCCACT TGTATTTC AGCTTATAAT GGTACAAAT
GACAGAGGCC CATTTACTCA CGCTGCCGGG ATCTCAGCTG GACGTCTTCG AACCGCGGT ACCGGGTGA ACAATAACG TCGAATATTA CCAATGTTTA

```

**FIG. 9J**

[illegible]

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## FIG. 9K

```

nlaIII      fnu4HI      fnu4HI      hinPI      hhaI/cfoI
styI        sfII mnlI   haeIII/palI
ncoI        bsaJI bglI   haeIII/palI ddeI
bslI        haeIII/palI bsaJI mnlI aluI      mnlI
aciI        mnlI mnlI aciI haeIII/palI      mnlI
bsaJI       GCAGAGCCG AGCGGCCCTC GGCCTCTGAG CTATTCCAGA AGTAGTCAGG
3201 CCAGTTCGG CCATTCTCG CCCCATGGCT GACTAATTTT TTTTATTAT GCAGAGCCG AGCGGCCCTC GGCCTCTGAG CTATTCCAGA AGTAGTCAGG
GGTCAAGCG GGTAGAGGC GGGTACCGA CTGATTAAAA AAAATAAATA CGTCTCGGC TCGCGGGAG CCGAGACTC GATAAGGTCT TCATCACTCC

xmaI        fnu4HI      hinPI      hhaI/cfoI
styI        mcrI      egi/xmaIII/ecI XI   thai
bsaJI       eaeI      fnuDII/mvni
blnI        bsrBI      tru9I      bstUI      bspMI
avrII       xhoI notI   tru9I      bsh1236I   scfI
haeIII/palI paer7I haeIII/palI hinPI      tru9I      pstI
stui        aval fnu4HI   pacI      hhaI/cfoI   mseI   bsgI
haeI        maeIII taqI cfrI   mseI      bshII      ahaIII/draI   maeIII
mnlI maeI   aluI      mnlI aciI aciI mseI asci   swaI   sse8387I   aluI   bsrI
3301 AGGCTTTTGT GGAGGCCTAG GCTTTTGCAA AAAGCTGTTA CCTCGAGCG CGCTTAATT AAGCGGCGC ATTTAAATCC TGCAGGTAAC AGCTTGGCAC
TCCGAAAAAA CCTCCGGATC CGAAACGTT TTTCGACAAAT GGAGCTCGCC GCGGAATTA TCCGCGCGG TAAATTTAGG ACGTCCATTG TCGAACCGTG

scrFI
mvaI
ecorII
dsav
bstNI
apvi(dcm+)
tru9I      fnu4HI      aluI      mboII
bsaJI      maeIII   mseI      bbvI   foki   nspBII   earI/ksp632I
3401 TGGCGTGT TTTACAAGT CGTGACTGG AAAACCTGG CGTTACCCAA CTTAATGCC TTGCAGCAC TCCCCCTTC GCCAGCTGGC GTAATAGCGA
ACCGCAGCA AATGTTGCA GCACGACCC TTTTGGACC GCAATGGGT GAATTAGCG NACGCTGTGT AGGGGGGAG CGGTGACCG CATTATCGCT

haeIII/palI
eaeI
cfrI
3401 TGGCGTGT TTTACAAGT CGTGACTGG AAAACCTGG CGTTACCCAA CTTAATGCC TTGCAGCAC TCCCCCTTC GCCAGCTGGC GTAATAGCGA
ACCGCAGCA AATGTTGCA GCACGACCC TTTTGGACC GCAATGGGT GAATTAGCG NACGCTGTGT AGGGGGGAG CGGTGACCG CATTATCGCT

sau3AI
mboI/ndeII(dam-)
dpnI(dam+)
dpnII(dam-)
sau96I
haeIII/palI
asuI
pvuI/bspCI
mnlI aciI mcrI
3501 AGAGCCCGC ACCGATCGC CTTCGCCAACA GTTCGTAGC CTGATGGCG AATGGCGCT GATCGGGTAT TTTCCTCTTA CGCATCTGTG CGGTATTTC
TCTCGGGCG TGGCTAGCG GAAGGTTGT CAACGCATCG GACTTACCG TTACCGCGGA CTAGCCATA AAGAGGAAT GCGTAGACAC GCCATAAAGT

```

**FIG. 9L**

3601

3701

3801

3901

4001

**FIG. 9M**

[illegible]

## FIG. 9N

hgiAI/aspHI  
 bsp1286  
 bsiHKA1  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+) bmyI  
 dpnII(dam-)  
 eco57I  
 hphI  
 sfanI  
 mboII(dam-)  
 alw44I/snoI  
 maeIII  
 taqI  
 alwI(dam-)  
 aciI  
 bstYI/xhoII  
 bsrI  
 dpnII(dam-)  
 alwI(dam-)  
 bstYI/xhoII  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 sau3AI  
 nspBII  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 alwI(dam-)  
 bstYI/xhoII  
 aciI  
 bstYI/xhoII  
 mboII  
 CTGGTGAAG TAAAGATGC TGAAGATCAG TTGGGTGCAC GAGTGGGTTA CATCGAATCG GATCTCAACA GCGGTAAGAT CCTGAGAGT TTTCGCCCGG  
 GACCACCTTC ATTTCTAGC ACTTCTAGC AACCCACGTG CTCACCAAT GTAGCTTGAC GTAGAGTTGT CGCCATTCTA GGAACCTCTA AAAGCGGGC  
 maeII  
 psp1406I  
 xmnI  
 asp700  
 AAGAACGTTT TCCAATGATG AGCACTTTTA AAGTCTGCT ATGTGCGCG GTATTATCCC GTGATGACGC CGGGCAAGAG CAACTCGGTC GCGGCATACA  
 TTCTTGCAAA AGGTTACTAC TCGTGAAAAT TTCAAGACGA TACACGCGC CATANTAGG CACTACTGCG GCCCGTTCTC GTTGAGCCAG CGCGGTATGT  
 rsaI  
 csp6I  
 bsrI  
 ddeI  
 CTATTCTCAG AATGACTTGG TTGAGTACTC ACCAGTCACA GAAAGCATC TTACGGATGG CATGACAGTA AGAGAATTAT GCAGTCTGC CATAACCATG  
 GATAAGAGTC TTAAGTGAAC AACTCATGAG TGGTCAGTGT CTTTTCGTAG ATGCCTACC GTACTGTCTAT TCTCTTAATA CGTCACGAGC GTATTGGTAC  
 fnu4HI  
 bbvI  
 nlaIII  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI  
 mnlI  
 aluI  
 aciI  
 nlaIII  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 maeIII  
 nlaIII  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 alwI(dam-)  
 nlaIII  
 alwI(dam-)  
 GGATCATGTA ACTCGCCTTG  
 TCACTATTGT GACGCCGCTT GAATGAAGAC TGTGTGCTAGC CTCCTGGCTT CCTCGATTGG CGAAAAACG TGTGTGACCC CCTAGTACAT TGAGCGGAAC  
 4601  
 4701  
 4801  
 4901



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## FIG. 90

```

mspI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI      hinPI
hpaII     mstI       avIII/fspI  bsrI      bsrI      bsrI      bsrI      bsrI      bsrI      bsrI
bsaWI     maeII      hhaI/cfoI  tru9I     tru9I     tru9I     tru9I     tru9I     tru9I     tru9I
nlaIV     maeII      psp1406I   mspI      mspI      mspI      mspI      mspI      mspI      mspI
5001 ATCGTTGGGA ACCGGAGCTG AATGAAGCCA TACCACGCA CGAGCGTGAC ACCAGCATGC CAGCAGCAAT GGCACCAACG TTGCGCAAC TATTAACTGG
TAGCAACCT TGGCCTCGAC TTACTTCGGT ATGGTTTGGT GCTCGCACTG TCGTGCTACG GTCGTCGTTA CCGTTGCTGC AACCGTTTG ATAATTGACC

mspI      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII
scrFI     nciI       dsav       cauII     cauII     cauII     cauII     cauII     cauII     cauII
xmaI      fokI       fokI       fokI       fokI       fokI       fokI       fokI       fokI       fokI
5101 CGAACTACTT ACTCTAGCTT CCGGGCAACA ATTAATAGAC TGGATGGAGG CGGATAAAGT TGCAGGACCA CTTCTGCGCT CGGCCCTTCC GGCTGCGCTGG
GCTTGATGAA TGAGATCGAA GGGCCGTTGT TAATTATCTG ACCTACCTCC GCCTATTCTA AGTCTCTGCT GAAGACGCGA GCCGGGAAGG CCGACCGACC

mspI      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII      hpaII
scrFI     nciI       dsav       cauII     cauII     cauII     cauII     cauII     cauII     cauII
xmaI      fokI       fokI       fokI       fokI       fokI       fokI       fokI       fokI       fokI
5201 TTTATTGCTG ATAAATCTGG AGCGGGTGAG CGTGGCTCTC GCGGTATCAT TGCAGCACTG GGGCCAGATG GTAGCCCTCT CCGTATCGTA GTTATCTACA
AAATAACGAC TATTAGACC TCGGCCACTC GCACCCAGAG CGCATATGTA ACCTGCTGAC CCGGCTCTAC CATTCGGGAG GGCATAGCAT CAATAGATGT

pleI      hinfI     eam1105I   foki      foki      foki      foki      foki      foki      foki
5301 CGACGGGGAG TCAGGCAACT ATGGATGAAC GAAATAGACA GATCGCTGAG ATAGGTGCTT CACTGATTA GCAATTGCTAA CTGTCAGACC AAGTTTACTC
GCTGCCCTC AGTCCGTTGA TACCTACTTG CTTTATCTGT CTAGCGACTC TATCCACGGA GTGACTTAAT CGTAACCAAT GACAGTCTGG TTCAAATGAG

truaI     sau3AI     sau3AI     sau3AI     sau3AI     sau3AI     sau3AI     sau3AI     sau3AI     sau3AI
sau3AI     hphI      hphI      hphI      hphI      hphI      hphI      hphI      hphI      hphI
mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-) mboI/ndeII(dam-)
5401 ATATATACTT TAGATTGATT TAAACTTCA TTTTAAATTT AAAAGGATCT AGGTGAAGAT CCTTTTGTAT AATCTCATGA CCAAATCCC TTAACGTGAG
TATATATGAA ATCTAATACTA ATTTGAAGT AAAAATTA TTTTCTCTAG TCCACTTCTA GGAANAACCTA TTAGACTACT GGTTTTAGGG AATTGCCACTC

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## FIG. 9P

```

sau3AI
mboI/ndeII[dam-]
dpmI[dam+] sau3AI
dpmII[dam-] mboI/ndeII[dam-]
bstYI/xhoII dpmI[dam+] fnuDII/mvnI
sau3AI alwI[dam-] dpmII[dam-] bstUI
mboI/ndeII[dam-] alwI[dam-] bsh1236I
dpmI[dam+] mboII[dam-] hinPI fnu4HI
dpmII[dam-] bstYI/xhoII hhaI/cfoI bbvI
5501 TTTTCGTTCC ACTGAGCTC AGACCCCGTA GAAAGATCA AAGCATCTTC TTTTCTGCG GCGTAATCTG CTGCTTGCAA ACAAAAAAAC
AAAAACAAGG TGAATCGCAG TCTGGGGCAT CTTTCTAGT TTCTAGNAG AACTCTAGGA AAAAAAGACG CGCATTAGAC GACGAACGTT TGTTTTTTGG

hgaI
dclI
sau3AI
mboI/ndeII[dam-]
dpmI[dam+]
dpmII[dam-]
alwI[dam-]
mspi
acii
nspBII
hpaII
alul
5601 CACCGCTACC AGCGGTGGTT TGTTCGCGG ATCAAGAGCT ACCAACTCTT TTTCGGAAGG TAACGTGGCTT CAGCAGAGCG CAGATACCAA ATACTGTCCT
GTGGCGATGG TCGCCACCAC ACAAACGGCC TAGTTCTCGA TGGTTGAGAA AAAGCTTCC ATTGACCGAA GTGCTCTGCG GTCTATGGTT TATGACAGGA

hmaI
bsII
maeI
scfI
acii
mnII
5701 TCTAGTGTAG CGGTAGTTAG GCCACCACTT CAAGAAGCTT GTAGCACCGC CTACATACCT CGCTCTGCTA ATCTGTGTAC CAGTGGCTGC TGCCAGTGGC
AGATCACATC GGCATCAATC CGGTGGTGAA GTTCTTGAGA CATCTGGCG CATGTATGGA GCGAGACGAT TAGGACAATG GTCACCCGACG ACGGTCAACG

rmaI
bsII
maeI
scfI
acii
mnII
alwNI
bbvI
bsrI
fnu4HI
5801 GATAAGTCGT GTCTTACCGG GTTGACTCA AGACGATAGT TACCGGATAA GCGGCGTGA CGGGGGGTTC GTGCACACAG CCCAGCTTGG
CTATTACGCA CAGATGGCC CAACCTGAGT TCTGTATCA ATGGCTATT CCGCGTGGCC AGCCCGACTT GCGCCCAAG CACGTGTGTC GGTCCGAACC

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SUBSTITUTE SHEET (RULE 26)



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## FIG. 10A

```

aluI
sstI
sacI
hgiI/II
hgiAI/aspHI
ecII/36II
bspI286
bsiHKAII
bmyI
banII
    1  TTCGAGCTCG CCGGACATG ATTATTGACT AGTTATTAAAT AGTAATCAAT TACGGGGTCA TTAGTTCATA GCCCATATAT GGAGTTCCGC GTTACATAAC
    2  AAGCTCGAGC GGGCTGTAAC TAATAACTGA TCAATAATTA TCATTAGTTA ATGCCCCAGT AATCAAGTAT CCGGTATATA CCTCAAGGCG CAATGTATTG
    3  taqI
    4  scrFI
    5  mvaI
    6  ecorII
    7  dsav
    8  aciI
    9  bglI bstNI
    10 sau96I
    11 haeIII/palI aciI
    12 asuI apyI[dcn+]
    13 101 TTACGGTAA TGGCCCGCT GGCTGACCG CCAACGACC CCGCCCATG ACGTCAATAA TGACGTATGT TCCCATAGTA ACGCCAAATAG GGACTTTCCA
    14 AATGCCATT ACCGGGCGA CCGACTGGC GGTGCTGGG GCGGGGTAAC TGCAGTTATT ACTGCATACA AGGTATCAT TCGGTTATC CTGAAAGGT
    15 maeII
    16 hinII/acyI
    17 ahaII/bsaHI
    18 aatII
    19 201 TTGACGTCAA TGGGTGGAGT ATTACGGTA AACTGCCCCA TTGGCAGTAC ATCAAGTGA TCATATGCCA AGTACGCCCC CTATTGACGT CAATGACGGT
    20 AACTGCAGT ACCACCTCA TAAATGCCAT TTGACGGGTG AACCGTCAAT TAGTTCACAT AGTATACGCT TCAATGCGGG GATAACTGCA GTTACTGCCA
    21 maeII
    22 hinII/acyI
    23 ahaII/bsaHI
    24 aatII
    25 301 AAATGGCCG CCTGGCATA TGCCCACTAC ATGACCTTAT GGGACTTTCC TACTTGGCAG TACATCTACG TATTAGTCAT CGCTATTACC ATGGTGATCC
    26 TTACCGGGC GGACCGTAAT ACGGTCATG ACGGTCATG TACTGGAATA CCTGAAAGG ATGAACCGTC ATGATAGTGC ATATACAGTA GCGATAATGG TACCACCTACG
    27 nlaIII
    28 styI
    29 ncoI
    30 dsai hphI aciI
    31 bsaJI sfaNI

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**FIG. 10B**

[illegible]

**FIG. 10C**

SUBSTITUTE SHEET (RULE 26)

**FIG. 10D**

[illegible]



## FIG. 10E

aciI  
 thai  
 fnuDII/mvnI  
 hgiAI/aspHI  
 bsp1286  
 bsiHKA1  
 bmyI bstUI  
 apaLI/snoI  
 alw44I/snoI  
 mnII bsh1236I nlaIV  
 1501 ACCTCGTGCA CGCGGATTTC GGCTCCAACA ATGTCCTGAC GGACAATGGC CGCATAACAG CGGTCAATGA CTGGAGCGAG GCGATGTTGC GGGATTCCCA  
 TGGAGCACGT GCGCCTAAG CCGAGGTTGT TACAGGACTG CCTGTACCG GCGTATTGTC GCCAGTAAGT GACCTCGCTC CGCTACAAGC CCCTAAGGGT  
 fnu4HI  
 aciI  
 haeIII/palI  
 eaeI  
 cfrI  
 nspBII  
 gsuI/bpmI  
 bsrI  
 mnII  
 tffI bslI  
 hinFI  
 fnu4HI  
 thai  
 fnuDII/mvnI  
 bstUI  
 bsh1236I  
 sacII/sstII  
 nspBII  
 kspI  
 dsal  
 bsaJI  
 aciI  
 fnu4HI  
 sau3AI aciI  
 mboI/ndelI[dam-]  
 dpnII[dam+]  
 dpnII[dam-]  
 alwI[dam-]  
 AGGATCGCGG  
 1601 ATACGAGGTC GCCAACATCT TCTTCTGGAG GCGGTGGTTG GCTTGTATGG AGCAGCAGAC GTACTTCGAG CGGAGGCATC CGGAGCTTGC AGGATCGCGG  
 TATGCTCCAG CGGTGTAGA AGAAGACCTC CGGCACCAAC CGACATACC TCGTCTCTG CATGAAGCTC GCCTCCGTAG GCCTCAAGC TCCTAGCGGC  
 dsal  
 haeIII/palI  
 mnII mnII bsaJI  
 mboII mnII gsuI/bpmI  
 mnII mboII  
 1701 CGGCTCCGGG CGTATATGCT CCGCAATTGT CTTGACCAAC TCTATCAGAG CTGGTTGAC GGCATTTTCG ATGATGCAGC TTGGGGCGCAG GGTTCATGCG  
 GCCGAGGCCC GCATATACGA GCGGTAACTG GAACCAACTG CCGTTAAAGC TACTACGTGC AACCCGGCTC CCAGCTACGC  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 cauII  
 nlaIV  
 1701 CGGCTCCGGG CGTATATGCT CCGCAATTGT CTTGACCAAC TCTATCAGAG CTGGTTGAC GGCATTTTCG ATGATGCAGC TTGGGGCGCAG GGTTCATGCG  
 GCCGAGGCCC GCATATACGA GCGGTAACTG GAACCAACTG CCGTTAAAGC TACTACGTGC AACCCGGCTC CCAGCTACGC  
 aluI  
 fnu4HI  
 bbvI  
 aluI nincII/hindII taqI sfaNI  
 hinPI  
 taqI  
 hhaI/cfoI sfaNI  
 hgaI  
 drdI



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## FIG. 10G

2001 CAACCTTGTTT ATTCAGCTT ATAATGGTTA CAATAAAGC AATAGCATCA CAATTTCAC AATAAAGCA TTTTTCAC TGCATTCTAG TTGTGGTTG  
 GTTGAACAA TAACGTCGAA TATTACCAAT GTTATTTCG TTATCGTAGT GTTAAAGTG TTATTTCGT AAAAAAAGTG ACCTAAGATC AACACCAAC  
 aluI rnaI  
 fnu4HI bsmI maeI  
 bbvI  
 sau3AI  
 mboI/ndeII(dam-) spoI  
 dpnI(dam+) maeI  
 dpnII(dam-) bsmI  
 pvuI/bspCI  
 mcrI  
 taqI(dam-) tru9I haeIII/palI  
 claI/bsp106(dam-) haeI  
 sau3AI fnu4HI styI  
 mboI/ndeII(dam-) bbvI ncoI  
 dpnI(dam+) xmnI hinPI dsai  
 dpnII(dam-) aseI/asnI/vspI bsaJI  
 aluI nlaIII aluI/dam-] asp700 hhaI/cfoI nlaIII mnlI  
 2101 TCCAAACTCA TCAATGATC TTATCATGTC TCGATCGATC GGGAAATTAAT TCGGCGCAGC ACCATGGCCT GAAATAACCT CTGAAGAAGG AACTTGTTA  
 AGGTTTGAGT AGTTACATAG AATAGTACAG ACCTAGCTAG CCCTTAATTA AGCCGGCTCG TGGTACCAGA CTTTATTGGA GACTTCTCC TTGAACCAAT  
 rsaI nlaIV  
 csp6I scrFI sfaNI  
 nlaIV mvai ppulOI  
 kpnI ecorII nsii/avaIII  
 hgiCI dsav nlaIII  
 banI bstNI sphI  
 asp718 aluI apyI(dcm+) nsPI  
 acc65I ddeI acil bsaJI nspHI  
 2201 GGTACCTTCT GAGGCGGAAA GAACCAAGT TCGAATGTGT GTGAGTTAGG GTGTGGAAG TCCCGAGGCT CCCAGCAGG CAGAAGTATG CAAGCATGC  
 CCATGGAAGA CTCCGCCITT CTGTCGTCGAC ACCTACACA CAGTCAATCC CACACCTTTC AGGGTCCGA GGGTCCGTC GTCTTCATAC GTTTCGTACG  
 scrFI nlaIV  
 mvai mvai ppulOI  
 ecorII nsii/avaIII  
 dsav nlaIII  
 bstNI bstNI sphI  
 apyI(dcm+) apyI(dcm+) nsPI sfaNI  
 sexAI bsaJI nspHI  
 2301 ATCTCAATTA GTCAGCAACC AGGTGTGGAA AGTCCCCAGG GTCCCCAGCA GGCAGAAGTA TGCAGAAGCAT GCATCTCAAT TAGTCAGCAA CCATAGTCCC  
 TAGAGTTAAT CAGTCGTTGG TCCACACCTT TCAGGGGTCC GAGGGTCTGT CCGTCTTCAT ACCTTTCGTA CGTAGAGTTA ATCAGTCGTT GGTATCAGGG  
 acil

**FIG. 10H**

[illegible]

**FIG. 10I**

[illegible]

**FIG. 10J**

[illegible]

## FIG. 10K

sstI  
 sacI  
 hgiII  
 hgiAI/aspHI  
 ecl136II  
 bsp1286  
 bsiHKA1  
 bmyI  
 haeIII/palI  
 sau96I aluI  
 asuI banII  
 hphI  
 eco109I/draII  
 maeIII alwNI ddeI  
 accI  
 haeIII  
 hgiII/espI  
 bpul102I  
 hgaI  
 ddeI fnu4HI  
 scfI mnlI bbvI  
 3401 AGGACAGCAC CTACAGCCTC AGCAGCACCC TGACGCTGAG CAAAGCAGAC TAGGAGAAAC ACAAGTCTA GCGCTGCGAA GTACCCCATC AGGCGCTGAG  
 TCGTGTCTG GATGTGGG GATGTGGG TCGTGTCTG ACTCGACTC GTTGTCTG ATGCTCTTG TGTTTCAGAT GCGGACGCTT CAGTGGGTAG TCCCGACTC  
 ddeI  
 celII/espI  
 bpul102I  
 hgaI  
 ddeI fnu4HI  
 scfI mnlI bbvI  
 3501 CTGCGCCGTC ACAAGAGCT TCAACAGGGG AGAGTGTAA GCTTCGATGG CCGCCATGGC CCACTTGT TATTGCAGCT TATAATGTT ACAATAAAG  
 GAGCGGCAG TGTCTCTCGA AGTTGTCCCG TCTCACAATT CGAAGCTACC GCGGTACCG GGTGAACAA ATAACGTCA ATATTACCA TGTATTTC  
 maeIII aluI maeI  
 taqI haeIII/palI  
 mseI  
 tru9I  
 hindIII  
 aluI  
 sfii ncoI  
 eaeI  
 cfrI bsaJI  
 fnu4HI  
 aluI  
 fnu4HI  
 bbvI  
 maeIII  
 sau96I  
 nlaIII  
 aciI haeIII/palI  
 fnu4HI asuI  
 bgli styI  
 aluI  
 hindIII  
 eaeI  
 cfrI bsaJI  
 mseI  
 taqI haeIII/palI  
 3601 CAATAGCATC ACAATTTCA CAAATAAGC ATTTTTCATC CTGCTTCTA GTTGTGTTT GTCCAACTC ATCAATGTAT CTTATCATGT CTGATCGAT  
 GTTATCGTAG TGTATAAGT GTTATAAGT TAAAAAAGT GAGTAAGT CAACACCCAA CAGGTTGAG TAGTTACATA GAATAGTACA GACCTAGCTA  
 sfanI apoI  
 bsmI maeI  
 rmaI  
 nlaIII alwNI ddeI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 pvuI/bspCI  
 mcrI  
 taqI(dam-)  
 claI/bsp106(dam-)  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 nlaIII alwNI ddeI

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## FIG. 10L

```

          haeIII/palI      rsal      csp6I      nlaIV      kpnI      hgiCI      banI      asp718      mnlI      pvuII      aluI      nspBII
          haeI              nlaIV      kpnI      hgiCI      banI      asp718      mnlI      pvuII      aluI      nspBII
          tru9I      fnu4HI styI      fnu4HI      fnu4HI      fnu4HI      fnu4HI      fnu4HI      fnu4HI      fnu4HI      fnu4HI      fnu4HI      fnu4HI
          mseI      bbvI      ncoI      mseI      bbvI      ncoI      mseI      bbvI      ncoI      mseI      bbvI      ncoI
          aseI/asnI/vspi      dsal      aseI/asnI/vspi      dsal      aseI/asnI/vspi      dsal      aseI/asnI/vspi      dsal      aseI/asnI/vspi      dsal
          xmnI      hinPI      bsajI      xmnI      hinPI      bsajI      xmnI      hinPI      bsajI      xmnI      hinPI      bsajI
          asp700      hhaI/cfoI nlaIII      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI      mnlI
          3701 CGGGAATTAA TTCGGCGCAG CACCATGGCC TGAATAAACC TCTGAAGAG GAACTTGGTT AGGTACCTTC TGAGGCGGAA AGAACCCAGT GTGGAATGTG
          GCCCTTAATT AAGCCGCGTC GTGGTACCGG ACTTTATTGG AGACTTCTC CTGGAACCAA TCCATGGAAG ACTCCGCTT TCTTGGTGA CACCTTACAC

          nlaIV      scrFI      mvaI      ecorII      dsav      bstNI      apyI[dcn+]      bsajI      nspHI      nlaIV
          scrFI      mvaI      ecorII      dsav      bstNI      apyI[dcn+]      bsajI      nspHI      nlaIV
          3801 TGTCAGTTAG GGTGTGMAA GTCCCCAGC TCCCCAGCAG GCAGAAGTAT GCAAGCATG CATCTCAATT AGTCAGCAAC CAGGTGTGGA AGTCCCCCAG
          ACAGTCAATC CCACACCTT CAGGGGTCCG AGGGGTCCG CGTCTTCATA CGTTTCGTAC GTAGAGTTAA TCAGTGTGTT GTCCACACCT TTCAGGGGTC

          ppulOI      nsII/avaIII      nlaIII      sphI      nspI      nspHI      nspI      nspHI      nspI      nspHI      nspI      nspHI
          ppulOI      nsII/avaIII      nlaIII      sphI      nspI      nspHI      nspI      nspHI      nspI      nspHI      nspI      nspHI
          3901 GCTCCCCAGC AGGCAGAAGT ATGCAAGCA TGCATCTCAA TTAGTCAGCA ACCATAGTCC CGCCCTAAC TCCGCCCATC CGCCCTTAA CTCGCCCCAG
          CGAGGGGTG TCGTCTTCA TAGCTTCTG ACGTAGAGTT AATCAGTCGT TGGTATCAGG GCGGGGATTG AGCGGGTAG GCGGGGATT GAGCGGGTTC

          fnu4HI      bgli      sfiI      haeIII/palI      mnlI      mnlI      ddel      haeIII/palI      bsajI      mnlI      aluI      haeIII/palI
          fnu4HI      bgli      sfiI      haeIII/palI      mnlI      mnlI      ddel      haeIII/palI      bsajI      mnlI      aluI      haeIII/palI
          4001 TTCGCCCAT TCTCCGCCC ATGGCTGACT AATTTTTTTT ATTTATGCAG AGGCCGAGC CGCTCGGCC TCTGAGCTAT TCCAGAAGTA GTGAGGAGGC
          AAGCGGGTA AGAGCGGGG TACCGACTGA TTAATAAATAA TAAATACGTC TCCGGTCCG GCGAGCCCG AGACTCGATA AGGTCTTCAT CACTCTCTCG

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## FIG. 10M

```

4101 TTTTGGAG CCTAGGCTT TTGCAAAAAG CTGTTAAGAG CTTGGCACTG GCCTGCTTT TACAACGTGG TGACTGGGAA AACCTGGCG TTACCCAAC T
AAAAACCTC CGGATCCGAA AACGTTTTTC GACAATTGTC GAACCGTGAC CGGCAGCAAA ATGTTGCAGC ACTGACCCCTT TTGGACCGC AATGGGTTGA

styI          scrFI
bsaJI          mvaI
blnI           ecorII
             dsav
             bstNI
             apyI(dcm+)   tru9I
             bsaJI maeIII mseI
             maeIII bsrI
             maeIII

             sau3AI
             mboI/ndeII(dam-)
             sau96I dpnI(dam+)
             haeIII/palI
             mmlI aciI dpnII(dam-)
             aluI mboII asuI pvuI/bspcI
             pvuII earI/ksp63II mcrI
             nspBII earI/ksp63II mcrI
             fnu4HI bbvI foki
             GCAGCACATC CCCCTTCGC CAGCTGGCGT AATAGCGAAG AGGCCGCGAC CGATCGCCCT TCCCAACAGT TCGGTAGCCT GAATGGCGAA
             ATTAGCGGAA CGTCGTGTAG GGGGGAAGCG GTCGACCGCA TTATCGCTTC TCCGGCGGTG GCTAGCGGSA AGGTTGTCA ACGCATCGGA CTTACCGCTT

             hinPI          hinPI          hinPI          hinPI          hinPI          hinPI          hinPI          hinPI
             hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI
             nlaIV          thal          thal          thal          thal          thal          thal          thal
             kasi          fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI
             hinII/acyI      bstUI          bstUI          bstUI          bstUI          bstUI          bstUI          bstUI
             hgiCI          bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I
             haeII          rsal          rsal          rsal          rsal          rsal          rsal          rsal
             aciI          scfI          scfI          scfI          scfI          scfI          scfI          scfI
             banI          fnu4HI          fnu4HI          fnu4HI          fnu4HI          fnu4HI          fnu4HI          fnu4HI
             sfaNI          tru9I          tru9I          tru9I          tru9I          tru9I          tru9I          tru9I
             ahaII/bsaHI      csp6I          csp6I          csp6I          csp6I          csp6I          csp6I          csp6I
             TGGCGCCTGA TGGGGTATTT TCTCCTTACG CATCTGTGCG GTATTTCACA CCGCATACGT CAAAGCAACC ATAGTACGCG CCTGTAGCG GCGCATTAAG
             ACCGCGGACT ACGCCATANA AGAGGAATGC GTAGACACGC CATAAAGTGT GGCATGCA GTTCTGTTGG TATCATCGC GGCATCGC CCGTAAATTC

             fnu4HI          fnu4HI          fnu4HI          fnu4HI          fnu4HI          fnu4HI          fnu4HI          fnu4HI
             hinPI          hinPI          hinPI          hinPI          hinPI          hinPI          hinPI          hinPI
             hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI
             thal          thal          thal          thal          thal          thal          thal          thal
             fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI    fnuDII/mvnI
             bstUI          bstUI          bstUI          bstUI          bstUI          bstUI          bstUI          bstUI
             bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I
             maeIII          maeIII          maeIII          maeIII          maeIII          maeIII          maeIII          maeIII
             bbvI          bsrBI          bsrBI          bsrBI          bsrBI          bsrBI          bsrBI          bsrBI
             hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI      hhaI/cfoI
             aciI          aciI          aciI          aciI          aciI          aciI          aciI          aciI
             maeIII          maeIII          maeIII          maeIII          maeIII          maeIII          maeIII          maeIII
             bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I      bsh1236I
             CGCGCGGGT GTGGTGGTTA CGCGCAGCGT GACCGCTACA CTTGCCAGCG CCCTAGCGCC CGCTCCTTTC GCTTCTTTC CCGCACGTTT
             CCGCCGCCCA CACCACCAAT CGCGCTGCGA CTGGCGATGT GAACGGTCCG GGGATCGCG GCGAGGAAAG CGAAGAGAGG GAAGGAAAGA GCGGTGCAAG

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nn11

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SUBSTITUTE SHEET (RULE 26)

## FIG. 10P

sau3AI mboI/ndeII(dam-) sau3AI  
 mboI/ndeII(dam-) mboI/ndeII(dam-)  
 dpmI(dam+) dpmI(dam+)  
 dpmI(dam-) dpmI(dam-)  
 l-styI/xhoII alwI(dam-)  
 bsrI nspBII alwI(dam-)  
 taqI alwI(dam-) acII bstyI/xhoII  
 5401 TCGAACTGGA TCTCAACAGC GGTAAAGATCC TTGAGAGTTT TCGCCCGGAA GAACGTTTC CAATGATGAG CACTTTTAAA GTTCGTCTAT GTGGCGCGGT  
 AGCTTGACCT AGAGTTGTGCG CCATTCTAGG AACTCTCAAA AGCGGGGCTT CTTCGCAAAAG GTTACTACTC GTGAAATTTT CAAGACGATA CACCGCGCCA  
 scrFI  
 nciI  
 mspI  
 hpaII  
 dsav  
 caulI  
 hinII/acyI  
 hgaI  
 ahaII/bsaHI bclI mcrI fnu4HI acII  
 5501 ATTATCCCGT GATGACCGG GGAAGAGCA ACTCGGTGCG CGCATACACT ATTCTCAGAA TGACTTGCTT GAGTACTCAC CAGTCACAGA AAGCATCTT  
 TAATAGGSCA CTACTCGGC CCGTTCTCGT TGAGCCAGCG GCGTATGTGA TAAGAGTCTT ACTGAACCAA CTCATGAGTG GTTCAGTGTCT TTTCGTAGAA  
 foki nlaIII fnu4HI bsvI nlaIII  
 5601 ACGGATGGCA TGACAGTAAG AGAATTATGC AGTGCTGCCA TAACCATGAG TGATAACACT GCGGCCAAT TACTTCTGAC AACGATCGGA GGACCGAAGG  
 TGCCTACCGT ACTGTCAATC TCTTAATACG TCACGACGCT ATTGGTACTC ACTATTGTA GCGCGGTTCA ATGAAGACTG TTGCTAGCCT CCGGCTTCC  
 sau3AI maeIII  
 mboI/ndeII(dam-) sau3AI nlaIV mspI  
 dpmI(dam+) mboI/ndeII(dam-)  
 alwI(dam-) dpmI(dam+) hpaII  
 nlaIII dpmI(dam-) dpmI(dam-) bsaWI aluI  
 5701 AGCTAACCGC TTTTTCAC ACATGCGGG ATCATGTAAC TCGCCTTGAT CGTTGGGAC CGGAGCTGAA TGAAGCCATA CCAAGCAGC AGCGTGACAC  
 TCGATTGGCG AAAAAACGTG TTGTACCCCC TAGTACATTG AGCGGAACCTA GCAACCCCTTG GCCTCGACTT ACTTCGTAT GTTTCGTGCG TCGCACTGCG

## FIG. 10Q

hinPI mspI  
 mstI hpaII  
 avIII/fspI aluI nciI scrFI  
 maeII hhaI/cfoI bsrI tru9I foki  
 pspl406I maeI dsav bsrI acil  
 5801 CACGATGCCA GCAGCAATGG CAACAACGTT TTAACCTGGC AACTACTTAC TCTAGCTTCC CGGCAACAAT TAATAGACTG GATGGAGGCG  
 GTGCTACGGT CGTCGTTACC GTTGTTCGAA CGGCTTTGAT AATTGACCGC TTGATGAATG AGATCGAAGG GCCGTTGTTA ATTATCTGAC CTACCTCCGC  
 fnu4HI  
 bsvI  
 sfaNI  
 bglI mspI  
 sau96I hpaII  
 haeIII/palI cfr10I  
 hinPI asuI mspI  
 hhaI/cfoI hpaII  
 5901 GATAAAGTTG CAGGACCACT TCTGGCTCG GCCCTTCCG CTGGCTGGTT TATTGCTGAT AAATCTGGAG CCGGTGAGCG TGGGTCTGCG GGTATCATTTG  
 CTATTTCAAC GTCTGTGTA AGACGGAGC CGGGAAGGCC GACCGACCAA ATAACGACTA TTTAGACCTC GCCCACTGCG ACCCAGAGCG CCATAGTAAC  
 sau96I  
 avalI  
 asuI  
 nlaIV hphI  
 gsuI/bpmI  
 bsmAI  
 bsaI bsh1236I bsvI  
 6001 CAGCACTGGG GCCAGATGGT AAGCCCTCCC GTATCGTAGT TATCTACACG ACGGGGAGTC AGGCAACTAT GGATGAACGA AATAGACAGA TCGCTGAGAT  
 GTCGTGACCC CGGTCTACCA TTCGGGAGGG CATAGCATCA ATAGATGTGC TGCCCTCAG CCTACTTGCT TATCTGTCT AGCGACTCTA  
 sau96I  
 asuI  
 nlaIV  
 bsrI haeIII/palI mnlI  
 eam1105I  
 foki  
 6101 AGGTGCCTCA CTGATTAGC ATTGGTAAGC GTGACACCAA GTTACTCAT ATATACCTTA GATTGATTTA AAATTCATT TTTAATTTAA AAGGATCTAG  
 TCCACGGAGT GACTAATTTCG TAACCATTGA CAGTCTGGTT CAAATGAGTA TATATGAAT CTAACATAAT TTGAAGTAA AATTAATTT TTCTAGATC  
 mnlI  
 nlaIV  
 hgiCI  
 bani  
 tru9I  
 mseI  
 maeIII  
 bphI  
 rmaI  
 sau3AI  
 mboI/ndeII(dam-)  
 dpnI(dam+)  
 dpnII(dam-)  
 tru9I  
 ahaIII/draI maeI  
 tru9I  
 mseI  
 bstVI/xhoII  
 alwI(dam-)  
 TCCACGGAGT GACTAATTTCG TAACCATTGA CAGTCTGGTT CAAATGAGTA TATATGAAT CTAACATAAT TTGAAGTAA AATTAATTT TTCTAGATC

**FIG. 10R**

[illegible]

## FIG. 10S

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6701 CGCCAGCCTT CCGAAGGGA GAAAGGCGG CAGGTATCCG GTAAGCGGCA GGGTCGGAC AGGAGAGCGC hhaI/cfoI aluI apyI(dcm+) apyI(dcm+)
GGGCTGCGAA GGGCTTCCCT CTTTCCGCT GTCCATAGGC CATTCGCCGT CCCAGCCTTG TCCTCTCGCG TGCTCCCTCG AAGTCCCCC TTTCGGGACC
scrFI mvaI scrFI
mvaI ecorII mvaI
dsav ecorII
bstNI dsav
bsaJI bstNI
6801 TATCTTTATA GTCTGTGCG GTTTCGCCAC CTCGTGACTTG AGCTCGATT TTGTGATGC TCGTCAGGGG GCGGAGCCT ATGGAAAAC GCCAGCAACG
ATAGAAATAT CAGGACAGCC CAAAGCGGTG GAGACTGAAC TCGCAGCTAA AACACTAGC AGCAGTCCCQ CCGCTCTGGA TACCTTTTG CCGTCGTGC
fnu4HI
aciI
thaI
fnuDII/mvni
bstUI
bsh1236I
nlaIV
aciI
sfanI
taqI
hgaI
mnlI drdI
6901 CGGCTTTTAT ACAGTTTCTG GCCTTTTCTG GCCTTTTCTG TCACATGTC TTYCTGCGT TATCCCTGA TTCTGTGGAT AACCGTATTA CCGCCTTTGA
GCCGGAANA TGCCAAGGAC CGGAAAACGA CCGTACAAAG AGTGTACAAG AAAGGACGCA ATAGGGGACT AAGACACCTA TTGCATAAT GCGGGAAC
bsaI
haeIII/palI nlaIV
scrFI
mvaI
ecorII
dsav
bstNI bsaI
apyI(dcm+)
haeIII/palI nspHI
haeIII/palI nlaIV haeI haeI afiIII
tfii
hinfi
aciI

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**FIG. 10T**

FIG. 10T

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[illegible]

	scrFI	mvaI	ecorII	dseV	bstNI	apyI dcm+	mspI	hpaII	bsaBI	aciI	bsrBI	alul	nlaiII	xmnI	asp700
7201	CACCCACAGGC	TTTACACTTT	ATGCTTCCGG	CTCGTATGTT	GTGTGGAATT	GTGAGCGGAT	AACAATTTCA	CACAGGAAAC	AGCTATGACC	ATGATTACGA					
	GTGGGGTCCG	AAATGTGAAA	TACGAAGGCC	GAGCATACAA	CACACCTTAA	CACCTGCCTA	TTGTTAAGT	GTGTCCCTTG	TCGATACTGG	TACTAATGCT					

```

tru9I
mseI
aseI/asnI/vspI
7301 ATTAA
TAATT

```

```
>length: 7305
```



## INTERNATIONAL SEARCH REPORT

International Application No  
PCT/US 95/09576

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/64 C12N15/67 C12N15/85 C12N9/72 C12N5/10

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DNA CLONING, VOLUME III, EDITED BY D.M. GLOVER, 1987 IRL PRESS, OXFORD, GB;, pages 189-212, A.M.C. BROWN AND M.R.D. SCOTT 'Retroviral vectors'	1-3, 7, 8
Y	see page 192, line 7 - page 196, line 5; figures 2,3  --- -/--	5, 6, 9-12, 16-21

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

## \* Special categories of cited documents:

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- \* "&" document member of the same patent family

Date of the actual completion of the international search

23 November 1995

Date of mailing of the international search report

08.12.95

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Hornig, H

## INTERNATIONAL SEARCH REPORT

Int. onal Application No

PCT/US 95/09576

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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X	CELL, vol. 37, no. 3, July 1984 CELL PRESS, CAMBRIDGE, MA, US;, pages 1053-1062, C.L. CEPKO ET AL. 'Construction and applications of a highly transmissible murine retrovirus shuttle vector' cited in the application	1-3,7,8
Y	pZIP-Neo SV(B)1 see figure 1	5,6, 9-12, 16-21
Y	--- MOL. CELL. BIOL., vol. 5, no. 3, March 1985 ASM WASHINGTON, DC, US, pages 431-437, A.D. MILLER ET AL. 'Generation of helper-free amphotrophic retroviruses that transduce a dominant-acting, methotrexate-resistant dihydrofolate reductase gene' see page 432, right column, line 5 - page 436, right column, line 7; figure 1	5,6, 9-12, 16-21
Y	WO,A,94 05784 (US) 17 March 1994  see the whole document	5,6, 9-12, 16-21
Y	--- EP,A,0 215 548 (ZYMOGENETICS INC ;UNIV WASHINGTON (US)) 25 March 1987  see the whole document	5,6, 9-12, 16-21
A	--- WO,A,92 17566 (GENENTECH INC) 15 October 1992 cited in the application see the whole document	1-21
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Int ional Application No

PCT/US 95/09576

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>PROC. NATL.ACAD SCI.,  vol. 86, February 1989 NATL. ACAD  SCI.,WASHINGTON,DC,US;,  pages 1041-1045,  M. VIVAUD ET AL. 'A 5' splice-region G-C  mutation in exon 1 of the human  beta-globin gene inhibits pre-mRNA  splicing: A mechanism for  beta+-thalassemia'  see the whole document  -----</p>	1-4

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Information on patent family members

International Application No

PCT/US 95/09576

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		DE-A- 3730599	07-07-88
		FR-A- 2603899	18-03-88
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		AU-B- 4134585	24-10-85
		AU-B- 5295890	30-08-90
		EP-A- 0385558	05-09-90
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		JP-A- 60243023	03-12-85
		JP-A- 6040942	15-02-94
		NO-B- 174934	25-04-94
		SG-A- 3994	10-06-94
		US-A- 4965199	23-10-90